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Comparative pharmacognosy atlas of *Pum Kutaja* (*Holarrhena antidysenterica* Wall. Ex A. Dc.) And *Stree* *Kutaja* (*Wrightia tinctoria* (Roxb.) R. Br.)

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Abstract

Kutaja is commonly used for amoebic dysentery, piles and skin diseases. Male and female varieties of *Kutaja* are mentioned in Ayurvedic classics which are sourced from *Holarrhena antidysenterica* Wall. ex A. DC. and *Wrightia tinctoria* (Roxb.) R. Br. respectively. Present study was undertaken to have a brief outlook of similarities and dissimilarities between the barks of both the plants. The samples were subjected to macro- and microscopic analysis, powder microscopy, physicochemical, phytochemical and HPTLC evaluation. Pharmacognostic analysis revealed both sources had crystals of calcium oxalate, stone cells and sclereids in common. However, phloem fibres were present only in *W. tinctoria*. Whereas, pericyclic fibres and starch grains were present in *H. antidysenterica*. Powder microscopy of *H. antidysenterica* showed parenchyma with starch grains and sclereids whereas, thin walled septate fibres and cut latex vessels were seen in *W. tinctoria*. HPTLC revealed similar R_f values in both the samples.

Keywords: Adulteration, Comparative pharmacognosy, *Kutaja*, Phytochemistry.

Introduction

Kutaja is one among the most commonly used medicinal plants and having two varieties such as *Pum* (male) and *Stree* (female) [1]. It is known for its Sangrahi (antidiarrheal) and Upashoshana (drying up) [2] properties, which makes it the drug of choice in *Atisara* (diarrhea). The two source plants considered for *Pum Kutaja* and *Stree Kutaja* are *Holarrhena antidysenterica* Wall. ex A. DC. and *Wrightia tinctoria* (Roxb.) R. Br. respectively [3], both belonging to family Apocynaceae. The bark of *H. antidysenterica* is considered as astringent, anthelmintic, amoebicidal and diuretic [4], and is used in amoebiasis [5], colic, dyspepsia, piles, diseases of the skin and spleen [4]. The bark of *W. tinctoria* is also known to be antidiarrheal [6], and is used in treatment of piles and skin disorders [7]. It is also known that the bark of *W. tinctoria* is commonly used as an adulterant of *Kurchi Bark* (*H. antidysenterica*) [7]. Moreover, studies like pharmacognosy, antibacterial activity and physicochemistry on seeds of these two species have been studied earlier [8]. This study however, is designed to evaluate the comparative account of pharmacognostic and phytochemical components of bark of both species.

Materials and Methods

Fresh plant materials were collected from Udupi District of Karnataka, authenticated by referring the regional floras [9] and voucher specimens (14031401-02) were deposited at the Pharmacognosy department of S.D.M. Centre for Research in Ayurveda and Allied Sciences, Kuthpady, Udupi. Plant samples were washed and shade dried at room temperature for 10 days. The dried plant material was pulverized into a fine powder using a pulverizer.

Macro-microscopic study

Systematic recording of macroscopic characters of barks of both plants were recorded using Canon IXUS digital camera according to the text book of Pharmacognosy [10]. Samples were preserved in FAA (Formalin - 5ml + Acetic acid - 5ml + 70% Ethyl alcohol - 90ml) fixative solution for more than 48 hours. The preserved specimens were cut into thin transverse section using a sharp blade, followed by staining with saffranine [10]. A pinch of the sample was mounted on a microscopic slide with a drop of glycerin-water. Magnifications of the figures were indicated by the pre-calibrated scale-bars using Zeiss Axio Vision software.

In order to supplement the descriptive part, photomicrographs in different magnifications of all necessary cells and tissues were taken in Zeiss Axio Lab trinocular microscope and Zeiss Stemi stereo microscope. Magnifications of the figures are indicated by the pre-calibrated scale-bars using Zeiss Axio Vision software.

Physicochemical evaluation

The percentage of physicochemical values like loss on drying, total ash, acid insoluble ash, alcohol soluble extractive, water soluble extractive were carried out as per the methods described in the Indian Pharmacopoeia [11].

Phytochemical study

Powdered material was extracted in ethanol using cold maceration and was tested for various classes of active chemical constituents, using standard prescribed methods as described in Trease and Evans [10].

High Performance Thin Layer Chromatography (HPTLC)

One gram of each powdered samples was extracted with 10 ml ethanol and kept for cold percolation for 24h and then filtered. Three and 6 μ l of the above samples were applied on a pre-coated silica gel F254 on aluminum plates to a band width of 7 mm using Linomat 5 TLC applicator. The plate was developed in toluene: ethyl acetate (9.0:1.0). The developed plates were visualized under UV 254, 366 nm and then scanned under UV 254 and 366 nm. The plate was derivatised with vanillin sulphuric acid reagent to see constituents not detected under UV. R_f , colour of the spots and densitometric scan were recorded [12].

