

# Journal of Pharmacognosy and Phytochemistry

J Journal of Ptampacogussy and Phylochemistry

Available online at www.phytojournal.com

E-ISSN: 2278-4136 P-ISSN: 2349-8234 JPP 2015; 4(1): 140-147 Received: 28-03-2015 Accepted: 30-04-2015

#### Priva Sharma

University Institute of Engineering and Technology Maharshi Dayanand University, Rohtak-124001, India.

#### Sonia Kapoor

University Institute of Engineering and Technology Maharshi Dayanand University, Rohtak-124001, India.

#### Correspondence: Sonia Kapoor

University Institute of Engineering and Technology Maharshi Dayanand University, Rohtak-124001, India.

## Biopharmaceutical aspects of Brassica vegetables

## Priya Sharma, Sonia Kapoor

#### Abstract

Brassica is a genre of plants belonging to family Brassicaceae commonly used as vegetables and oilseed. The members of Brassicaceae are devoured by natives throughout the world. They comprise important food crops in Europe, Japan, China and India and thus symbolize a vital portion of human regimen worldwide. Brassica vegetables frequently exploited for food include broccoli, cauliflower, Brussels sprouts, cabbage and certain seeds. They are important source of bioactive compounds and nutrients like Vitamin E and C, soluble fiber, enzymes owing antioxidant activity for example peroxidase, superoxide dismutase (SOD) and catalase and carotenoids which have persuasive antiviral, antibacterial and anticancer activity. Brassica vegetables have positive impacts on human vigour which are somewhat accredited to their composite blend of phytochemicals possessing antioxidant activity that fights against free radicals by acting on complementary and transformed levels. They basically lead to the stimulation of detoxification enzymes, induce immune system, preclude oxidative stress, diminution of threat of cancers, obstruction of carcinogenic mutations and malignant transformation besides lessening explosion of cancerous cells. These vegetables are also a major source of valuable metabolites, which embrace anthocyanins, terpenes, S-methyl cysteine sulfoxide, sulforaphane, selenium, coumarins and glucosinolates. Glucosinolates breakdown into diverse metabolic products which act as modulators to protect against DNA damage. They may abolish or nullify innumerable mutagenic and carcinogenic factors through consequently inhibiting DNA methylation which mainly occurs via the initiation of enzymatic systems I and II phase of xenobiotics metabolism. Genetic makeup, environmental impact, cultivation strategy, biochemistry of plants and type of processing and storage, fundamentally outline the concentration and bioavailability of these constituents. Therefore, we can say that the consumption of vegetables including Brassica species is strongly connected with the prevention against threat of numerous types of chronic diseases like Alzheimer's disease, cardiovascular disease, diabetes, cancer, age-related efficient waning and cataracts etc. and thus plays a crucial function in maintaining healthy human life.

**Keywords:** Anticancer, *Brassica* vegetables, glucosinolates, isothiocyanates, neurodegenerative diseases, phytochemicals, sulforaphanes.

## 1. Introduction

Plant-centred diets comprise substantial quantities of bioactive compounds, which deliver necessary health profits yonder simple nourishment [1]. The consumption of a diet rich in *Brassica* vegetables has progressive insinuations for human health which have been implied by various epidemiological evidences (Fig.1) [1, 2]. Greater devotion has been waged towards edible plants in the last decades, exclusively those that are rich in phytochemicals (secondary metabolites) and nowadays, a special interest is developing in the antioxidant activity of such phytochemicals [1]. Cruciferous vegetables rich diet has been related with inferior rates of coronary heart disease and cancer [3-8]. Plant-based terpenes, flavonoids, phenols, glucosinolates, isoflavones together with many other compounds that are existing in the daily diet are described to have anticarcinogenic and antioxidant properties plus a catholic gamut of anti-tumor actions [4, 7, 9, 10]. These protective effects of *Brassica* vegetables are mainly accredited to the presence of vast quantities of glucosinolates, that distinguishes them from other vegetables. Vegetables of the *Brassica* genus, including broccoli, kale, cabbage, Brussels sprouts, cauliflower, black and brown mustard, kohlrabi, root crops like turnips and rape subsidize utmost to glucosinolates intake [11].

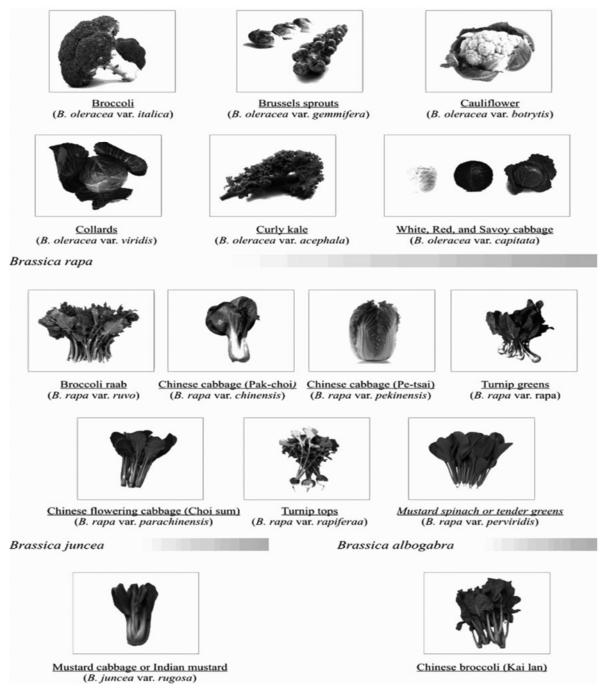


Fig 1: Scientific and dialect names of marketed Brassicaceae [2].

