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# Evaluation of curcumin compound from methanolic extracts of *Curcuma longa* by high performance thin layer chromatography

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## Abstract

*Curcuma longa* (turmeric) is a medicinal plant belonging to the family Zingiberaceae. It is widely used for the health care of humans, livestock and poultry. Turmeric is a potent antioxidant, anti-inflammatory, antimutagenic, antimicrobial, and anticancer agent. Curcumin is a polyphenol extracted from roots of turmeric. In the present study, fresh roots of *Curcuma longa* was collected from Herbal Garden, Ethno Veterinary Herbal Product Research and Development Centre, Tamil Nadu Veterinary and Animal Sciences University, Orathanadu, Thanjavur District, Tamil Nadu and evaluated for the identification and relative quantification of curcumin by using a simple, sensitive and accurate High Performance Thin Layer Chromatography method.

Keywords: Curcumin, turmeric, polyphenol, high performance thin layer chromatography

# Introduction

Medicinal plants are widely used for curing various diseases in animal kingdom. Phytochemicals play a major role in plant defence mechanisms against predators and pathogens (Divisha *et al.*, 2018) <sup>[6]</sup>. They are majorly classified into carotenoids and polyphenols which include phenolic acids, flavonoids, stilbenes and lignans (Molyneux *et al.*, 2007) <sup>[8]</sup>. *Curcuma longa*, commonly known as Turmeric, a rhizomatous herbaceous perennial plant belonging to the ginger family Zingiberaceae, which is native to tropical South Asia. It is a medicinal plant that has a very long history of medicinal use, dating back nearly 4000 years. In Southeast Asia, turmeric is used not only as a principal spice but also as a component in religious ceremonies. Because of its brilliant yellow color, turmeric is also known as "Indian saffron". The rhizome, from which the turmeric is thought to have many medicinal properties including strengthening the overall energy of the body, relieving gas, dispelling worms, improving digestion, regulating menstruation, dissolving gallstones, and relieving arthritis. Many South Asian countries use it as an antiseptic for cuts, burns, and bruises, and as an antibacterial agent (Benzie and Wachtel-Galor, 2011) <sup>[3]</sup>.

Curcumin (diferuloylmethane), a water-insoluble polyphenol extracted from the herb Curcuma longa L. has been widely used in traditional Indian and Chinese medicine as well as in the food industry. This polyphenol has been demonstrated for several types of biological and pharmacological activities, including anti-inflammatory, anti-oxidant and anti-cancer properties, etc. (Mishra et al., 2017)<sup>[7]</sup>. This compound is the most important active ingredient responsible for the biological activity of turmeric It is the natural yellow pigment present in the roots of turmeric (Badreldin et al., 2006)<sup>[2]</sup>. Chemical synthesis of curcumin analogues has resulted in compounds with stronger anti-oxidant and cancer chemoprotective activities (Youssef et al., 2004) <sup>[11]</sup>. Curcumin protects human keratinocytes from xanthine oxidase injury by its anti oxidant activity (Bonte et al., 1997)<sup>[4]</sup>. Several studies have estimated phenol contents in medicinal plants using Thin Layer Chromatography (Ranganathan and Punniamurthy, 2013)<sup>[10]</sup> and using High Performance Thin Layer Chromatography (Aftab et al., 2015) <sup>[1]</sup>. In the present study, a sensitive, simple and accurate HPTLC method was established to identify and evaluate curcumin from turmeric sample cultivated at herbal garden of Ethno Veterinary Herbal Product Research and Development Centre, Tamil Nadu Veterinary and Animal Sciences University, Orathanadu, Thanjavur District, Tamil Nadu.

## Materials and Methods Plant Source

Fresh roots of *Curcuma longa* was collected from Herbal Garden, Ethno Veterinary Herbal Product Research and Development Centre, Tamil Nadu Veterinary and Animal Sciences University, Orathanadu, Thanjavur District, Tamil Nadu. The roots were cleaned, shade-dried and powdered for further analysis.

### **Preparation of methanolic extract**

25g of the powdered sample was extracted with 125ml of methanol for 6 hours in a Soxhlet apparatus (Uma *et al.*, 2009). The extracts were subjected to Rotary evaporator to allow the solvent to evaporate completely. Standard curcumin was purchased from Sigma Aldrich (USA). All the solvents used in the study were of HPLC grade.