Results

Holarrhena antidysenterica Wall. ex A. DC. dried stem bark appear in small recurved pieces that are 3 to 5 cm long and 0.5 to 2 cm thick, outer surface dark brownish, longitudinally wrinkled and bearing horizontal lenticels, inner surface brownish and rough [Fig. 1.1].

Fig 1: Macroscopy of *H. antidysenterica* and *W. tinctoria*



Fig. 1.1. *H. antidysenterica*

Outer surface of *Wrightia tinctoria* R. Br. dried stem bark, faintly longitudinally and transversely striated and intermittently displays small circular lenticels. Inner surface rough, having fibres and is buff in colour. [Fig. 1.2].



Fig 1.2: *W. tinctoria*

Transverse section (TS) of stem bark of *H. antidysenterica* shows periderm, a wide stratified cortex and secondary phloem. Periderm consists of thin-walled and somewhat rectangular cork cells and 2 layers of phellogen. Phelloderm parenchymatous, containing prisms of calcium oxalate crystals and a few starch grains. Cortex shows groups of lignified pitted stone cells of different shapes upto 0.3 mm in size. In phloem region prisms, sclereids and stone cells can be seen. Medullary rays bi- or tri-seriate in different regions. Phloem parenchyma shows prisms of calcium oxalate crystals and starch in abundance. Phloem fibres absent. [Fig. 2].

Fig 2: Microscopy of bark of *Holarrhena antidysenterica*

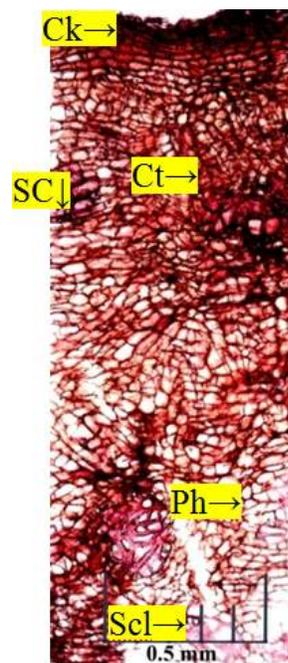


Fig 2.1: TS of entire bark

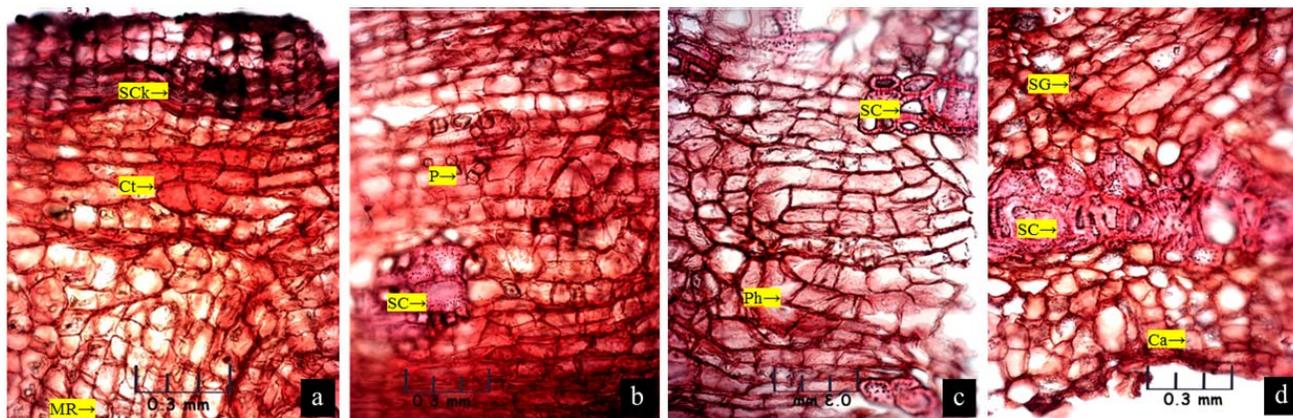


Fig 2.2 a: Stratified cork, cortex; b. phloem region with prisms, stone cells; c. sclereids group, d. starch grains and cambium

TS of stem bark of *W. tinctoria* shows outermost multilayered cork, and parenchymatous cortex. Cortex interspersed with stone cells; phloem region shows fibers, medullary rays and laticiferous tubes and occasional isolated stone cells. Cortex made up of 12 to 15 layers traversed with usually isolated

prismatic crystals of calcium oxalate. Sieve tubes can be seen. Phloem wide, traversed with uni and sometimes multi-seriate medullary rays, isolated lignified fibres, and oval to circular wide latex canal. [Fig. 3].