Glucosinolates are hydrolysed into two principal components i.e., isothiocyanates and indoles owing anti-carcinogenic properties. Glucosinolates together with their consequent products have been reported to reduce the menace of certain types of cancers in humans like lung, breast, colon, rectum, and prostate cancers and thereby provide vital health benefits [10]. An enzyme called myrosinase is present in the membranes of plant cells which is responsible for the hydrolysis of glucosinolates. This enzyme comes in contact with glucosinolates only when plant gets damaged (e.g. by cutting or chewing) and as a result hydrolysis takes place. All glucosinolates present in *Brassica* family primarily show a familiar basic frame, with having difference in only their side chain (R) where R may be an alkenyl, aryl, alkyl, alkylthioalkyl, indolylmethyl or 3-hydroxyalkyl (Fig. 2) [12].

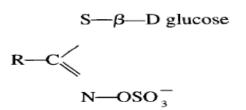


Fig 2: Basic structure of glucosinolates.

The hydrolysis products of glucosinolates entail equimolar amounts of sulfate, glucose and aglucon. The aglucones are unsteady and experience additional reactions that may lead to the production of various useful components, like nitriles, thiocyanates, isothiocyanates, or indoles (Fig. 3) [12, 13]. The

behaviour of these hydrolysis products may vary depending principally upon the presence of cofactors, the hydrolysis conditions and the side chain of glucosinolate [12].

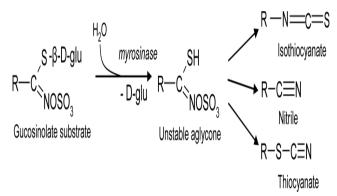


Fig 3: Conversion of Glucosinolates into Isothiocyanates by Plant Myrosinase [13]

Several experimental studies have demonstrated that the intake of isothiocyanates and indoles in animals (after a carcinogen) is associated with the reduction of tumour incidence and multiplicity [14-17]. The plausible inhibitory activity of isothiocyanates and indoles in opposition to tumorigenesis is mainly because of their capability of influencing enzyme activities of phase 1 and 2 biotransformation and as a result several processes correlated with chemical carcinogenesis get manipulated, such as DNA binding potential of carcinogens and their metabolism [18-20].

Owing to the wide range of bioactive compounds in *Brassica* genus, the attention of food scientists has shifted towards the hidden function of isolated phytochemicals. For instance, an essential nutrient of *Brassica* members i.e., Vitamin C, deficiencies an integrative approach to recognize its functions on health across with the habituation of food matrix on its bioavailability and rest of bioactive components in their usual food concentrations [21-26].

## 2. Phenolic Compounds

The health potential of Brassica vegetables are partially attributed to their intricate fusion of phytochemicals owing antioxidant activity. Recently, considerable research has been aimed at the detection of plant derived natural antioxidants which can be utilized for human consumption for prevention of non-transmissible chronic diseases and promotion of health. Phenolic compounds are one of the most vital groups among phytochemicals that possess antioxidant capacity [27]. The term "Phenolic compounds" refers to a large number of compounds, nearly 8000, extensively distributed all over the plant kingdom exemplified by having at least one aromatic ring with involvement of one or more hydroxyl groups. They are one of the most important secondary metabolites in plants and are produced via shikimic acid pathway [1]. The aromatic amino acid phenylalanine act as a precursor for their biosynthesis and the enzyme involved is known as phenylalanine ammonialyase (PAL).

Phenolics range from single aromatic-ringed, modest and low molecular weight compounds to multifarious and heavily stemmed polyphenols and tannins (Fig. 4) [28-30]. They have been classified depending on the arrangement and number of their carbon atoms in flavonoids (flavanones, flavones, anthocyanidins, flavan-3-ols, isoflavones, flavonols and many others) and non-flavonoids (stilbenes, hydroxycinnamates,

phenolic acids and others) [29]. Phenolics are usually discovered conjugated to organic acids and sugars [1]. The most pervasive and diverse group of polyphenols in *Brassica* species are hydroxycinnamic acids and the flavonoids (mainly anthocyanins and flavonols). But the composition of phenolic compounds can be pretty diverse among crops from the same species or even in the single species itself.

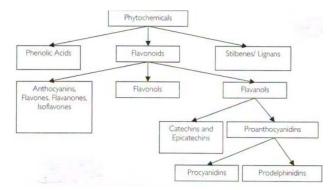


Fig 4: Classification of Phytochemicals [30].

The qualitative and quantitative profiles of different *Brassica* species have been elucidated showing their diverse polyphenol composition as well as antioxidant activity [1]. For example, Phenolic profiles of several *Brassica* species has been comprehensively reviewed by Podsedek [31]. The highest antioxidant activity was exhibited by red cabbage followed by green cabbage, while Chinese cabbage, Mustard cabbage and Chinese white cabbage have been shown to possess lower antioxidant potential as compared to these two [31]. According to a study done by Podsedek *et al.* [32] as compared to *B. oleracea*, red cabbage and Brussels sprouts have 5 to 2.2-fold higher antioxidant activity than savoy and white cabbages which may be attributed to different range of bioactive compounds in the respective species.

