Pharmacological activity investigation of *Callicarpa attenuate*

Md. Rafayet Hossain, Mahmud Hasan Rumi, Farhan Hasin Arik, Tahmid Khurshed and Amina Ferdous Chowdhury

**Abstract**

*Callicarpa attenuate* plant extract used to assess its different biological activity. Extract was made by soaking the dried plant powder in methanol. After comparing with the standard we found that Methanol extract of the sample gave the activity against all the experimented microbes of ZI (zone of inhibition) against *E. coli* and *B. subtilis*. After performing the antioxidant, thrombolytic, anti diabetic, hypoglycemic and cytoxic activity assay of methanol extract of sample plant we saw that it has a good biological activity that can be used as a potential traditional medicine.

**Keywords:** *Callicarpa attenuate*, antioxidant, antimicrobial, anti diabetic, hypoglycemic, thrombolytic activity

**Introduction**

Plants are playing an intrinsic role of all living organisms of planet and earth by proving different features like medicines, food, clean air, clothing, shade and shelter [9]. It is unknown to us when and where people first used plants for treating different diseases, but the relation between health and plants has existed for many years [2]. Medicinal plants can be recognized by depending on their different therapeutic properties and through trial and error method. According to World Health Organization (WHO), 25% of prescribed medicines are come from natural sources which were used in traditional method in ancient era [9]. In modern era, medicinal herbs are being used tremendously and accepted in urban areas also due to their effectiveness, availability, low toxicity and affordability. Moreover, medicinal plants have greater acceptance because many synthetic drugs that are used for treating many diseases like gonorrhea, diabetes, tuberculosis and so on are showing inefficiency along with increase the drug resistance of different antibiotics and the cost of prescribed drug are at higher price [6]. Medicinal plants are full of different bio-resource of traditional medicine, modern medicine, nutraceuticals, folk medicine pharmaceutical intermediates and chemical entities for synthetic drugs [7]. There are different effective ways in which primary and secondary metabolites from the plants sources can be discovered, identified and isolated for studying further in medicinal plants [4]. The primary metabolites include carbohydrates, amino acid, proteins, chlorophyll where secondary metabolites include alkaloids, saponins, flavonoids etc. These metabolites mainly protect human health from different diseases [9]. *Callicarpa attenuata* is belonging to Verbenaceae family. This plant is widely distributed in many countries including Hainan, Sichuan, Guangdong, Guangxi, Yunnan India, Taiwan, Indonesia, Laos, Philippines, Singapore, Malaysia, Myanmar, Thailand, and Vietnam [3]. The shrub about 5-15 ft. tall, stem color is light gray, much branched slender, flowers numerous, bisexual, actinomorphic [1]. The genus *Callicarpa* has the presence of flavonoids, essential oils, and terpenoids, fatty acids, flavones, lignans, monoterpenes, phenylpropanoids, phytosterols, sesquiterpenes, and triterpenes [5]. It has also various important medicinal uses such as relieve rheumatic pain, treat diarrhea and dysentery, prevent rashes, ease digestion, bark of the plant is used to apply on cuts and wounds etc. [3]. This study is aimed at providing information about the properties of plant like antioxidant, anti diabetic, cytotoxic, analgesic and hypoglycemic activity by using plant extraction of *Callicarpa attenuate*.

**Methods and Materials**

**Collection of plant materials:**

The leaf part of *Callicarpa attenuate* plant was collected in May, 2017 from Chittagong hill tract. After collection, the National Herbarium Bangladesh (NHB), Mirpur, and Dhaka authenticated the plant material and provided a plant identification number, which was 47696.
Preparation of the extract
At first, the leaves part was washed with fresh water to remove the unwanted dust particles and plant scrap. After that, the cleaned leaves were dried under the sun for a day. Then the leaves were again dried for 1 hour at 30-40 °C in hot air oven. By using a high capacity grinding machine, the dry and crusty leaves were ground. After that, at a normal ambient temperature (22-25 °C) around 900 g of ground powder was soaked in 2.5 L of methanol for a period of 2 days with occasional stirring. With the help of cotton filter (pore size: 110nm) filtration was done and rotary evaporator was used at 100 rpm at 30 °C to evaporate the maximum amount of solvent. For vaporizing the solvent completely from the extract, the leaf extract was kept under laminar airflow cabinet. Moreover, it was used to avoid any possibility of microbial growth in the extract while drying. Finally, 22.4 g of plant leaf extract was obtained and kept in dry and cool place and proper labeling was done. After that, this extract was used to conduct antioxidant, brine shrimp lethality assay, thrombolytic, anti-diabetic, antimicrobial and hypoglycemic studies.