Fig 3: Microscopy of bark of *Wrightia tinctoria*

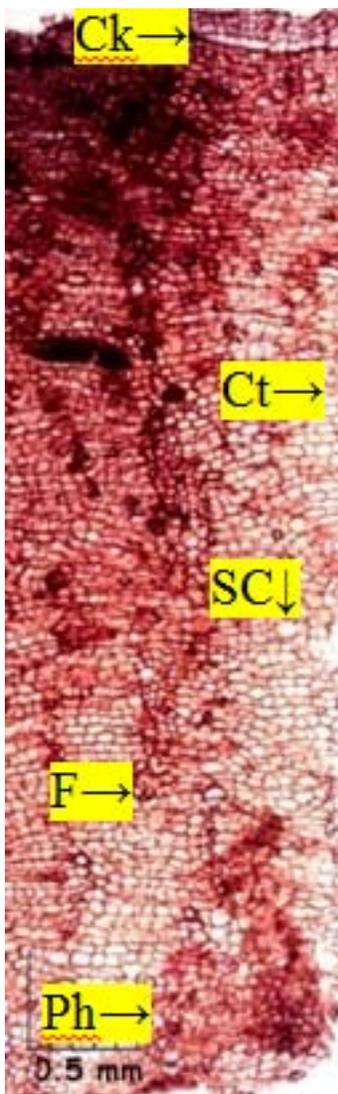


Fig 3.1: TS of entire bark

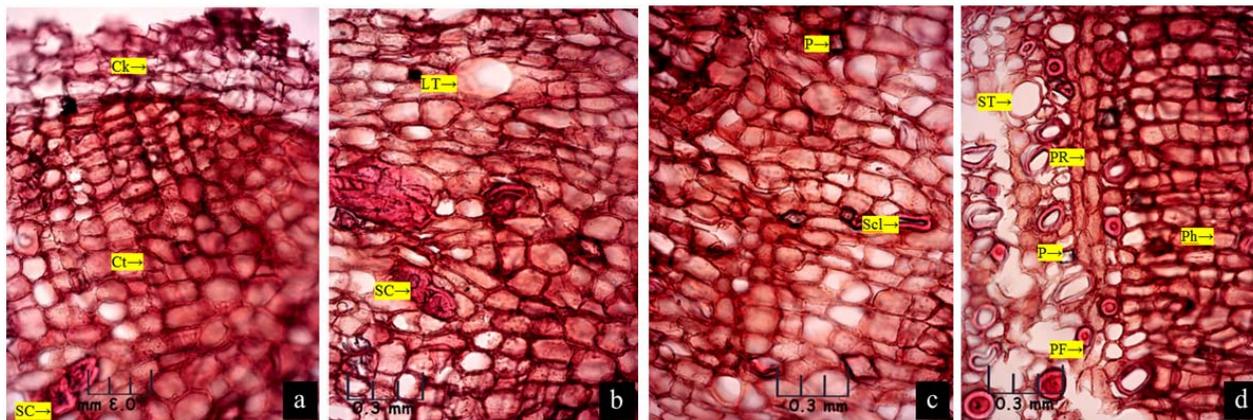


Fig 3.2 a: Cork and cortex; b. cortex with latex tubes and stone cells; c. phloem region with fibres and prisms; d. phloem rays, fibres and prisms

Powder of *H. antidysenterica* is light brown, taste bitter; parenchyma, thick walled with contents and starch grains, transversely and obliquely cut thin walled cork cells, groups of phloem tissue and few sclereids visible. [Fig. 4].
 stone cells having different sizes and shapes. Cortical