## 3. Health Benefits of Brassica Vegetables

The potential health benefits of polyphenols mainly depend on their bioavailability and disbursed amount [1]. The concern in the biological possessions and the bioavailability of flavonoids and phenolics in food plants is being rising over the past few decades. Attracting insects for pollination and seed dispersion is one of the important supplementary roles of phenolic compounds. Moreover, they also provide resistance against fungi, insects, bacteria and viruses by acting as vital hormone controllers of plants. Likewise, because of their potential health-promoting effects, phenolic compounds have been intensively investigated in recent years [26, 33-36].

Phytochemicals from *Brassica* vegetables may act on distinctive and complementary levels [36] and they possess many advantages for the health benefits for human beings including enzyme inhibition, anti-inflammatory, antimicrobial, cytotoxic and vascular antitumor activity, antiallergic etc., but the antioxidant potential of phenolics is their most crucial attribute [31, 32, 35, 37-40]. Moreover, phenolic compounds owe other vital aspects such as the inhibition of nitrosation reactions, chelating metal ions and production of hydrogen peroxide in the incidence of some metals etc. They also have the aptitude of scavenging electrophiles and consequently, they act by blocking the onset of wide range of human diseases [31, 41-43]

The phenolic compounds impart positive effects on human health by virtue of antioxidant activity associated with their

chemical structure which bestows redox properties. These compounds can play a vital role in decomposition of peroxides, adsorbing and neutralizing reactive oxygen species (ROS) and quenching of singlet and triplet oxygen. Reactive oxygen species (ROS) can instigate DNA damage, protein and lipid oxidation, modulation of gene expression and base modification in the body [36]. Moreover, they also play a central role in etiopathology of diseases, like atherosclerosis, vasospasm, cancers, stroke, heart attack and liver injury. ROS, originate in the body from various processes of oxidation, are significant ingredients of the resistant mechanisms against infection. However, if generated in excess, these free oxygen radicals may lead to the damage of the tissue [1]. Disparity between ROS and antioxidants may lead to oxidative stress due to accumulation of lipid peroxides. The various reasons of oxidative stress includes antioxidant scarcity in the diet or increased production of free radicals by stress, smoking, environmental contaminations, which immigrate into water and food (heavy metals, pesticides, nitrates, nitrites, nitrosamines etc.) [44]. Other risks are mutagenic and carcinogenic substances present in food products due to inappropriate storage conditions (eg. mycotoxins) or heat treatment (e.g. heterocyclic aromatic amines, acrylamide, genotoxic lipid peroxidation products) [36]. The existence of antioxidants, and chiefly their release during heat treatment, suggest that cabbage can effectively protect other food ingredients against thermo-oxidative changes [45].

Consumption of vegetables including Brassica species has been strongly connected with the reduced risk of chronic diseases, like cancer, cardiovascular disease, Alzheimer's disease, diabetes, age-related functional decline and cataracts etc. [46-48]. Vitamin C which is a vital component of Brassica vegetables plays a major role in the maintenance of body functions. It demonstrates a broad array of therapeutic properties that include anticarcinogenic, antioxidant, iron absorption promoter and also acts as a cofactor in the synthesis of collagen. The health-promoting properties of vitamin C have been validated by several reports [49]. Vitamin C and phenolic compounds account for 80 % natural antioxidant activity in *Brassica* vegetables [31]. In cabbage and broccoli, vitamin C is accountable of 10-12% of the total antioxidant capacity [31]. Vitamin C is involved in a broad array of essential reactions in the body responsible for human fitness despite of that the phenomenon is not fully understood yet. Many health promoting properties of Vitamin C have been suggested such as the protection against free radicals, its antioxidant properties, cytoprotective functions like prevention of DNA mutation, repairing amino acid residues to save the integrity of protein and protection against lipid peroxidative damage etc. [50-52].

## 4. Role of Isothiocyanates (ITC)

The existence of glucosinolates in cruciferous vegetables is their most prominent and distinctive chemical property. Both the glucosinolates and their hydrolysis products i.e., isothiocyanates are eminent armaments to counter carcinogenesis which signifies that if these vegetables are consumed in large amount, they may dampen the hazard of several forms of cancer. Around 130 chemically diverse glucosinolates have been recognized [53]. In animal models, isothiocyanates (both natural and synthetic) have captivated mounting and significant research as efficient and imperative protectors against chemical carcinogenesis, since the early 1960s [54]. However, due to unavailability of satisfactory

quantities of these compounds, only a few glucosinolates have been examined which seem to be very effective in impeding carcinogenesis [54].

Specific attention has been focused on the isothiocyanates, befalling in *Brassica* vegetables, for their potential anti-cancer abilities <sup>[36]</sup>. However, the structure of ITCs have a major impact on transactivation of NF- κB and also possess anti-inflammatory action, but only a few studies have been carried out <sup>[55, 56]</sup>. ITCs, specific to diverse range of cruciferous vegetables, can prevent human from a number of diseases like cancer, chronic–degenerative diseases, diabetes, cardiovascular diseases and neurodegeneration etc. <sup>[57]</sup>. Constant inflammation plays a critical role in sundry human illnesses and ITCs reduce inflammation by permanently inactivating the migration of inhibitory factor of macrophages and restraining of cyclooxygenase 2 <sup>[57]</sup>.

Sulforaphane (SFN), one of the most important ITCs of *Brassica* vegetables has been established to possess a range of defensive effects in models of cancer and tissue injury. There are two types of xenobiotic metabolizing enzymes i.e., Phase I and II which are expressed in epithelial cells (including those of colon) and in liver whose equilibrium can be amended by sulforaphane [58]. Glutathione transferase family (GST) represents an important type of Phase II enzymes that can metabolize the products of Phase I activity and thus, may lead to the formation of water-soluble and sedentary conjugates which are promptly defecated in urine [58].