# Identification of curcumin from turmeric using High Performance Thin Layer Chromatography (HPTLC)

The chromatographic procedure was carried out with silica gel 60 F254 Thin Layer Chromatographic (TLC) plate (Merck). Seven different volumes-1,2,3,4,5, 6 and 7µL of standard curcumin solution (1 mg/mL) and duplicates of 2, 4, 6 and 8 µL of turmeric samples (50 mg/ml) were applied to TLC plates by using CAMAG Linomat 5 sample applicator. Type of the application was banding and applied on Y axis of the plate with 8.0mm distance and the length was also maintained at 8.00 mm distance. The plate was then developed at room temperature in a CAMAG twin-trough vertical development chamber (20x10) already saturated for 20 min with the mobile phase of toluene: acetic acid (8:2 v/v). The solvent front position was maintained at 70 mm. After the development process, the plate was subjected to CAMAG visualizer2 for screening the developed plate using tungsten light source at a wavelength of 540 nm, maintaining a scanning speed of 20 mm/s and slit dimension of 6.0 mm x 0.45 mm. The scanning procedure was carried out using vision CATS software (version 2.5.18262.1). The detection of curcumin present in turmeric samples was observed on the HPTLC plates in the form of bands that held similar Retardation factor (Rf) values as that of curcumin standard. The Retardation factor (Rf) which indicates the ratio of the distance travelled by the compound to the distance travelled by the solvent in a given time was measured for both standard and sample spots.

### **Relative quantification of Curcumin in turmeric samples**

The area calibration was measured for substance curcumin @ 540nm using CAMAG scanner (S/N: 250410). Four reference (Standard) samples and five turmeric samples were subjected for creating standard curve for the evaluation of curcumin from turmeric samples.

### **Results and Discussion**

The table 1 showed the Rf values analysed by High Performance Thin Layer Chromatography (HPTLC) for curcumin standard and turmeric samples indicating the presence of curcumin in the turmeric samples. Fig.1 and fig.2 showed development of TLC plate with remission of 540 nm and identification of curcumin in turmeric sample with Rf value, respectively. Several research works utilized HPTLC for identification and evaluation of medicinal phytochemistry in view of its sensitivity and accuracy. Mishra et al. (2017)<sup>[7]</sup> also identified curcumin in plasma samples by using HPTLC method. Corina et al (2019)<sup>[5]</sup> analysed iridoids from Lamium album using HPTLC methods. Principal component was analysed for root extracts of Withania coagulans by using this chromatogram (Preethi et al., 2014)<sup>[9]</sup>. Studies so far conducted with CAMAG HPTLC used win CATS software whereas the present study was conduted with the sophisticated recent software known as vision CATS software. Moreover, no reports are available with regard to evaluation of curcumin from turmeric cultivated at Thanjavur district of Tamil Nadu.

**Table 1:** HPTLC –Curcumin profile of methanolic extract of turmeric samples (curcumin-1mg/ml; turmeric samples-50mg/ml)

Track	Rf	Height	Area	Assigned substance
Curcumin standard	0.397	0.1700	0.0040	Curcumin
Curcumin standard	0.384	0.1665	0.0038	Curcumin
Curcumin standard	0.384	0.1994	0.0055	Curcumin
Curcumin standard	0.385	0.2188	0.0063	Curcumin
Curcumin standard	0.384	0.2336	0.0065	Curcumin
Curcumin standard	0.381	0.2408	0.0071	Curcumin
Curcumin standard	0.384	0.2051	0.0063	Curcumin
Turmeric sample	0.387	0.2059	0.0052	Curcumin
Turmeric sample	0.384	0.2106	0.0052	Curcumin
Turmeric sample	0.384	0.2552	0.0073	Curcumin
Turmeric sample	0.385	0.2595	0.0074	Curcumin
Turmeric sample	0.381	0.2754	0.0087	Curcumin
Turmeric sample	0.381	0.2824	0.0089	Curcumin
Turmeric sample	0.382	0.2993	0.0100	Curcumin
Turmeric sample	0.385	0.3025	0.0100	Curcumin

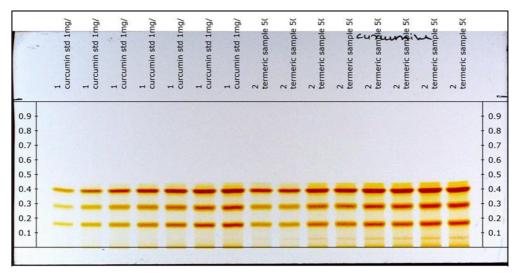


Fig 1: HPTLC Plate developed under visible remission (540nm) showing bands of standards and samples ~ 1932 ~