Fig 4: Powder microscopy of *Holarrhena antidysenterica*

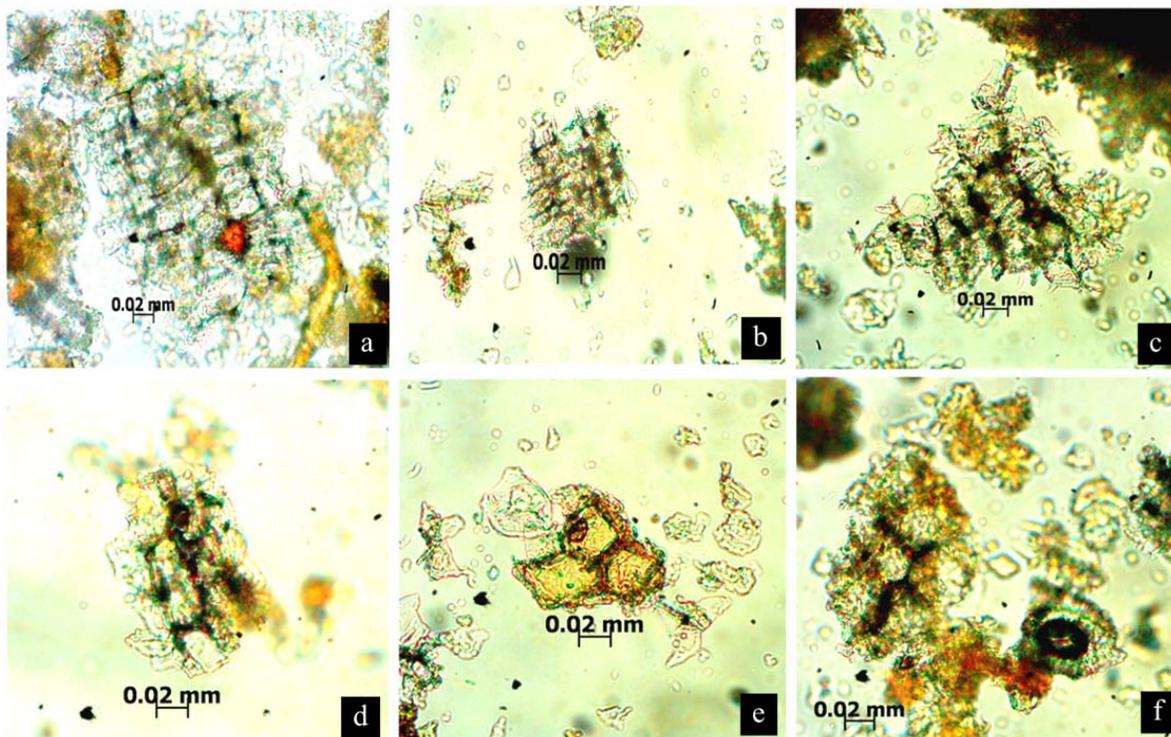
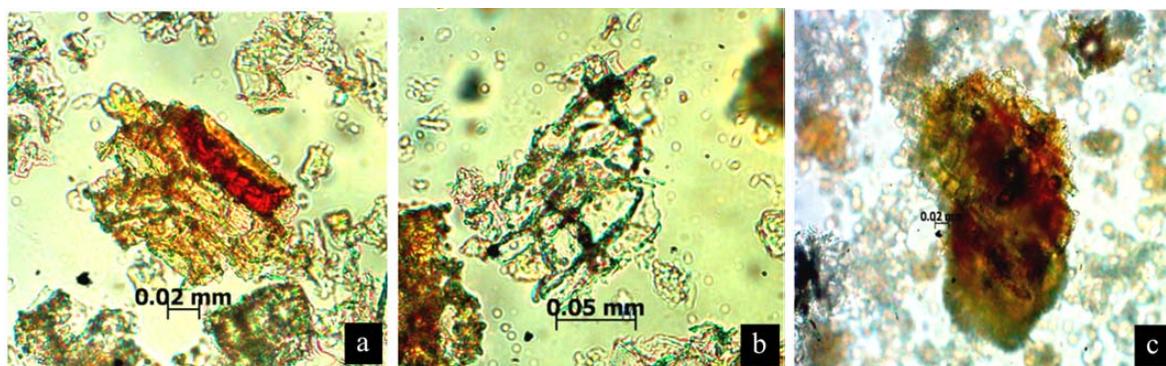


Fig 4.1 a: Transversely cut cork; b. Cork cells; c. Obliquely cut cork cells; d. Cortical parenchyma; e. Parenchyma with content; f. Stone cell



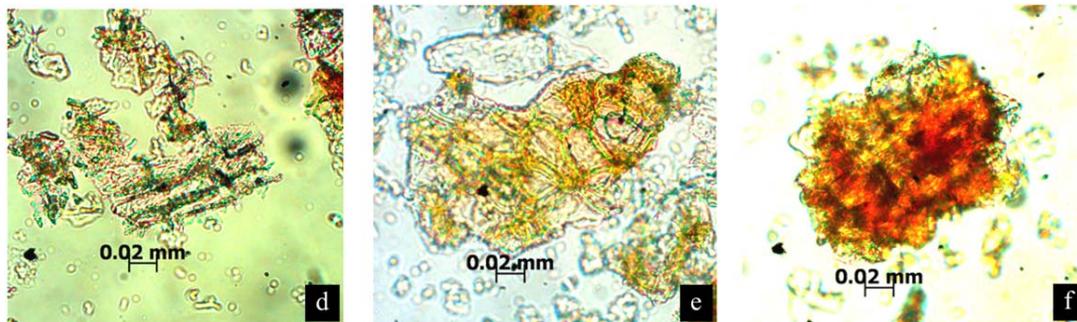


Fig 4.2 a: Parenchyma with content; b. Parenchyma with starch grains; c. Stone cell group; d. Phloem tissue; e. Thick walled cortical parenchyma; f. Sclereids

W. tinctoria powder shows presence of the fragments of quadrangular, pentagonal and hexagonal cork cells in surface view and sectional view, chlorenchyma of cortex and prisms of calcium oxalate; parenchymatous cells with content,

fragments of thin walled septate fibres; fibers with pitted walls; longitudinally cut latex vessels filled with granular contents. [Fig.5].

