



ISSN 2278- 4136

ZDB-Number: 2668735-5

IC Journal No: 8192

Volume 1 Issue 6

Online Available at [www.phytojournal.com](http://www.phytojournal.com)

## Journal of Pharmacognosy and Phytochemistry

# Molecular Target-Oriented Phytochemical Database and Its Application to the Network Analysis of Action Mechanisms of Herbal Medicines

Hitoshi Kojo<sup>\*1</sup>, Yukihiro Eguchi<sup>1</sup>, Toshikazu Miyagishima<sup>1</sup>, Kimiko Makino<sup>1,2</sup> and Hiroshi Terada<sup>1,2</sup>

1. Systems PharmaSciences Research Organization, Japan  
[E-mail: [kojoh@systemspharma.org](mailto:kojoh@systemspharma.org)]
2. Faculty of Pharmaceutical Sciences, Tokyo University of Sciences, Japan

---

Kampo medicines, the Japanese adaptation of traditional Chinese medicines, are formed by combining several herbs containing multiple phytochemicals. The considerable ambiguity of pharmacological profiles of Kampo medicines is expected to be clarified by identifying the molecular targets of constituent phytochemicals and analyzing the combined effects of the phytochemicals on the pharmacological pathways formed by those targets. To facilitate this line of study, we constructed paired databases named PhytodamaTarget and PhytodamaTaxon DBs which treat molecular targets of phytochemicals and constituent phytochemicals of plant taxa, respectively, by utilizing information from the literature. We then used the databases to explore possible mechanisms of synergism in analgesic activity between *Glycyrrhiza glabra* and *Paeonia lactiflora*

---

**Keyword:** Phytochemicals, Database, Molecular Target, Kampo Medicines, Synergism, Analgesic

### 1. Introduction

Kampo and traditional Chinese medicines are characterized by a complex composition with multiple pharmacologically active phytochemicals<sup>[1]</sup>. A tremendous number of these medicines have been developed by combining various specified herbs to achieve the greatest possible pharmacological effect<sup>[2]</sup>. However, most of the mechanisms of pharmacological action of Kampo medicines have yet to be fully elucidated at a molecular level<sup>[3]</sup>. To elucidate these molecular mechanisms, it is necessary to identify the molecular targets of the herbal ingredients and to clarify the synergism or antagonism of molecular

actions. The purpose of this study is to construct a phytochemical database that facilitates identification of the molecular targets of phytochemicals contained in Kampo formulations and to examine its usefulness for exploring the molecular mechanisms behind the medicines actions.

### 2. Methods

#### 2.1 Database generation

The data for molecular targets and the taxonomy of producers of phytochemicals were compiled from both primary and secondary sources, particularly Gideon Polya's Biochemical Targets of Plant Bioactive Compounds<sup>[4]</sup>. The chemical

information on phytochemicals was also obtained from primary and secondary sources, the latter of which included databases such as PubChem (<http://pubchem.ncbi.nlm.nih.gov/>) and NikkajiWeb ([http://nikkajiweb.jst.go.jp/nikkaji\\_web/pages/top\\_e.html](http://nikkajiweb.jst.go.jp/nikkaji_web/pages/top_e.html)). PhytodamaTarget DB and PhytodamaTaxon DB were constructed by using Microsoft Office Access 2003.

## 2.2 Graphical presentation of relationship between phytochemicals and their molecular targets

CellDesigner 4.2, a tool for modeling biochemical networks with a graphical user interface developed by H. Kitano *et al.*<sup>[5]</sup>, was used to present graphically the relationship between phytochemicals and their molecular targets.

## 2.3 Screening of molecular targets involved in analgesic activity

A list of 371 genes involved in pain was obtained from the database PainGenesdb (<http://www.jbldesign.com/jmogil/enter.html>). The molecular targets of phytochemicals involved in analgesic activity were identified among the targets of either *Glycyrrhiza globra* or

*Paeonia lactiflora* by checking whether they were included in the pain gene list.