Chemicals
The chemicals were gallic acid [Sigma-Aldrich, USA], sodium chloride [Sigma-Aldrich, USA], Folin-Ciocalteu reagent [Sigma-Aldrich, USA], vincristine sulphate [Sigma-Aldrich, USA], 2,2-Diphenyl-1-Picrylhydrazyl (DPPH) [Sigma-Aldrich, USA], sodium carbonate [Merck, India] and ascorbic acid (ASA) [Merck, India], dimethyl sulfoxide (DMSO) [Fisher Scientific, UK], Castor oil (WELL’s Heath Care, Spain), 0.9% sodium chloride solution (normal saline) (Orion Infusions Ltd., Bangladesh), charcoal meal (10% activated charcoal in 5% gum acacia), and loperamide (Square Pharmaceuticals Ltd., Bangladesh) were used for anti-diarrheal activity test, and dimethyl sulfoxide (DMSO) (Sigma-Aldrich, USA) and sodium chloride (Sigma) were used for cytotoxic activity test. All the chemicals used in this study were of analytical grade.

Anti-oxidant activity
Total phenolic content (TPC)
The phenols were oxidized by Folin-Ciocalteu in ionic phenolic solution. When the solution became yellow to dark blue, it is understood that the oxidation has been completed. After that, this color changed mixture measured in 760 nm in UV spectrophotometer. Finally, the value of the absorbance plotted in gallic acid calibration curve and data was evaluated as gallic acid equivalents (GAE).

Total Flavonoid content
Aluminum chloride was used to determine the total amount of flavonoids. Firstly, 0.5 ml of plant extract was made the final volume of 1 ml for reaction medium (MeOH/HO/CH,COOH=14:5:1) which was then mixed with Aluminum chloride reagent (4 ml, 133 mg of AlCl₃ × 6 H₂O and 400 mg of CH,COONa dissolved in 100 ml H₂O). After 5 minute, the absorbance was measured at 430 nm. Based on the calibration curve, total flavonoid content was calculated and it was expressed as gram equivalents.

DPPH free radical scavenging assay
The antioxidative activity of *Callicarpa attenuate* was determined by performing DPPH free radical scavenging assay. To run this assay, different concentrations of plant extracts were mixed with 2, 2-diphenyl-1-picrylhydrazyl (DPPH) solution. In methanol or aqueous solution, free radicals were generated due to delocalization of the free electrons and a deep purple colored solution is produced. Then absorbance of different concentration solutions was measured at 517 nm in UV spectrophotometer. The decreasing value of DPPH at 517 nm is directly proportional to the radical scavenging activity. Percentage of inhibition of DPPH free radical (1%) was calculated by using the following equation:

\[
(1%) = \frac{\text{Absorbance of blank} - \text{Absorbance of sample}}{\text{Absorbance of blank}} \times 100
\]

50% of inhibition of the extract concentration was calculated from the graph and the percentage of inhibition was plotted against extract concentration.

Cytotoxic activity
Brine shrimp lethality assay
In this assay, *Artemia salina* shrimp was used. Its offspring was hatched in replicated seawater to cultivate nauplii. Here, calculated amount of dimethyl-sulfoxide (DMSO) was added with sample and desired concentration of sample was prepared by dilution. The counted nauplii were placed in vials that contained approximately 5 mL simulated seawater with visual inspection. With the help of micropipette, various concentrations of samples were added to tubes. Here, vincristine sulfate was used as standard. The sample containing tubes were then placed in a dry place for 24 hours at room temperature. At the last, after 24 hours, the survived nauplii were counted. Percentage (%) of mortality was calculated by using the following equation:

\[
\text{Percentage of mortality= (Number of nauplii taken - Number of nauplii alive)/ Number of nauplii taken} \times 100
\]

50% of lethal concentration of extract concentration was calculated from the graph plotted percentage of mortality against concentration.