Fig 5: Powder microscopy of *Wrightia tinctoria*

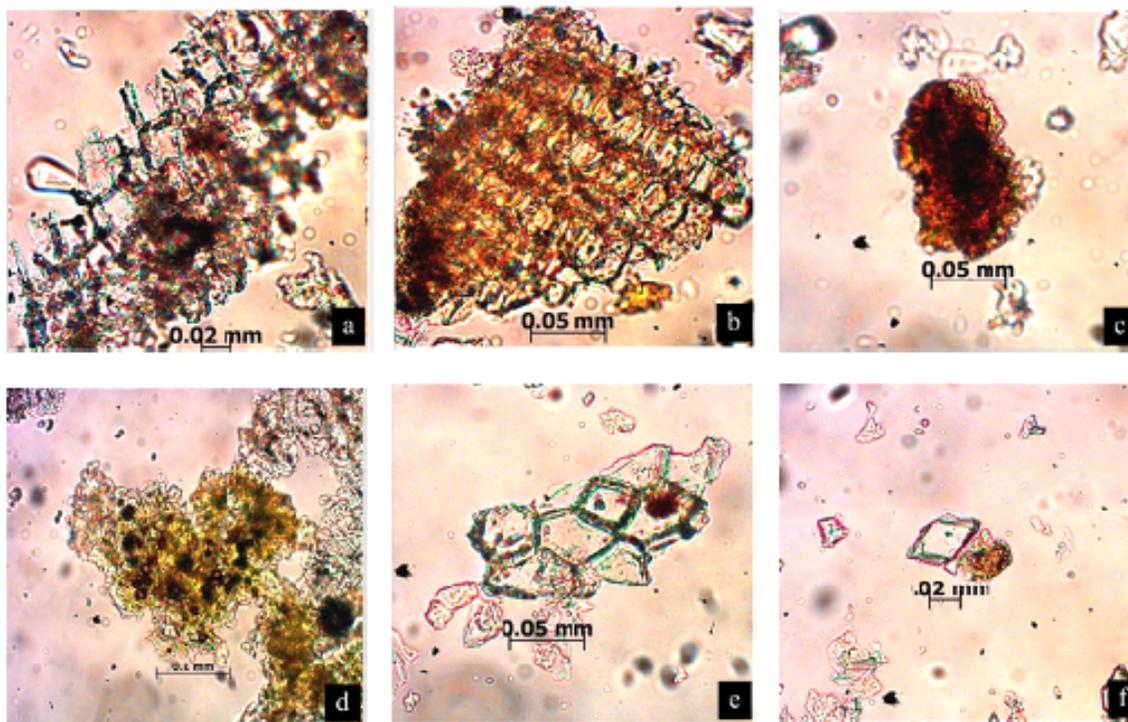


Fig. 5.1 a: Obliquely cut cork cells; b. Transversely cut cork cells; c. Parenchyma with content; d. Chlorenchyma of cortex; e. Cork cells in surface view; f. Prisms of calcium oxalate

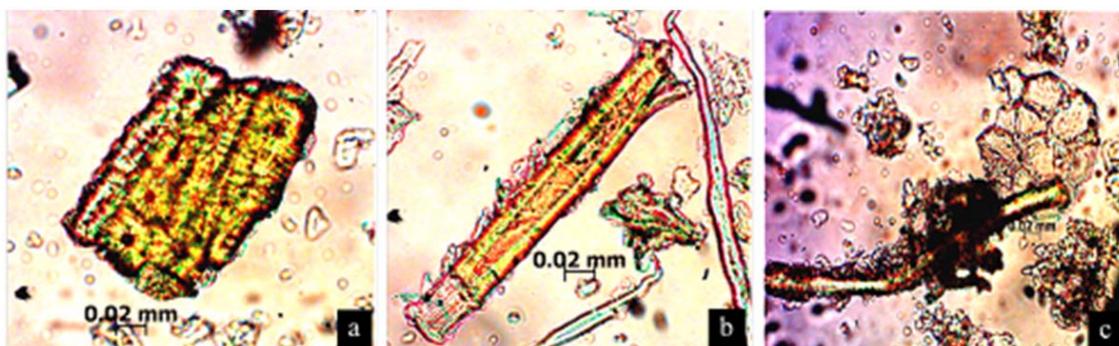




Fig 5.2 a: Sclereid group; b. Fragment of fibre; c. Fragment of fibre; d. Pitted fibres; e. Latex vessel; f. Pitted fibres

Comparative pharmacognostic characters

Many similar and dissimilar characters were observed between the barks as such and barks in powdered form of *H. antidysenterica* and *W. tinctoria*. The dissimilarities are as: the inner surface of *H. antidysenterica* bark is brown in colour and rough whereas *W. tinctoria* bark has pale brown and smoother inner surface. The stone cells are arranged throughout the section, and are arranged in concentric tangential bands in *H. antidysenterica* and are present only in cortical region in case of *W. tinctoria*. Prisms of calcium oxalate are comparatively less in *W. tinctoria* than in *H. antidysenterica*. Phloem fibres are present in *W. tinctoria* and absent in *H. antidysenterica*. Whereas pericyclic fibres are absent in *W. tinctoria* and present in *H. antidysenterica*.