#### 5. Acute Neurodegeneration

**a.) Ischemic brain injury:** Basically involves specific biochemical mechanisms, like the origination of ROS, glutamate-mediated excite-toxicity and apoptosis together with inflammation <sup>[59]</sup>. In case of neonatal hypoxia model, <sup>[60, 61]</sup> scrutinized that the expressions of Nrf2 and HO-1 were pointedly increased by SF which was escorted by reduced infarct volume.

**b.) Traumatic Brain Injury (TBI):** Initiated by external mechanical force and characterized by the mutilation of brain <sup>[62]</sup>. Recently it has been shown that administration of SF after TBI reduces the cerebral edema and BBB impairment in rats <sup>[63, 64]</sup>. Also Zhao *et al.* <sup>[64]</sup> reported that after the interval of 24 h and 3 days following TBI, the attenuation of SF resulted in the loss of aquaporin-4 (AQP4) channel in the core of injury together with the increment of AQP4 protein levels in the penumbra region. AQP4 channels functions as to clear the excess water and thus maintain the water homeostasis of brain which was proved by monitoring the diminution in cerebral edema only at 3 days of TBI <sup>[65]</sup>.

#### 6. Chronic Neurodegeneration

**a.) Alzheimer's Disease:** It is the most ordinary neurodegenerative disease suffered by older people that results in dementia. The consequences of Alzheimer's Disease involves impairment of at least one cognitive function and progressive decline in memory  $^{[66]}$ . In this context, the neuroprotective effects of SF against oxidative stress has been revealed by Kwak *et al.*  $^{[67]}$ , in terms of cytotoxicity elicited by hydrogen peroxide and formation of protein carbonyl which occurs as a result of induction of proteasome expression in Neuro2A cells (murine neuroblastoma) by SF. Park *et al.*  $^{[68]}$  also demonstrated the capacity of SF to protect the neuronal cells from  $A\beta 1$ –42-mediated cytotoxicity and increase in activities of proteasome, in other similar cellular models.

b.) Parkinson's Disease: Parkinson's disease, a type of neurodegenerative disease related with age, is characterized by congregation of neuronal inclusions identified as Lewy bodies and with gradual loss of dopaminergic (DA) neurons in the substantia nigra pars compacta [69]. However, the exact etiology of PD has not been fully elucidated, it mainly has an environmental [70, 71] or a genetic [72] origin, or a combination of both as recommended by many reliable theories. The crucial protein which is involved in the pathogenesis [73-75] of this disorder is a prime component of Lewy body inclusions namely  $\alpha$ -synuclein protein [76] as demonstrated by diverse genetic studies. Although it has been detected that a superfluous of  $\alpha$ -synuclein protein can cause loss of DA neuron, but the mechanism by which mutations in this gene lead to neuron loss and precise biological function of  $\alpha$ synuclein are still not clear [77].

SF is also capable of significantly diminishing the levels of DA quinone in dopaminergic cell lines, for example SK-N-BE(2)C, CATH.a in addition to mesencephalic dopaminergic neurons (aroused by BH4 and 6- hydroxydopamine (6-OHDA) [78] as indicated by in vitro studies. According to Han *et al.* [79], SF can safeguard dopaminergic cells from the cytotoxicity of 6-OHDA and BH4 as the activity of NQO1 enzyme and mRNA level are amplified and the amount of quinone-modified proteins is diminished by SF treatment.

#### 7. Cancer Prevention

The development of cancer is a multifaceted and prolonged process involving initiation, elevation and progression [80]. The consumption of Cruciferous vegetables seems to lower the threat of certain types of cancers, like prostate, colorectal and renal cancers due to the presence of glucosinolates [81]. However, the evidence for breast, oral and lung cancers is not as strong [81]. The extracts of fresh *Brassica* vegetables including Brussels sprouts, cauliflower and broccoli has been shown to possess the highest antioxidant and anti-proliferative activities on carcinoma HT-29 cells and moreover, the tested samples were free from any genotoxic activity [80].

The anti-myeloma activity of the phenethyl isothiocyanate and sulforaphane on human primary myeloma tumor cells along with panel of myeloma cell lines showed that isothiocyanates may hold powerful anti-myeloma activities together with the enhancement of action of additional anti-multiple myeloma agents [82].

The chief descriptive of the flavonol subclass in Brassica vegetables, i.e., quercetin (found at high concentration in broccoli) has gained significant attention in recent years because of its high potential for preventing the oxidation of LDL by chelating transition metal ions and scavenging free radicals [1]. These properties are deliberated due to a 4-oxo function at the C-ring, unsaturation at the C-ring and dihydroxylated B-ring [1]. Hence, quercetin can prevent humans from various kinds of fatal diseases, for example, chronic inflammation, atherosclerosis and most importantly cancer by induction of enzymes which can detoxify 30 types of carcinogens that may cause cancer and through retardation of oxidative degradation [83-85]. The reduction of the risk of prostate cancer through the intake of diet rich in broccoli have been suggested through different epidemiological studies. It has been shown that if one or more portions of broccoli is consumed per week, it can significantly lessen the incidence and the development of prostate cancer from confined to belligerent forms [86, 87].