Thrombolytic activity
The normal blood flow to the cells and tissues can be hampered due to thrombus as it blocks the blood vessel which can lead to lack of blood and oxygen. There are some thrombolytic medications like urotokinase, clopifogrel, and streptokinase remove this thrombus and cells and tissues are remained in normal conditions. For this assay, fresh human blood was collected. Then, they were taken in three different pre-weighted sterile microbes and incubated for 45 minutes at 37 °C. The upper fluid was entirely dispensed from all microtube lines when the clot was appeared. As a standard streptokinase was used and as a negative control distilled, water was used. 100 microliter of plant extract was taken in each tube and incubated for 90 minutes at 37 °C. Next, liquid that was released from the clot was removed and the tubes were weighted again to observe the weight difference when the clot disruption occurred. Percentage of clot lysis was calculated by following equation:

\[
(\%) = \frac{\text{released clot weighted}}{\text{clot weight after release}}
\]

Antimicrobial assay
Disc Diffusion Assay Method
In recent years, different studies are developing as antimicrobial agents to fight antibiotics resistance from different sources and highest concentration has given to screen and evaluate the antimicrobial activity. By using disc diffusion assay method, antimicrobial activity of *Callicarpa attenuate* was evaluated. *E. coli* bacteria (gram negative) and
**Bacillus subtilis** bacteria (gram positive) were used in this study. Mular Hinton agar (MHA) was used as media in this assay. Firstly, every petri dish was autoclaved for sterilization and 20 ml of MHA was poured in every petri dish. After that, the plates were kept for a time being to be settled. With the help of cotton swab, the nutrient broth of bacterial strains was incubated in MHA. Small disc of filter paper was made by using paper punch machine and then different concentrations of plant extract (200 mg/mL and 400 mg/mL) were used to swallow that filter paper. When the discs become dry, they were transferred to the petri dishes and kept in incubator for 24 hours at 37°C. After 24 hours the zone of inhibition were calculated and for keeping the contamination limited, whole experiment was done under laminar flow.

**Hypoglycemia activity**

The anti-diabetic activity of the plant leaves was evaluated with glucose tolerance test. The test was done in two different ways like orally and intraperitoneally.

**Oral glucose tolerance test**

In Oral glucose tolerance test, 24 healthy mice were fasted for 18 hrs. Then they were divided into four groups that contained six mice in each group. Here, 0.9% (w/v) normal saline was given to group I. Group II was received Glibenclamide (250 mg/kg). In addition, group IV and V was received methanol plant extract of 200 mg/kg and 400 mg/kg respectively. After 30 minutes, glucose (3g/kg) was fed. After that at 0, 30, 90, and 120 minutes of glucose administration blood sample were taken from retro-orbital sinus and glucose level was estimated by glucose oxidase-peroxidase method.

**Intraperitoneal glucose tolerance test**

Initially 24 mice were fasted for 18 hours and then they were divided into four groups that contain six rats each. The group of negative control received only 0.9 percentage (w/v) normal saline and standard group received Glibenclamide (250 mg/kg) while the samples were administered the plant extract (200 mg/kg and 400 mg/kg respectively). After 30 minutes, glucose solution (3g/kg) was injected intraperitoneally. At different time after giving glucose solution like t=0, t=30 minutes, t=90 minutes and t=120 minutes, blood sampling was taken and glucose level was determined by using glucose oxidase peroxidase method.

**Antidiarrheal activity**

Two different tests were conducted to evaluate the antidiarrheal activity of the experimented plant.

**Castor Oil-Induced Diarrhea in Rats**

Normal healthy 24 rats were fasted for 18 hours. The rats were divided into 4 groups (n=6). Group I was given normal saline (0.9% w/v) orally and Group II received Loperamide (5 mg/kg) as standard group. Groups III-IV received plant extract (200 and 400 mg/kg b. wt, respectively). After 1 hour, all groups received castor oil 1 mL each orally. Next, all the rats were placed in cages with adsorbent papers and observed for 4 hours for the presence of characteristic diarrheal droppings. 100% was considered as the total number of feces of control group. The activity was expressed as % of inhibition of diarrhea. The % of inhibition was measured by using following formula:

\[
\text{Percent (\%)} \text{ inhibition of defecation} = \left[ \frac{(A-B)}{A} \right] \times 100
\]

Where A is mean number of defecation time caused by castor oil and B is mean number of defecation time caused by drug or extract.