Physico-chemical and phytochemical results

Loss on drying at 105° C was found to be almost same in both the samples. Total ash content was found to be more in *W. tinctoria* (12.86) than in *H. antidysenterica* (7.05). Similar results were observed in case of acid insoluble ash with values of 0.49 and 0.19 respectively. Water soluble extractive was more in *H. antidysenterica* (27.04) than in *W. tinctoria* (24.70). Alcohol soluble extractive did not show much variation, though. [Table-1]

Table 1: Physico-chemical parameters of *H. antidysenterica* and *W. tinctoria*

Parameter	Results n=3 %w/w	
	<i>H. antidysenterica</i>	<i>W. tinctoria</i>
Loss On Drying	10.42	10.48
Total ash	7.05	12.86
Acid Insoluble Ash	0.19	0.49
Alcohol Soluble Extractive	10.25	10.46
Water Soluble Extractive	27.04	24.70

Alkaloids, carbohydrates, resins, steroids, saponins, tannins and terpenoids were present in both the species. The presence of coumarins, flavonoids and phenol was seen only in *H. antidysenterica*. [Table-2]

Table 2: Results of preliminary phytochemical tests of *H. antidysenterica* and *W. tinctoria*

Test	<i>H. antidysenterica</i>	<i>W. tinctoria</i>
Alkaloid	+	+
Carbohydrate	+	+
Carboxylic acid	-	-
Coumarins	+	-
Flavonoids	+	-
Phenol	+	-
Quinone	-	-
Resins	+	+
Steroid	+	+
Saponins	+	+
Tannin	+	+
Terpenoid	+	+

Three bands were seen under *H. antidysenterica* with R_f values of 0.08, 0.48 and 0.83 under 366nm. Whereas in *W. tinctoria* only two bands were seen with 0.08 and 0.03 R_f . Seven bands were seen post derivatisation under *H. antidysenterica* with R_f values 0.07, 0.13, 0.16, 0.30, 0.34, 0.51 and 0.80. While eight bands were observed under *W. tinctoria* with R_f values 0.15, 0.21, 0.35, 0.46, 0.52, 0.61, 0.67 and 0.90. [Table-3] On spectral comparison between both, the R_f values overlapping in both the samples under short and long UV radiations are 0.01, 0.73, 0.82 and 0.01 respectively. These graphs show the overlapping zones present in *H. antidysenterica* and *W. tinctoria*. [Fig. 6-9]

Table 3: R_f value of Alcohol extract of the Samples of *Holarrhena antidysenterica* and *Wrightia tinctoria*

At 254nm		At 366nm		Post Derivatisation	
HA	WT	HA	WT	HA	WT
-	-	0.08 (DF. Blue)	0.08 (LF. Blue)	0.07 (L. Purple)	-
-	-	0.48 (LF. Blue)	-	0.13 (L. Purple)	-
-	-	0.83 (DF. Blue)	0.83 (DF. Blue)	0.16 (L. Purple)	0.15(L. Purple)
-	-	-	-	-	0.21(L. Purple)
-	-	-	-	0.30 (D. Purple)	-
-	-	-	-	0.34 (D. Purple)	0.35 (L. Purple)
-	-	-	-	-	0.46 (D. Purple)
-	-	-	-	0.51 (D. Purple)	0.52 (D. Purple)
-	-	-	-	-	0.61 (L. Purple)
-	-	-	-	-	0.67 (L. Purple)
-	-	-	-	0.80 (D. Purple)	-
-	-	-	-	-	0.90 (D. Purple)