The inhibition of the expansion of colorectal cancer in animal

models has been indicated through some experimental evidences, when given either before, or after, treatment with a carcinogen [88]. It has been shown that a diet enriched in sinigrin, which is a glucosinolate precursor of allyl isothiocyanate, can suppress the induction of enhanced level of apoptosis and mitosis in the colorectal crypts of rats, 48 h after treatment with the colon carcinogen dimethyl hydrazine (DMH) in an in vivo study [89]. However, it resulted in the substantial destruction of aberrant crypt foci (kind of precancerous lesions) [88]. But, importantly, there was no momentous effect on kinetics of crypt cell and their apoptosis, in the colon of control rats (not treated with a carcinogen) due to sinigrin [90]. The effects similar to those of AITC on HT29 cells have been shown to be exerted by the juice of uncooked Brussels sprout tissue (prepared by mechanical disruption) in vitro [91]. Moreover, in a rodent model, there was a marked increment in the apoptosis of crypt cell after treatment with DMH by the same juice given by gavage [91]. All these results collectively suggest that a discriminatory effect in contradiction of the evolution of colorectal epithelial cells carrying DNA damage may be exerted by the isothiocyanates together with their dietary components in vivo. Nevertheless, it has not yet been recognized that the intake of diet rich in breakdown products of glucosinolates leads to the induction of suppression and apoptosis of the cell cycle in the colorectal mucosa of human, hence warrants further research.

#### 8. Conclusions

Brassica vegetables have long been regarded as excellent source of nutraceuticals attributed to the structural diversity of the bioactive components and their wide array of beneficial effects on human health. These possess a large gamut of therapeutic effects, like antibacterial, antifungal, antitumor, anti-mutagenic, anti-inflammatory, neuroprotective and antioxidative activities that have been demonstrated and validated through many in vivo and in vitro pharmacological studies. Increased consumption of these vegetables is highly desirable in recent times. The numerous types of supplements containing purified compounds which are derived from *Brassica* species, should be viewed with prudence. The protective effects from Brassica vegetables may be increased through contentenhanced functional foods, however, these should be tested for safety. Diverse kinds of cooking methods like boiling, frying and microwave cooking etc. should also be optimized in order to maintain the original and natural content of bioactive compounds in processed foods. Therefore, we can conclude that Brassica vegetables play a vital role in prevention of nontransmissible chronic diseases and maintenance of healthy wellbeing.

### 9. References

- Cartea ME, Francisco M, Soengas P, Pablo Velasco P. Phenolic Compounds in *Brassica* Vegetables. Molecules 2011; 16:251-280. doi:10.3390/molecules16010251.
- Dominguez-Perles R, Mena P, Garcia-Viguera C, Moreno DA. *Brassica* Foods as a Dietary Source of Vitamin C: A Review. Critical Reviews in Food Science and Nutrition 2014; 54:1076-1091.
- Drewnowski A, Gomez-Carneros C. Bitter taste, phytonutrients, and the consumer: a review. Am J Clin Nutr 2000; 72:1424-35.
- Craig WJ. Phytochemicals: guardians of our health. J Am Diet Assoc 1997; 97:S199-204.
- 5. Beecher GR. Phytonutrient's role in metabolism: effects

- on resistance to degenerative processes. Nutr Rev 1999; 57:S1-6.
- Lichtenstein AH. Soy protein, isoflavones and cardiovascular disease risk. J Nutr 1998; 128:1589-92.
- Potter JP. Food, nutrition and the prevention of cancer: a global perspective. Washington, DC: World Cancer Research Fund, 1997.
- Steinmetz KA, Potter JD. Vegetables, fruit and cancer prevention: a review. J Am Diet Assoc 1996; 96:1027-39.
- 9. Rhodes MJC. Physiologically-active compounds in plant foods: an overview. Proc Nutr Soc 1996; 55:371-84.
- Zhang Y, Talalay P, Cho C-G, Posner GH. A major inducer of anti carcinogenic protective enzymes from broccoli: isolation and elucidation of structure. Proc Natl Acad Sci U S A 1992; 89:2399-403.
- 11. Fenwick GR, Heany RK, Mullin WJ. Glucosinolates and their breakdown products in food and food plants. CRC Crit. Rev. Food Sci Nutr.1983; 18:123-201.
- Dorette TH, Verhoeven 1R, Goldbohm A, PoppelGV, Verhagen H, Piet A. van den Brandt. Epidemiological Studies on *Brassica* Vegetables and Cancer Risk 1996; 5:733-748.
- 13. http://www.mdpi.com/1422-0067/10/8/3371/htm [date accessed May 1, 2015].
- 14. Morse MA, Eklind KI, Hecht SS, Chung FL. Inhibition of tobacco-specific nitrosamine 4-(N-Nitrosomethylamino)-l-(3-pyridyl)-l-butanone(NNK) tumorigenesis with aromatic isothiocyanates. Scientific Publ. Lyon, France: IARC 1991; 105:529-534.
- 15. Sugie S, Okumura A, Tanaka T, Mori H. Inhibitory effects of benzyl isothiocyanate and benzyl thiocyanate on diethylnitrosamine-induced hepato-carcinogenesis in rats. Jpn J Cancer Res 1993; 84:865-870.
- Dashwood RH, Arbogast DN, Fong AT, Pereira C, Hendricks JD, Bailey GS. Quantitative inter-relationships between aflatoxin B 1 carcinogen dose, indole-3-carbinol anti-carcinogen dose, target organ DNA adduction and final tumor response. Carcinogenesis (Lond.) 1989; 10:175-181.
- Tanaka T, Moil Y, Morishita Y, Hara A, Ohno T, Kojima T *et al.* Inhibitory effect of sinigrin and indole-3-carbinol on diethyl nitrosamine induced Hepatocarcinogenesis in male ACI/N rats. Carcinogenesis (Lond.) 1990; II:1403-1406
- 18. Zhang Y, Talalay P. Anticarcinogenic activities of organic isothiocyanates: chemistry and mechanisms. Cancer Res 1994; 54:1976s-1981s.
- 19. Boone CW, Kelloff GJ, Malone WE. Identification of candidate cancer chemopreventive agents and their evaluation in animal models and human clinical trials: a review. Cancer Res 1990; 50:2-9.
- McDanell R, McLean AEM. Chemical and biological properties of Indole glucosinolates (glucobrassicins): a review. Food Chem. Toxicol 1988; 26:59-70.
- 21. Blot WJ, Li JY, Taylor PR, Guo W, Dawsey S, Wang GQ *et al.* Nutrition intervention trials in Linxian, China: Supplementation with specific vitamin/mineral combinations, cancer incidence, and disease-specific mortality in the general population. J. Natl. Cancer I. 1993; 85:1483-1492.
- Bjelakovic G, Nikolova D, Gluud LL, Simonetti RG, Gluud C. Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: Systematic review and meta-analysis. J. Am.