**Magnesium sulfate induced diarrhea**

In the similar protocol as for castor oil induced diarrhea was followed for magnesium sulfate induced diarrhea. Initially, 24 healthy rats were fasted for 18 hours. The rats were divided into four groups that contained 6 rats each. Normal saline (0.9% w/v) was given to group I. Loperamide (5 mg/kg) was given to group II and methanol plant extract (200 mg/kg and 400 mg/kg) was given to group III and IV respectively. After 60 minutes, 1 mL of magnesium sulfate solution was administrated orally and placed in cages lined with adsorbent papers and observed for 4 hours to see the presence of characteristic diarrheal dropping. 100% was considered as total number of feces of control group and % of inhibition was calculated.

**Result and discussion**

**Antioxidant activity**

**Total phenolic content (TPC)**

In total phenolic content test, Gallic acid was used as standard and methanol extract of leaves which was used as a sample. The absorbance of the sample plotted in Gallic acid calibration curve. The absorbance of the plant extract was found 0.453 and TPC value was 67.71 GAE/g against the absorbance of the sample which indicates that the plant has antioxidant activity.

**Total flavonoid content**

The content of total flavonoid of the plant extracts was measured spectrophotometrically by using the aluminium chloride colorimetric assay. The flavonoid content of the extracts was expressed as mg quercetin equivalent per gram of the extract and it is 341.90 QE/g against the absorbance of 0.491 that indicates the present of flavonoid content.

**DPPH free radical scavenging assay**

It is known that DPPH free radical scavenging activity is increasing along with increasing concentration of the methanol extract. As the reference standard, ascorbic acid was used in this experiment for which IC₅₀ value was 55.60 µg/mL. on the other hand, the IC₅₀ value of the methanol extract of the sample plant was 251.82 µg/mL. this result indicates the presence of antioxidant activity which is less significant.

<table>
<thead>
<tr>
<th>Table 1: Evaluation of DPPH free radical scavenging activity of methanol extract of <em>Callicarpa attenuate</em>.</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Sample</strong></td>
</tr>
<tr>
<td>---------------</td>
</tr>
<tr>
<td>Standard</td>
</tr>
<tr>
<td>Sample (methanol extract)</td>
</tr>
</tbody>
</table>
Cytotoxic activity

This brine shrimp lethality assay was used to assess the cytotoxic property of methanol extract of plant material. Here, different concentrations standard and sample were plotted that provided different percentages of mortality. Percentage of mortality was found to increase along with the increasing concentrations of standard and methanol extract. This study indicates the methanol extract of plant material has cytotoxic activity.

Thrombolytic activity

<table>
<thead>
<tr>
<th>Name of the sample</th>
<th>W1</th>
<th>W2</th>
<th>W3</th>
<th>W4</th>
<th>W5</th>
<th>% of clot lysis</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plant extract</td>
<td>0.742</td>
<td>1.558</td>
<td>0.79</td>
<td>0.347</td>
<td>0.372</td>
<td>73.46</td>
</tr>
<tr>
<td>Standard</td>
<td>0.795</td>
<td>1.519</td>
<td>0.272</td>
<td>0.057</td>
<td>0.057</td>
<td>18.90</td>
</tr>
<tr>
<td>Blank</td>
<td>0.795</td>
<td>1.478</td>
<td>0.163</td>
<td>0.468</td>
<td>0.115</td>
<td>22.5</td>
</tr>
</tbody>
</table>

Here, W1 = Micro-tube weight, W2 = Clot with micro-tube weight, W3 = Clot with micro-tube weight after clot disruption, W4 = Clot weight after clot disruption, W5 = Released clot weight. Plasminogen enzyme is usually activated by thrombolytic agents and it also removes fibrin bonds in blood, as a result, the clot becomes soluble and blood flow is restored. Here, methanol extract showed much lower level of thrombolytic activity than standard. Standard gave 18.90% clot lysis, distilled water was used as a negative control, which provided 22.5% clot lysis and methanol extract of plant leaves showed 73.46% clot lysis. After comparing the clots lysis value of plant extract with the positive control value, it was observed that plant material revealed thrombolytic activity but less than standard.