## 3. Results and Discussion

### 3.1 Construction of PhytodamaTarget and PhytodamaTaxon Databases

We constructed two databases named PhytodamaTarget and PhytodamaTaxon DBs, the former covering information on molecular targets of phytochemicals together with their chemical properties, and the latter providing information on the taxonomy of the producers of phytochemicals together with herbal information. The basic information in PhytodamaTarget DB consists of phytochemical names, phytochemical synonyms, target names, target subfamilies, target categories, modes of action, IC<sub>50</sub>/EC<sub>50</sub>, K<sub>d</sub>/K<sub>i</sub>, chemical structure, chemical formulae, smiles strings, molecular weights, biosynthetic pathways, links to other databases such as PubChem, KEGG (<http://www.genome.jp/kegg/>) and GeneCards (<http://www.genecards.org/>) and references (Fig.1) whereas that in PhytodamaTaxon DB consists of taxonomic names including genus, family, order, clade, Japanese, Chinese and English herb names, and phytochemical names (Fig.2).

Fig.1. Representative form of PhytodamaTarget DB.

Phytochemical_ID	Phytochemical_name	Target_ID	Target_name	Mode of action	IC50/EC50 (uM)	Kd/Ki (uM)	Remark
C00645	berbamine	T08700	calmodulin	inhibitor			
C00645	berbamine	T02900	muscarinic acetylcholine receptor	antagonist		0.2	
C00645	berbamine	T02000	voltage-gated Ca <sup>2+</sup>	inhibitor			

Fig. 2. Representative form of PhytodamaTaxon DB.

The screenshot displays the PhytodamaTaxon DB interface for the genus *Curcuma*. The top section contains taxonomic details:

- G\_ID:** G00391
- Genus:** Curcuma
- Family:** Zingiberaceae
- Order:** Zingiberales
- Clade:** Commelinids
- 科 (Family):** ショウガ科
- Herb name (Japanese):** ウコン, ウコン, 秋ウコン, 紫ウコン, ムラサキガジュツ
- Herb name (Chinese):** 郁金 (Curcuma wenyujin), 姜黄 (Curcuma longa), 莪朮 (Curcuma aeruginosa)
- Western herb name (Japanese):** ウコン
- Western herb name (English):** Turmeric (Curcuma longa)

The bottom section, titled "taxon\_phytochemical", shows a list of phytochemicals associated with the genus:

G_ID	Genus	phytochemical ID	phytochemical
G00391	Curcuma	C00176	1,8-cineole
G00391	Curcuma	C00517	aerugioidiol
G00391	Curcuma	C00882	curcumenol
G00391	Curcuma	C00883	curcumenone
G00391	Curcuma	C00884	curcumin
G00391	Curcuma	C00885	curcumin I

The data set used in the classification of targets: target category, target name and target subfamily designates pharmacologically distinct groups such as G protein-coupled receptors, gene product families such as receptor families, and gene product subfamilies such as receptor subtypes, respectively. The plant classification was performed according to the APG III system<sup>[6]</sup>.

Registered numbers of phytochemicals, targets, plant genres and herbs in the databases are 2408, 210, 1345 and 159, respectively.

### 3.2 Retrieval of Targets of Herbs

Identification of molecular targets of phytochemicals contained in herbs is helpful in elucidating the pharmacological effect of herbs. This identification is easily accomplished by combined use of PhytodamaTarget and PhytodamaTaxon DBs. Namely, data on plant genus can be obtained instantly together with that on its constituent phytochemicals and their targets by using the "subform" and "query" functions of Microsoft Access (Fig. 3).

Fig. 3. Presentation of phytochemicals and their targets corresponding to a plant genus record.

The screenshot displays the PhytodamaTarget interface for the genus *Glycyrrhiza*. The top section contains taxonomic details:

- G\_ID:** G00579
- Clade:** Eurosids I
- Order:** Fabales
- Family:** Fabaceae
- Genus:** Glycyrrhiza
- 科 (Family):** マメ科
- Herb name (Japanese):** カンゾウ
- Herb name (Chinese):** 甘草 (Glycyrrhiza uralensis)
- Western herb name (Japanese):** カンゾウ
- Western herb name (English):** Liquorice (Glycyrrhiza glabra)

The bottom section, titled "phytochemical\_taxon\_target", shows a list of phytochemicals and their targets:

Phytoche	Phytochemical name	Target ID	Target category	Target name	Target subfamily name	Mode of [IC50/EC50] Kd/Ki
C00204	18- $\alpha$ -glycyrrhetic acid	T09100	signal-regulated prot protein kinases(PKA, CDPK, CDK, PKC, PKI)	PKA, PKC	inhibitor	6(PKA), 1
C00205	18- $\beta$ -glycyrrhetic acid	T13300	cytosolic hormone re 17- $\beta$ -hydroxysteroid oxidoreductase		inhibitor	30
C00205	18- $\beta$ -glycyrrhetic acid	T13100	cytosolic hormone re estrogen receptor		agonist	0.9
C00205	18- $\beta$ -glycyrrhetic acid	T15100	digestion and metabi chymotrypsin (CHY), trypsin (TRY), elastase	elastase	inhibitor	
C00205	18- $\beta$ -glycyrrhetic acid	T05800	G protein-coupled re glucagon receptor		?	
C00205	18- $\beta$ -glycyrrhetic acid	T01300	ion pumps, trans- at Na,K-ATPase		inhibitor	
C00205	18- $\beta$ -glycyrrhetic acid	T09200	signal-regulated prot PKC		inhibitor	
C00205	18- $\beta$ -glycyrrhetic acid	T12900	cytosolic hormone re 11- $\beta$ -hydroxysteroid dehydrogenase		inhibitor	
C00205	18- $\beta$ -glycyrrhetic acid	T12700	cytosolic hormone re androgen transport-steroid binding globulin		inhibitor	0.5
C00205	18- $\beta$ -glycyrrhetic acid	T12800	cytosolic hormone re corticosteroid receptors		antagonist	0.004
C00205	18- $\beta$ -glycyrrhetic acid	T13000	cytosolic hormone re cortisol transport-cortisol binding globulin		inhibitor	10
C00205	18- $\beta$ -glycyrrhetic acid	T17500	digestion and metabi fatty acid desaturase		inhibitor	~0.01
C00205	18- $\beta$ -glycyrrhetic acid	T18800	digestion and metabi PLA2		binds PL	
C00205	18- $\beta$ -glycyrrhetic acid	T09100	signal-regulated prot protein kinases(PKA, CDPK, CDK, PKC, PKI)	PKA, PKC	inhibitor	6(PKA), 1
C00205	18- $\beta$ -glycyrrhetic acid	T12800	cytosolic hormone re corticosteroid receptors		antagonist	0.002
C00276	3,4-dihydroxychalcone	T15500	digestion and metabi oxidative phosphorylation		uncouple	
C00309	3-monoacyloxy-1-glycyrrhetic acid	T12900	cytosolic hormone re 11- $\beta$ -hydroxysteroid dehydrogenase		inhibitor	
C00309	3-monoacyloxy-1-glycyrrhetic acid	T13300	cytosolic hormone re 17- $\beta$ -hydroxysteroid oxidoreductase		inhibitor	16