- Med.Assoc.2007; 297:842-857.
- Loria CM, Klag MJ, Caulfield LE, Whelton PK. Vitamin C status and mortality in US adults Am J Clin Nutr 2000; 72:139-145.
- 24. Frei B, Lawson S. Vitamin C and cancer revisited. Proc. Natl. Acad. Sci. USA 2008; 105:11037-11038.
- Li Y, Schellhorn HE. New developments and novel therapeutic perspectives for vitamin C J Nutr 2007a; 137:2171-2184.
- Kim SY, Yoon S, Kwon SM, Park KS, Lee-Kim YC. Kale Juice improves coronary artery disease risk factors in hyper cholesterolemic men Biomed Environ Sci 2008; 21:91-97.
- 27. Jahangir M, Kim HK, Choi YH, Verpoorte R. Health-affecting compounds in *Brassica*ceae. Compr. Rev. Food Sci Food Safety 2009; 8:31-43.
- Crozier A, Jaganath IB, Clifford MN. Phenols, polyphenols and tannins: An overview. In Plant Secondary Metabolites: Occurrence, Structure and Role in the Human Diet; Crozier, A., Clifford, M., Ashihara, H., Eds.; Blackwell: Oxford, UK, 2006, 1-24.
- Pereira DM, Valentao P, Pereira JA, Andrade PB. Phenolics: From Chemistry to Biology. Molecules 2009; 14:2202-2211.
- 30. http://www.todaysdietitian.com/newarchives/090313p70.s html [date accessed May 1, 2015].
- Podsedek A. Natural antioxidants and antioxidant capacity of *Brassica* vegetables: A review. Lwt-Food Sci. Technol 2007; 40:1-11.
- Podsedek A, Sosnowska D, Redzynia M, Anders B. Antioxidant capacity and content of *Brassica oleracea* dietary antioxidants. Int J Food Sci Technol 2006; 41:49-58
- Crozier A, Jaganath IB, Clifford MN. Dietary phenolics: Chemistry, bioavailability and effects on health *Nat* Prod Rep 2009; 26:1001-1043.
- Vallejo F, Tomas-Barberan FA, Garcia-Viguera C. Potential bioactive compounds in health promotion from broccoli cultivars grown in Spain. J Sci Food Agric 2002; 82:1293-1297.
- De Pascual-Teresa S, Moreno DA, Garcia-Viguera C. Flavanols and Anthocyaninsin Cardiovascular Health: A Review of Current Evidence. Int J Mol Sci 2010; 11:1679-1703
- 36. Kapusta-Duch J, Aneta Kopeć A, Piątkowska E, Borczak B, Leszczyńska T. The beneficial effects of *Brassica* vegetables on human health, Rocz Panstw Zakl Hig Nr 2012; 63(4):389-395.
- Plumb GW, Price KR, Rhodes MJC, Williamson G. Antioxidant properties of the major polyphenolic compounds in broccoli. Free Radical Res 1997; 27:429-435.
- 38. Cushnie TPT, Lamb AJ. Antimicrobial activity of flavonoids. Int. J. Antimicrob. Agents 2005; 26:343-356.
- Chu YH, Chang CL, Hsu HF. Flavonoid content of several vegetables and their antioxidant activity. J Sci Food Agric 2000; 80:561-566.
- Fukumoto LR Mazza G. Assessing antioxidant and pro oxidant activities of phenolic compounds. J. Agric. Food Chem 2000; 48:3597-3604.
- 41. Skandrani I, Limem I, Neffati A, Boubaker J, Ben Sghaier M, Bhouri W. Assessment of phenolic content, free radical scavenging capacity genotoxic and anti-genotoxic effect of aqueous extract prepared from Moricandia