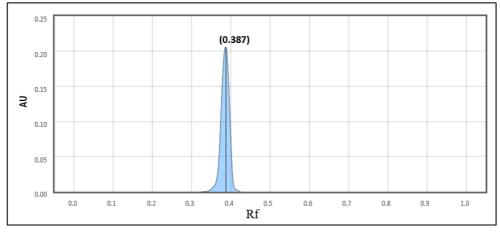


Fig 2: HPTLC plate track peak for identification of curcumin in turmeric sample

For quantification purpose, calibration curve was plotted with five reference samples and four turmeric samples in the CAMAG scanner. Coefficient variation (CV) was 2.49% and correlation coefficient was 99.308583%. Based on the calibration curve, curcumin present in the turmeric sample was evaluated to be 1.631 mg in 50 mg of turmeric sample. The calibration curve was linear within a range of 1.631mg/ml of turmeric sample. Fig.3 showed the calibration curve for relative quantification of curcumin in the turmeric sample.

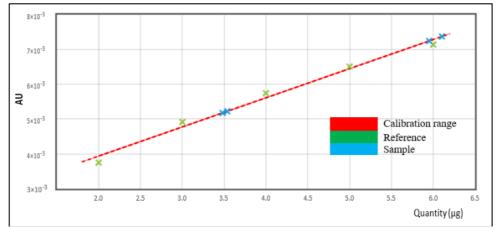


Fig 3: HPTLC calibration curve for evaluation of curcumin in turmeric sample (50mg/ml)

# Conclusion

The results obtained in the present study suggest that the roots of *Curcuma longa* cultivated at herbal garden of Ethno Veterinary Herbal Product Research and Development Centre, Veterinary College and Research Institute, Orathanadu contain curcumin, which has potential for use in the health care of animal kingdom.

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# References

- Aftab A, Asif H, Mohd M, Shah AK, Hani AAA, Anil B. Quantification of total phenol, flavonoid content and pharmacognostical evaluation including HPTLC fingerprinting for the standardization of *Piper nigrum* Linn fruits. Asian Pacific Journal of Tropical Biomedicine. 2015; 5(2):101-107.
- Badreldin HA, Husnia M, Salwa AN, Amel OB, Gerald B. Some biological properties of curcumin: A review. Natural Products Communications. 2006; 1(6):509-521.

- 3. Benzie IFF, Wachtel GS. Chapter 13-Turmeric the gold spice: Herbal medicine- Biomolecular and clinical aspects (II Edition)-CRC Press/Taylor and Francis, 2011.
- 4. Bonte F, Noel-Hudson MS, Wepierre J, Meybeck A. Protective effect of curcuminoids on epidermal skin cells under free oxygen radical stress. Planta Medica 1997; 63:265-266.
- 5. Corina B, Rasit I, Minerva P. Haemostatic activity of butanolic extracts of *Lamium album* and *Lamium purpurium* aerial parts, Acta Pharm. 2019; 69:443-449.
- Divisha R, Ranganathan V, Vijayakaran K, Elamaran A. Evaluating *Ocimum basilicum* and *Ocimum tenuiflorum* leaf extracts for the presence of phenolic compounds. Journal of Pharmacognosy and Phytochemistry. 2018; 7(6):2453-2456.
- 7. Mishra J, Ankit T, Kshipra M, Sarada SKS. HPTLC: A tool for determination of curcumin in mammalian samples. International Journal of Pharmacognosy and Phytochemical Research. 2017; 9(4):498-501.
- 8. Molyneux RJ, Lee ST, Gardner DR, Panter KE, James LF. Phytochemicals: the good, the bad and the ugly?. Phytochemistry. 2007; 68(22-24):2973-85.
- 9. Preethi MP, Sangeetha U, Pradeepa D, Valizadeh M, Kalaiselvi S. Principal compound analysis and HPTLC fingerprinting of *in vitro* and field grown root extracts of

*Withania coagulans*. International Journal of Pharmacy and Pharmaceutical Sciences. 2014; 6(5):480-488.

- 10. Ranganathan V, Punniamurthy N. Estimation of phenol contents in glycirrizha glabra by thin layer chromatography and spectrophotometry. International Journal of Agricultural Sciences and Veterinary Medicine 2013; 1(3):24-26.
- 11. Youssef KM, El-Sherbeny MA, El-Shafie FS, Farag HA, Al-Deeb OA, Awadalla SR. Synthesis of curcumin analogues as potential antioxidant, cancer chemopreventative agents. Archiv der Pharmazie (Weinheim). 2004; 337:42-54.