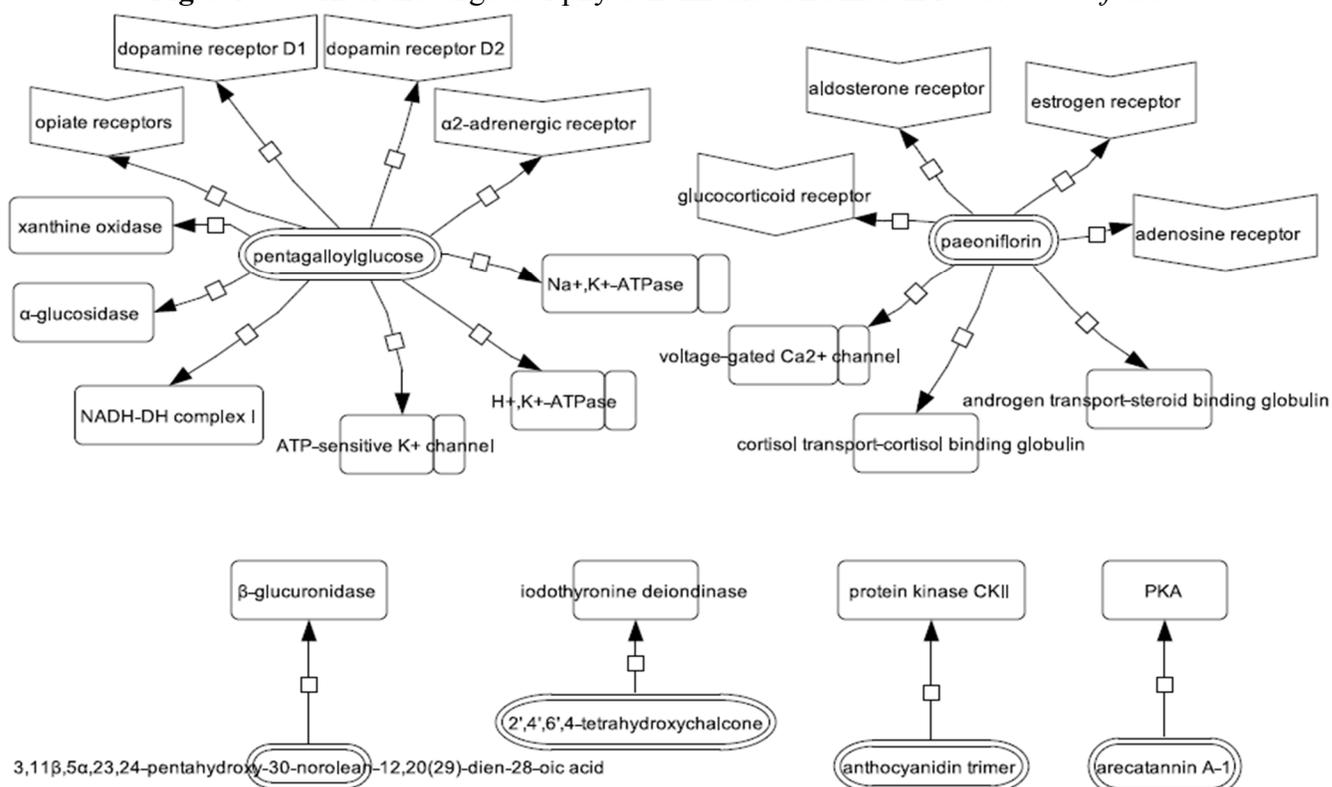


Meanwhile, other mechanisms of the analgesic activity of *Shakuyakukanzoto* and its constituents have been reported. Omiya *et al.* found that *Shakuyakukanzoto* showed antinociceptive activity in diabetic mice and assumed the activity to be caused by activation of the descending noradrenergic neurons<sup>[10]</sup>. Liu *et al.* ascribed the antinociceptive effect of paeoniflorin in mice to its ability to increase the binding and antinociceptive effect of an adenosine A1 agonist by binding with A1 receptors<sup>[11]</sup>. According to our database, *Paeonia lactiflora* contains a constituent with  $\alpha$ 2-adrenergic agonist activity, namely  $\beta$ -1,2,3,4, 6-penta-O-galloyl-D-glucose (PGG), together with paeoniflorin. Further studies are necessary to conclude whether PGG, paeoniflorin or same as yet unidentified ingredient is responsible for the analgesic activity of *Shakuyakukanzoto* in diabetic mice. The molecular mechanism behind the synergistic analgesic effect of *Shakuyakukanzoto* remains largely unclear despite extensive studies. To elucidate it, it is necessary to clarify the

interactions among constituents whose molecular targets are involved in the analgesia. Accordingly, the molecular targets of phytochemicals contained in *Glycyrrhiza globra* and *Paeonia lactiflora* were first retrieved by combined use of our databases. The number of hits for phytochemicals in *Glycyrrhiza globra* and *Paeonia lactiflora* was 29 and 6, respectively and the total number of hits for the molecular targets of these phytochemicals was 28 and 21, respectively (Fig.4 and Fig.5). The number of molecular targets common to both genres was 6. Furthermore, some molecular targets were shared by multiple phytochemicals within the genus (Fig.4).

The symbols used to represent molecules and actions are as follows; ovoids, concave hexagons, and divided and simple round-cornered squares represent phytochemicals, receptors, ion channels and generic proteins, respectively. Arrows represent actions of phytochemicals against targets.