- arvensis leaves. Food Chem Toxicol 2010; 48:710-715.
- 42. Ackland ML, Van de Waarsenburg S, Jones R. Synergistic antiproliferative action of the Flavonolsquercetin and kaempferol in cultured human cancer cell lines. *In Vivo* 2005; 19:69-76.
- Fresco P, Borges F, Marques MPM, Diniz C. The Anticancer Properties of DietaryPolyphenols and its Relation with Apoptosis. Curr. Phar Des 2010; 16:114-134.
- Lee J, Koo N, Min DB. Reactive Oxygen Species, Aging, and Antioxidative Nutraceuticals. Compr. Rev. Food Sci. Food Saf 2004; 3:21-33.
- Kusznierewicz B, Piasek A, Lewandowska J, Śmiechowska A, Bartoszek A. Anti-carcinogenic properties of white cabbage. Technologia Jakość 2007; 6(55): 20-34.
- Cohen JH, Kristal AR, Stanford JL. Fruit and vegetable intakes and prostate cancer risk. J. Natl Cancer Inst 2000; 92:61-68.
- Knekt P, Kumpulainen J, Jarvinen R, Rissanen H, Heliovaara M, Reunanen A. Flavonoids intake and risk of chronic diseases. Am. J Clin Nutr 2002; 76:560-568.
- 48. Yokozawa T, Kim HY, Cho EJ, Choi JS, Chung HY. Antioxidant Effects of isorhamnetin 3, 7-Di-O-β-d-glucopyranoside isolated from mustard leaf (*Brassica juncea*) in Rats with Streptozotocin-Induced Diabetes. J.Agr. Food Chem 2002; 50:5490-5495.
- Mart N, Mena P, C'anovas JA, Micol V, Saura D. Vitamin C and the role of citrus juices as functional food.Nat. Prod. Commun 2009; 4:677-70047.
- Hoey BM, Butler J. Therepai of oxidized amino acids byantioxidants. Biochim. Biophys. Acta 1984; 791:212-218.
- 51. Barja G, Lopez-Torres M, Perez-Campo R, Rojas C, Cadenas S, Prat J. Dietary vitamin C decreases endogenous protein oxidative damage, malon dialdehyde, and lipid peroxidation and maintains fatty acid unsaturation in the guinea pig liver. Free Radical Biol. Med. 1984; 17:105–115.
- Lutsenko EA, Carcamo JM, Golde DW. Vitamin C prevents DNA mutation induced by oxidative stress. J. Biol. Chem 2002; 277:16895-16899.
- 53. Fabre N, Poinsot V, Debrauwer L, Vigor C, Tulliez J, Fouraste I *et al.* Characterization of glucosinolates using electro spray ion trap and electro spray quadrupole time of flight mass. Phytochemical Analysis 2007; 18:306-319.
- Hecht SS. Chemoprevention of Cancer by Isothiocyanates, Modifiers of Carcinogen Metabolism. J. Nutr 1999; 129:768S-774S.
- 55. Jeong WS, Kim IW, Hu R, Kong AN. Modulatory properties of various natural chemopreventive agents on the activation of NF- kappa B signaling pathway. Pharmaceut Res 2004; 21:661-670.
- 56. Rose P, Won YK, Ong CN, Whiteman M. Betaphenylethyl and 8-methylsulphinyloctyl isothiocyanates, constituents of watercress, suppress LPS induced production of nitric oxide and prostaglandin E2 in RAW 264.7 macrophages. Nitric Oxide 2005; 12:237-243.
- 57. Fimognari C, Turrini E, Ferruzzi L, Lenzi M, Hrelia P. Natural isothiocyanates: Genotoxic potential versus chemoprevention. Mutat Res 2012; 750(2):107-131.
- Ian T, Johnson V. Glucosinolates in the human diet. Bioavailability and implications for human health. Phytochemistry Reviews 2002; 1:183-188.

- Zhao J, Kobori N, Aronowski J, Dash PK. Sulforaphane reduces infarct volume following focal cerebral ischemia in rodents. Neuroscience Letters 2006; 393(2-3):108-112.
- Ping Z, Liu W, Kang Z et al. Sulforaphane protects brains against hypoxic-ischemic injury through induction of Nrf2-dependent phase 2 enzyme. Brain Research 2010; 1343:178-185.
- Tarozzi A, Angeloni C, Malaguti M, Morroni F, Hrelia S, Hrelia P. Sulforaphane as a Potential Protective Phytochemical against Neurodegenerative Diseases, Hindawi Publishing Corporation Oxidative Medicine and Cellular Longevity Volume Article ID 415078, 10, 2013. pageshttp://dx.doi.org/10.1155/2013/415078.
- Maas AI, Stocchetti N, Bullock R. Moderate and severe traumatic brain injury in adults. The Lancet Neurology 2008; 7(8):728-741.
- Zhao J, Moore AN, Redell JB, Dash PK. Enhancing expression of Nrf2-driven genes protects the blood-brain barrier after brain injury. Journal of Neuroscience 2007; 27(38):10240-10248.
- Zhao J, Moore AN, Clifton GL, Dash PK. Sulforaphane enhances aquaporin-4 expression and decreases cerebral edema following traumatic brain injury. Journal of Neuroscience Research 2005; 82(4):499-506.
- 65. Manley GT, Binder DK, Papadopoulos MC, Verkman AS. New insights into water transport and edema in the central nervous system from phenotype analysis of aquaporin-4 null mice. Neuroscience 2004; 129(4):983-991.
- 66. Tannenberg RK, Scott HL, Tannenberg AEG, Dodd PR. Selective loss of synaptic proteins in Alzheimer's disease: evidence for an increased severity with APOE ε4. Neurochemistry International 2006; 49(7):631-639.
- 67. Kwak MK, Cho JM, Huang B, Shin S, Kensler TW. Role of increased expression of the proteasome in the protective effects of sulforaphane against hydrogen peroxide mediated cytotoxicity in murine neuroblastoma cells. Free Radical Biology and Medicine 2007; 43(5):809-817.
- Park HM, Kim JA, Kwak MK. Protection against amyloid beta cytotoxicity by sulforaphane: role of the proteasome. Archives of Pharmacal Research 2009; 32(1):109-115.
- Dawson TM, Dawson VL. Molecular pathways of neurodegeneration in Parkinson's disease. Science 2003; 302(5646):819-822.
- Calne S, Schoenberg B, Martin W, Uitti RJ, Spencer P, Calne DB. Familial Parkinson's disease: possible role of environmental factors. Canadian Journal of Neurological Sciences 1987; 14(3):303-305.
- 71. Schoenberg BS. Environmental risk factors for Parkinson's disease: the epidemiologic evidence. Canadian Journal of Neurological Sciences 1987; 14(3):407-413.
- 72. Ktada T, Asakawa S, Hattori N *et al.* Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. Nature 1998; 392(6676):605-608.
- Polymeropoulos MH, Lavedan C, Leroy E et al. Mutationin the α-synuclein gene identified in families with Parkinson's disease. Science 1997; 276(5321):2045-2047.
- 74. Kruger R, Kuhn W, Muller T *et al.* Ala30Pro mutation in the gene encoding α-synuclein in Parkinson's disease. Nature Genetics 1998; 18(2):106-108.
- 75. Zarranz JJ, Alegre J, Gomez-Esteban JC *et al.* The new mutation, E46K, of α-synuclein causes Parkinson and