*L-Light, D-Dark, F-Fluorescent

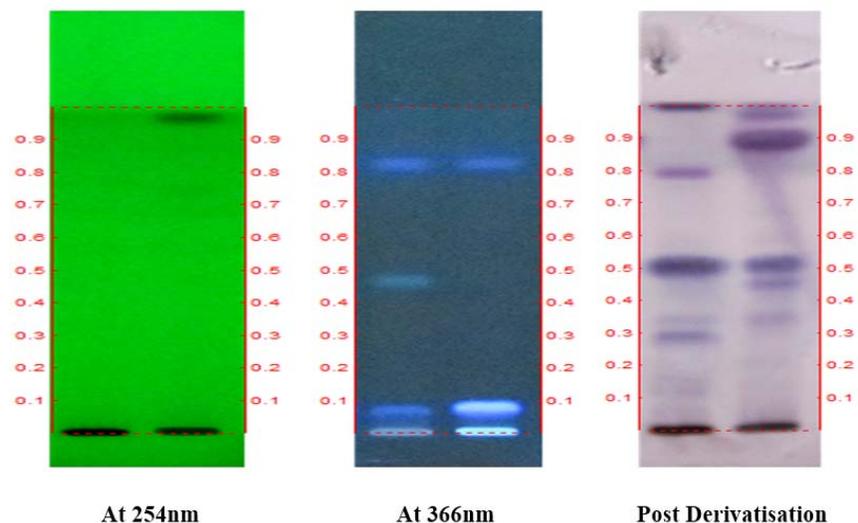


Fig 6: HPTLC photodocumentation of Alcohol extract of *Holarrhena antidysenterica* and *Wrightia tinctoria*

Track 1- Alcohol extract of *Holarrhena antidysenterica* 6µl

Track 2 – Alcohol extract of *Wrightia tinctoria* 6µl

Solvent system – Toluene: Ethyl acetate: (9:1)