**Fig.5.** Network of the targets of phytochemicals contained in *Paeonia lactiflora*.

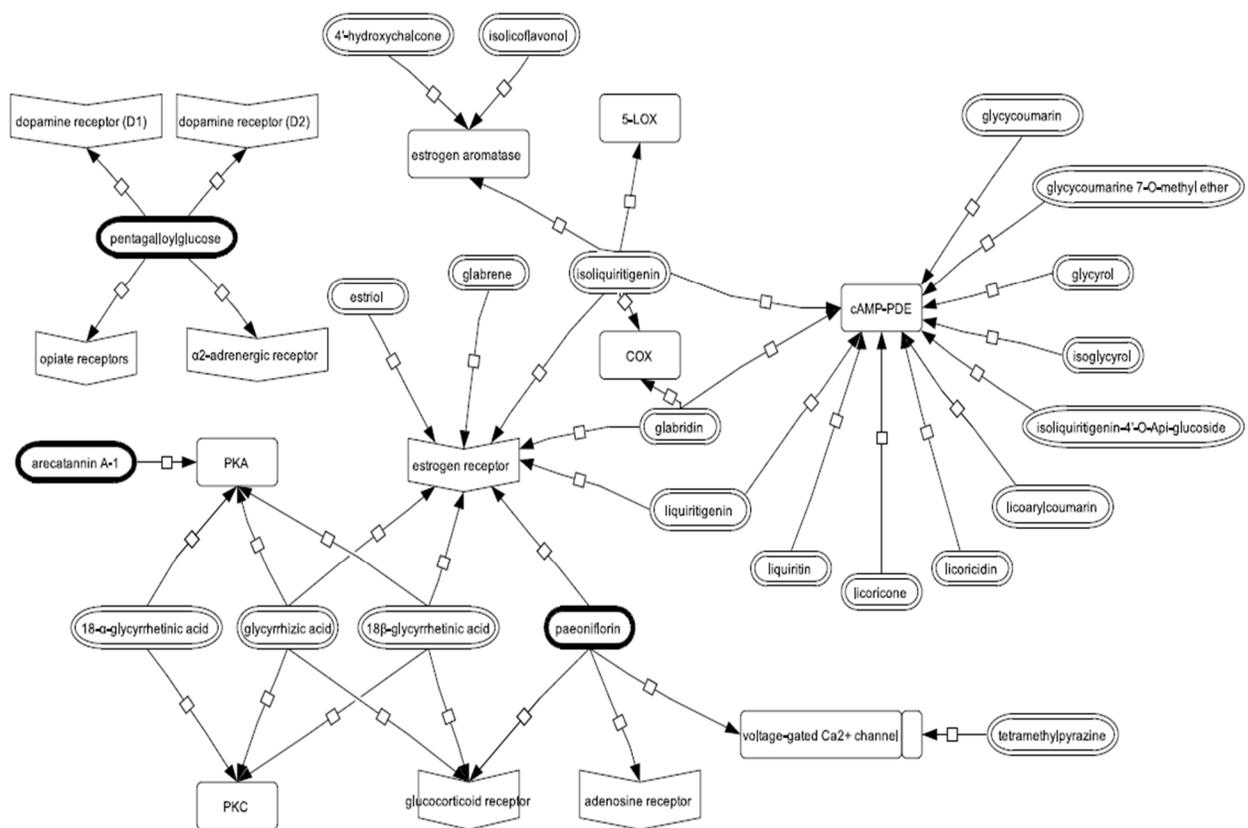


The symbols used are explained in the legend of Fig. 4.

To identify the molecular targets involved in analgesic activity among those listed, those coinciding with the gene products registered in the pain genes database<sup>[12]</sup> were screened. Eight out of 28 targets for *Glycyrrhiza gloabra* and 8 of 21 targets for *Paeonia lactiflora* were hit. They are cyclooxygenase, estrogen aromatase, the estrogen receptor, 5-lipoxygenase, protein kinase A, protein kinase C, cAMP phosphodiesterase and voltage-gated Ca<sup>2+</sup> channels for *Glycyrrhiza gloabra* and the adenosine receptor, dopamine receptors, the estrogen receptor, opiate receptors, protein kinase A, voltage-gated Ca<sup>2+</sup> channels and the  $\alpha$ 2-adrenergic receptor for *Paeonia lactiflora*. Although the glucocorticoid receptor, a target for

both genuses, is missing in the pain genes database, it may be added as a pain gene, referring to the literature<sup>[13,14]</sup>. Among the targets identified as pain-related, cyclooxygenase, 5-lipoxygenase and estrogen aromatase are involved in the biosynthesis of algesic and analgesic substances<sup>[15,16]</sup>, the adenosine, estrogen and glucocorticoid receptors are receptors for algesic or analgesic substances<sup>[13,14,17,18]</sup>, the  $\alpha$ 2-adrenergic, dopamine and opiate receptors are receptors for neurotransmitters<sup>[19-21]</sup>, protein kinases A, C and cAMP phosphodiesterase are involved in postreceptor signaling pathways<sup>[22]</sup> and voltage-gated Ca<sup>2+</sup> channels are involved in neural transmission including regulation of the release of neurotransmitters<sup>[23]</sup>. Fig. 6 summarizes the interaction between the herbal ingredients and their molecular targets involved in analgesia.