Antimicrobial assay

The plant extract showed antimicrobial activity at all concentrations tested with a broad spectrum of activity, inhibiting against the growth of both Gram positive and Gram-negative bacteria. The antimicrobial potential was especially showed against E. coli and B. subtilis, when exposed to 400 mg/mL of methanol extract of plant and made it impossible when exposed to 200 mg/mL of methanol extract of dried leaves as shown in the table. These results indicate that the antimicrobial activity of the plant extract is not as significant as standard.

Hypoglycemia activity

Table 4: Oral glucose tolerance test in rats as a part of hypoglycemic activity of leaves of Callicarpa attenuate.

<table>
<thead>
<tr>
<th>Group</th>
<th>Dose (mg/kg)</th>
<th>Initial (mmol/L)</th>
<th>30 min. (mmol/L)</th>
<th>90 min. (mmol/L)</th>
<th>120 min. (mmol/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Standard</td>
<td>-</td>
<td>5.23±0.777</td>
<td>19.85±0.777</td>
<td>9.6±0.777</td>
<td>3.11666±0.777</td>
</tr>
<tr>
<td>MEPG 200</td>
<td>400</td>
<td>14.15±1.6467</td>
<td>11.15±1.6467</td>
<td>4.25±1.6467</td>
<td>2.73±1.6467</td>
</tr>
</tbody>
</table>

Table 5: Intra peritoneal glucose tolerance test in rats as a part of hypoglycemic activity of the leaves of Callicarpa attenuate.

<table>
<thead>
<tr>
<th>Group</th>
<th>Dose (mg/kg)</th>
<th>Initial (mmol/L)</th>
<th>30 min. (mmol/L)</th>
<th>90 min. (mmol/L)</th>
<th>120 min. (mmol/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Standard</td>
<td>-</td>
<td>20.333±0.825631</td>
<td>11.633±0.825631</td>
<td>4.95±0.825631</td>
<td>2.883±0.825631</td>
</tr>
<tr>
<td>Control</td>
<td>200</td>
<td>14.66667±0.750333</td>
<td>10.46667±0.750333</td>
<td>4.25±0.750333</td>
<td>4.5±0.750333</td>
</tr>
<tr>
<td>MEPG 400</td>
<td>400</td>
<td>16.15±1.263329</td>
<td>7.716667±1.263329</td>
<td>4.5±1.263329</td>
<td>4.5±1.263329</td>
</tr>
</tbody>
</table>
From the Table 3 and 4 we can say that our sample plant has the ability to act as a potential hypoglycemic medicine. Here MEPG denotes methanol extract of *Callicarpa attenuate*. In both the cases which means in oral and intraperitoneal we saw that the administered glucose level go low as the time increases. If we compare them the intraperitoneal administration of glucose got a high blood glucose level at a short time and it went to low level at a short period of time.

### Antidiarrheal activity

<table>
<thead>
<tr>
<th>Group</th>
<th>Dose (mg/kg)</th>
<th>Castrol oil induced diarrhea</th>
<th>MgSO₄induced diarrhea</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Total number of faeces in 4 hours</td>
<td>Percentage of inhibition</td>
</tr>
<tr>
<td>Control</td>
<td>_</td>
<td>21.33±3.501</td>
<td>48.59</td>
</tr>
<tr>
<td>Standard</td>
<td>8.833±1.839</td>
<td>15.33±2.943</td>
<td>29.13</td>
</tr>
<tr>
<td>MEPG200</td>
<td>200</td>
<td>7.833±1.714</td>
<td>55.28</td>
</tr>
<tr>
<td>MEPG400</td>
<td>400</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

A significant reduction in the number of defection instances was observed with all the test doses of the extract compared with the control group and standard group. There was graded reduction in intestinal fluid volume in graded MEPG extracts. MEPG (400 mg/kg) showed the reduction in the intestinal fluid volume with significant difference as compared with control group and standard group and % inhibition was 55.28% and 48.99% for castor oil induced diarrhea and magnesium sulfate induced diarrhea.

### Conclusion

The plant has been brought into effective action in various traditional uses of which some have been proved clinically. Further studies can be conducted on secondary metabolites to explore more activities. This review can be helpful in promoting research that can help to develop new agents for therapeutic applications based on bioactive chemical compounds. Therefore, this plant is significantly used for the treatment and prevention of diseases.

### References