Fig 7: HPTLC Densitometric Scan at 254 nm

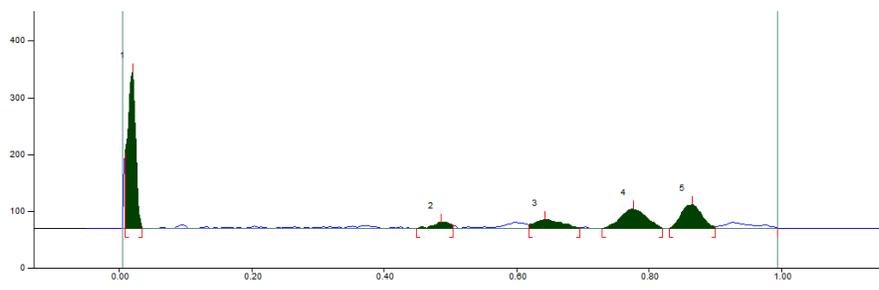


Fig 7a: *Holarrhena antidysenterica*

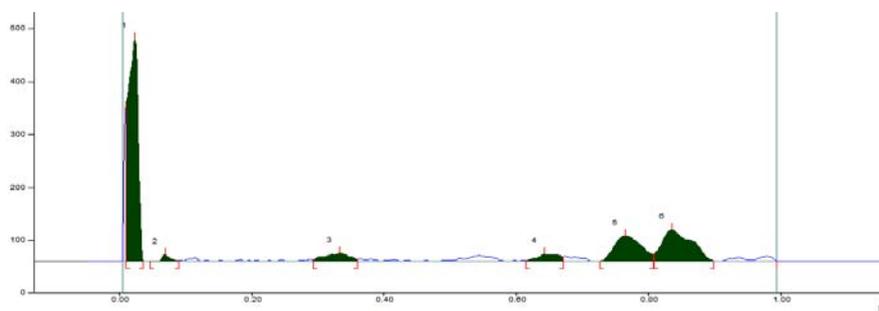


Fig 7b: *Wrightia tinctoria*

Fig 8: HPTLC Densitometric Scan at 366nm

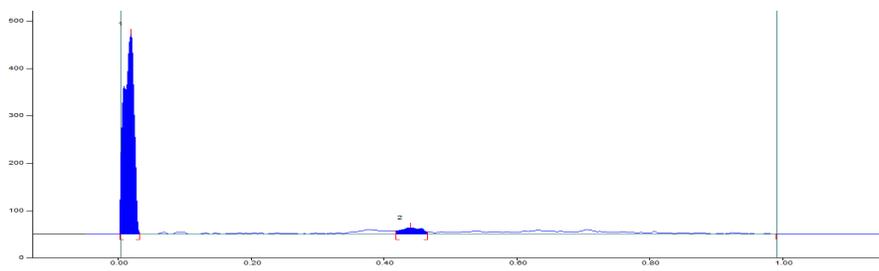


Fig 8a: *Holarrhena antidysenterica*

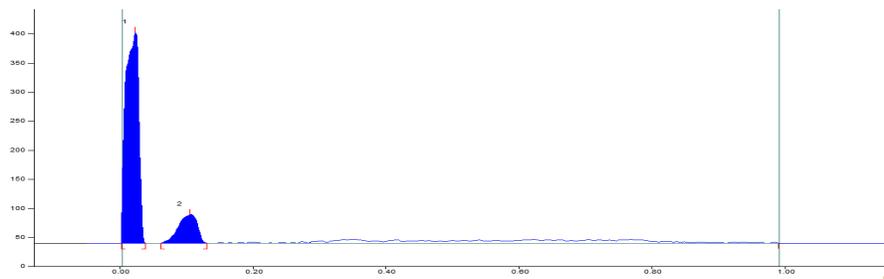
Fig 8b: *Wrightia tinctoria*

Fig 9: 3-D Chromatogram

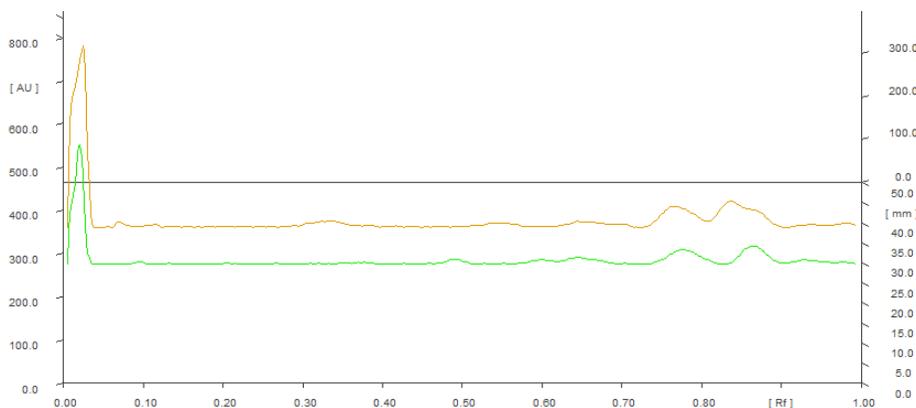


Fig 9a: 3-D Chromatogram at 254 nm

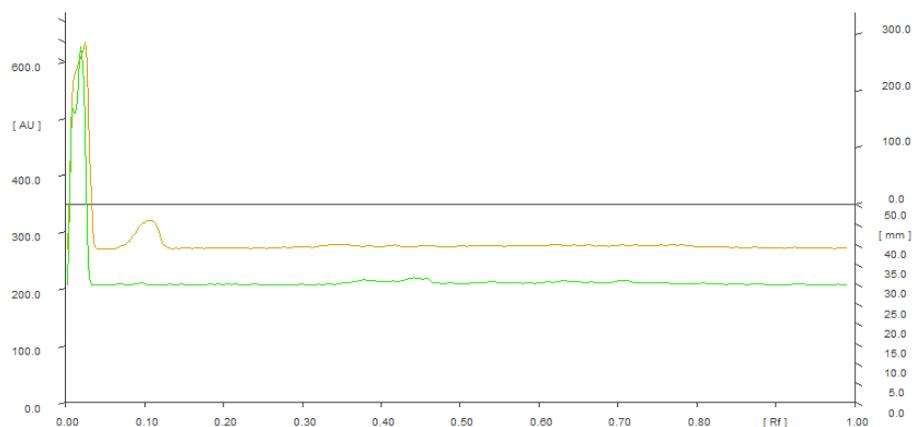


Fig 9b: 3-D Chromatogram at 366nm

Discussion

Habit and habitat of both plants are similar as both of them belong to same family. Recurved shape and dark brownish colour of *H. antidysenterica* bark plays an important role in differentiating between both. The crystals of calcium oxalate are present in both the samples and have prismatic form. Stone cells/sclereids commonly occur in hard outer coats of seed and fruits and in the bark. In *H. antidysenterica* lignified pitted stone cells can be seen. Fibres are usually differentiated on the basis of the tissue in which they occur. Pericyclic fibers are seen only in *H. antidysenterica* bark which differentiates it from *W. tinctoria* bark. Fibres are absent in phloem of *H. antidysenterica* but presence of isolated lignified fibres in the phloem of *W. tinctoria* is a distinguishing character. Starch occurs in the form of granules (commonly known as starch grains). Their shape and size are characteristics of the species. They are found in the bark of *H. antidysenterica* and are absent in the other sample. More total ash value in *W. tinctoria*

indicates presence of more amounts of extraneous matter in it. Comparatively presence of more silicaceous matter in *W. tinctoria* may be considered as a reason for more acid-insoluble ash value in it. The existence of close relationship between constituents of plants and their taxonomical status is established by extensive phytochemical screening. Such similarity can be seen in both these samples under study, as both of them belong to Apocynaceae family. Still *H. antidysenterica* showed presence of coumarins, flavonoids and phenol, which separates it from *W. tinctoria*. Overlapping zones in HPTLC indicates similarity in chemical components present in both samples. However they have to be evaluated pharmacologically and clinically.

Conclusion

W. tinctoria which is commonly used as an adulterant of *H. antidysenterica* bark, shows similar morphology and habitat. Pharmacognostic evidences show the presence of calcium

oxalate crystals, stone cells and sclereids as key characters for differentiation. Phytochemically they are found to be similar with R_f value 0.01 and 0.73 in common when compared spectrally. This study is useful in the identification of both the species as well as in their differentiation from each other. Further scientific evaluation at molecular level, marker compounds and pharmacological confirmation is required however.

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