**Fig.6.** Network of the pain-involved targets of phytochemicals contained in either *Glycyrrhiza gloabra* or *Paeonia lactiflora*.



The symbols are the same as those in Fig.4 except that ovoids with double outlines represent phytochemicals contained in *Glycyrrhiza glabra* whereas ovoids with a thick outline represent those in *Paeonia lactiflora*.

The network of herbal ingredients and their pain-involved molecular targets shows that there are hubs, targets highly connected with ingredients, such as cAMP phosphodiesterase and the estrogen receptor, suggesting their possible greater roles for analgesic activity of the herbal medicine. Cyclic AMP phosphodiesterase catabolizes cAMP which is a pain-mediating second messenger.<sup>[24]</sup> Hence, the inhibitory activity of phytochemicals against cAMP phosphodiesterase would lead to pain enhancement. However, contrary to this speculation, cAMP phosphodiesterase inhibitors were shown to elevate nociceptive thresholds in the central nervous system by increasing antinociceptive natural epoxy-fatty acids<sup>[25]</sup>. The estrogen receptor was shown to be involved in lowering nociceptive thresholds by using the estrogen receptor knockout female mice although its mechanism has yet to be deciphered<sup>[26]</sup>. The phytochemicals would exert analgesic activity not by their agonistic activity but by their antagonistic activity against the estrogen receptor in the presence of endogenous estrogen<sup>[27]</sup>. Meanwhile, among targets shared by multiple components, those shared by components from different herbs, such as voltage-gated Ca<sup>2+</sup> channels, the glucocorticoid receptor, the estrogen receptor and protein kinase A, are particularly noteworthy since one possible mechanism of synergism between agonists/antagonists or inhibitors is differential activities against a shared target. However, targets other than those mentioned above could be involved in the synergism, because simultaneous activities against targets closely related to analgesic activity is another possible mechanism of synergism. Though the ideas discussed above need to be examined further in wet experiments, the information provided by our database should help to elucidate the molecular

mechanism of synergism in the analgesic activity of *Shakuyakukanzoto*. Furthermore, PhytodamaTarget and PhytodamaTaxon DBs should prove useful for analyzing the synergism or antagonism of pharmacological activities among herbal components of Kampo and other traditional medicines.

#### 4. Conclusion

PhytodamaTarget and PhytodamaTaxon DBs were shown to facilitate identification of the molecular targets of phytochemicals contained in Kampo formulations and to be useful for exploring the molecular mechanisms of the synergistic pharmacological actions of these medicines.

#### 5. References

1. Ehrman TE, Barlow DJ, and Hylands PJ. Phytochemical databases of Chinese herbal constituents and bioactive plant compounds with known target specificities. *J. Chem. Inf. Model.* 2007; 47, 254-263.
2. Yu F, Takahashi T, Moriya J, Kawaura K, Yamakawa J, Kusaka K et al. Traditional Chinese medicine and Kampo: a review from the distant past for the future. *J. Int. Med. Res.* 2006; 3, 231-239.
3. Uezono Y, Miyano K, Suzuki M, Sudo Y, Shiraishi S and Terawaki K. A review of traditional Japanese medicines and their potential mechanism of action. *Curr Pharm Des.* 2012 May 23 Epub ahead of print
4. Polya G. Biochemical Targets of Plant Bioactive Compounds: A Pharmacological Reference Guide to Sites of Action and Biological Effects. Taylor & Francis, London, 2003.
5. Funahashi A, Tanimura N, Morohashi M and Kitano H. CellDesigner: a process diagram editor for gene-regulatory and biochemical networks. *BIOSILICO* 2003; 1, 159-162.
6. The Angiosperm Phylogeny Group. An update of the angiosperm phylogeny group classification for the orders and families of flowering plants: APG III. *Bot. J. Linn. Soc.* 2009; 161, 105-121.
7. Hidaka T, Shima T, Nagira K, Ieki M, Nakamura T, Aono, Y et al. Herbal medicine Shakuyaku-kanzo-to