- Lewybody dementia. Annals of Neurology 2004; 55(2):164-173.
- 76. Spillantini MG, Schmidt ML, Lee VM, Trojanowski JQ, Jakes R, Goedert M. α-synuclein in Lewy bodies. Nature 1997; 388(6645):839-840.
- Cuervo AM, Stafanis L, Fredenburg R, Lansbury PT, Sulzer D. Impaired degradation of mutant αsynucleinbychaperone-mediated autophagy. Science 2004; 305(5688):1292-1295.
- Yoon NS, Cho Y, Lee SY, Choi HJ, Hwang O. Inactivation of aconitase by tetrahydrobiopterin in Dargic cells: relevance to PD. Experimental Neurobiology 2010; 19(1):23-29.
- 79. Han JM, Lee YJ, Lee SY *et al.* Protective effect of sulforaphane against dopaminergic cell death. Journal of Pharmacology and Experimental Therapeutics 2007; 321(1):249-256.
- Ferrarini L, Pellegrini N, Mazzeo T, Miglio C, Galati S, Milano F et al. Anti-proliferative activity and chemoprotective effects towards DNA oxidative damage of fresh and cooked *Brassica*ceae. Brit J Nutr 2011; 17:1-9
- Thomson CA, Dickinson S, Bowden T. Cruciferous vegetables, isothiocyanates, indoles, and cancer prevention. Nutrition and Health: Bioactive Compounds and Cancer. Springer Science+Business Media, LLC 2010, 535-566.
- 82. Jakubikova J, Cervi D, Ooi M, Kim K, Nahar S, Klippel S *et al.* Anti-tumor activity and signaling events triggered by the isothiocyanates, sulforaphane and phenethyl isothiocyanate, in multiple myeloma. Haematol 2011; 96(8):1170-9.
- 83. Ackland ML, Van de Waarsenburg S, Jones R. Synergistic antiproliferative action of the flavonols quercetin and kaempferol in cultured human cancer cell lines. *In Vivo* 2005; 19:69-76.
- 84. Fresco P, Borges F, Marques MPM, Diniz C. The Anticancer Properties of Dietary Polyphenols and its Relation with Apoptosis. Curr Pharm Des 2010; 16:114-134
- 85. Llorach R, Espin JC, Tomas-Barberan FA, Ferreres F. Valorization of cauliflower (*Brassica oleracea* L. var. *botrytis*) by-products as a source of antioxidant phenolics. J. Agric. Food Chem 2003; 51:2181-2187.
- 86. Kirsh VA, Peters U, Mayne ST, Subar AF, Chatterjee N, Johnson CC *et al.* Prospective Study of Fruit and Vegetable Intake and Risk of Prostate Cancer. Journal of the National Cancer Institute 2007; 99(15):1200-1209.
- 87. Traka M. Broccoli Consumption Interferes with Prostate Cancer Progression: Mechanisms of Action. Acta Horticulturae 2010; 867(5):19 -25.
- 88. Chung FL, Conaway CC, Rao CV, Reddy BS. Chemoprevention of colonic aberrant crypt foci in Fischer rats by sulforaphane and phenethyl isothiocyanate. Carcinogenesis 2000; 21:2287-2291.
- 89. Smith TK, Lund EK, Johnson IT. Inhibition of dimethylhydrazine-induced aberrant crypt foci and induction of apoptosis in rat colon following oral administration of the glucosinolate sinigrin. Carcinogenesis 1998; 19:267-273.
- Bird RP. Role of aberrant crypt foci in understanding the pathogenesis of colon cancer. Cancer Lett. 1995; 93:55-71
- 91. Smith TK, Clarke R, Scott J, Johnson IT. Raw Brussels

sprouts block mitosis in colorectal cancer cells (HT29) and induce apoptosis in rat colonic mucosal crypts *in vivo*. In: Johnson IT & Fenwick GR (eds) Dietary Anticarcinogens and Antimutagens: Chemical and Biological Aspects. Royal Society of Chemistry, Cambridge, 2000.