- reduces paclitaxel-induced painful peripheral neuropathy in mice. *Eur. J. Pain.* 2009; 13, 22-27.
8. Kimura M, Kimura I and Nojima H. Depolarizing neuromuscular blocking action induced by electropharmacological coupling in the combined effect Paeoniflorin and Glycyrrhizin. *Japan. J. Pharmacol.* 1985; 37, 395-399.
9. Satoh H and Tsuro K. Pharmacological modulation by Shakyakukanzoto (Shao-Yao-Gan-Cao-Tang) and the ingredients in rat intestinal smooth muscle. *Chinese Medicine* 2011; 2, 62-70.
10. Omiya Y, Suzuki Y, Yuzurihara M, Murata M, Aburada M, Kase Y et al. Antinociceptive effect of Shakyakukanzoto, a Kampo medicine, in diabetic mice. *J. Pharmacol. Sci.* 2005; 99, 373-380.
11. Liu DZ, Zhao FL, Liu J, Ji XQ, Ye Y and Zhu XZ. Potentiation of adenosine A1 receptor agonist CPA-induced antinociception by paeoniflorin in mice. *Biol. Pharm. Bull.* 2006; 29, 1630-1633.
12. Lacroix-Fralish ML, Ledoux JB and Mogil JS. The Pain Genes Database: An interactive web browser of pain-related transgenic knockout studies. *Pain* 2007; 131, 3.e1-3.e4.
13. Chrousos GP and Kino T. Glucocorticoid signaling in the cell: expanding clinical implications to complex human behavioral and somatic disorders. *Ann. N. Y. Acad. Sci.* 2009; 1179, 153-166.
14. Aasboe V, Raeder JC and Groegaard B. Betamethasone reduces postoperative pain and nausea after ambulatory surgery. *Anesth. Analg.* 1998; 87, 319-323.
15. Steinmeyer J. Pharmacological basis for the therapy of pain and inflammation with nonsteroidal anti-inflammatory drugs. *Arthritis. Res.* 2000; 2, 379-385.
16. Smith YR and Sto CS. Pronociceptive and antinociceptive effects of estradiol through endogenous 26pioid neurotransmission in women. *J. Neurosci.* 2006; 26, 5777-5785.
17. Jacobson KA. Introduction to adenosine receptors as therapeutic targets. *Handb. Exp. Pharmacol.* 2009; 193, 1-24.
18. Gupta S, McCarson KE, Welch KM and Berman NE. Mechanisms of pain modulation by sex hormones in migraine. *Headache.* 2011, 51, 905-922.
19. Fairbanks CA, Stone LS and Wilcox GL. Pharmacological profiles of alpha 2 adrenergic receptor agonists identified using genetically altered mice and isobolographic analysis. *Pharmacol. Ther.* 2009; 123, 224-238.
20. Jarcho JM, Mayer EA, Jiang ZK, Feier NA and London ED. Pain, affective symptoms, and cognitive deficits in patients with cerebral dopamine dysfunction. *Pain* 2012; 153, 744-754.
21. Simon EJ, Hiller JM. The opiate receptors. *Annu. Rev. Pharmacol. Toxicol.* 1978; 18, 371-394.
22. Cheng JK and Ji RR. Intracellular signaling in primary sensory neurons and persistent pain. *Neurochem. Res.* 2008; 33, 1970-1978.
23. Perret D and Luo ZD. Targeting voltage-gated calcium channels for neuropathic pain management. *Neurotherapeutics.* 2009; 6, 679-692.
24. Taiwo YO, Bjerknes LK, Goetzl EJ and Levine JD. Mediation of primary afferent peripheral hyperalgesia by the cAMP second messenger system. *Neuroscience* 1989 32, 577-580.
25. Inceoglu B, Wagner K, Schebb NH, Morisseau C, Jinks SL, Ulu A, Hegedus C, Rose T, Brosnan R and Hammock BD. Analgesia mediated by soluble epoxide hydrolase inhibitors is dependent on cAMP. *Proc Natl Acad Sci USA.* 2011, 108, 5093-5097.
26. Li L, Fan X, Warner M, Xu XJ, Gustafsson JA, Wiesenfeld-Hallin Z. Ablation of estrogen receptor alpha or beta eliminates sex differences in mechanical pain threshold in normal and inflamed mice. *Pain* 2009, 143, 37-40.
27. Collins BM, McLachlan JA, Arnold SF. The estrogenic and antiestrogenic activities of phytochemicals with the human estrogen receptor expressed in yeast. *Steroids* 1997, 62, 365-372.