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A.N.M. Alamgir

Department of Botany, University of
Chittagong, Chittagong-4331,
Bangladesh

Minhajur Rahman

Department of Botany, University of
Chittagong, Chittagong-4331,
Bangladesh

Ataur Rahman

Department of Botany, University of
Chittagong, Chittagong-4331,
Bangladesh

Phytochemical Characteristics, Antimitotic, Cytotoxic and Antiinflammatory Activities of *Coccinia grandis* (L.) J. Voigt

A.N.M. Alamgir, Minhajur Rahman, Ataur Rahman

ABSTRACT

Coccinia grandis (L.) J. Voigt, grows in the wild all over Bangladesh, is known to exhibit certain therapeutic properties. In the present work, leaf extract of *C. grandis* was considered for qualitative assessment of its secondary metabolite contents such as alkaloids, flavonoids, glycosides, saponins, sterol and tannins. Antimitotic, cytotoxic and antitumor activities of the ethanolic extract of leaf were also determined. Leaf extract showed strong inhibitory effect on seed germination, seedling's root growth of wheat and brine shrimp lethality with LC₅₀ at 24.20 µg/ml. Antitumor assay showed inhibition of tumor induction on potato discs due to *Agrobacterium tumefaciens* by 66% at 1000ppm without any lethal activity against the bacterial agent. Results of anticarcinogenic activities of the leaf extract were discussed in relation to its secondary metabolites content.

Keywords: *Coccinia grandis*, Ethanolic Extract, Secondary Metabolites, Anticarcinogenic Activities.

1. Introduction

Traditional medicinal plants have been recognized for their therapeutic benefits for centuries. However, there is still lack of evidence for the clarification of their typical mechanisms of action. *Coccinia grandis* (L.) J. Voigt or Ivy gourd of Cucurbitaceae, an indigenous plant to Bangladesh and other South Asian countries, grows luxuriantly as tendril climber all over Bangladesh in the wild. *Coccinia grandis* leaf is traditionally used in anorexia, cough, diabetic, wounds and biliary-hepatic disorders. It has been reported to have antiinflammatory, antioxidant, antimutagenic, antimicrobial, antiulcer, hepatoprotective, expectorants and analgesic activities [1]. But the systemic effects and mechanisms of its anticancer properties like antimitotic, cytotoxic and antitumor activities have not been investigated in relation to its secondary metabolites content. Therefore, we aimed to investigate these properties in the present work along with the qualitative assessment of its secondary metabolites.

2. Materials and Method

Leaf extract of *Coccinia grandis* (L.) J. Voigt was used in the present work. For extraction, the collected tender leaf sample from a naturally grown plant was cleaned, chopped, air dried at room temperature and finally ground into coarse powder. About 100 g powder was macerated with ethanol (1:5) in a sealed container for 5 days at room temperature with occasional shaking. Extract was filtered through Whatman No.1 filter paper and evaporated to dryness under vacuum below 50 °C to get about 3 g blackish extract. The extract thus prepared was kept at 4 °C for future use. Methods of Cromwell [2], Aplin and Cannon [3], Wall *et al.* [4], Farnsworth [5], Bhattacharjee and Das [6] were followed for qualitative assessment of different secondary metabolites. Alkaloid contents were assessed with the help of Dragendorff (D), Wagner (W), Mayer (M), Hager (H) and Tannic acid (T) reagents. The relative abundance of secondary metabolite contents in the extract was expressed by – or + signs, ranging up to 4+, signifying its abundance in degrees. Antimitotic activity of the leaf extract was determined according to Turker and Camper [7] using wheat seeds at germination and seedling growth stage. Cytotoxicity of the leaf extract was determined according to Meyer *et al.* [8] using brine shrimp nauplii and antiinflammatory activity on potato discs due to *Agrobacterium tumefaciens* was assessed according to Galsky *et al.* [9] using ethanol extract of leaf. Each set of experiment was replicated thrice.

Correspondence:

A.N.M. Alamgir

Department of Botany, University of
Chittagong, Chittagong-4331,
Bangladesh
Email: alamgiranm@yahoo.com

3. Result and Discussion

The curative value of medicinal herbs largely depends on their secondary metabolites, especially alkaloids, terpenoids and phenolic compounds. Results given in Table 1 show that alkaloid

test by all five reagents gave positive result in the order of 2+ to 3+ signifying its presence in the leaf in substantial quantity. Among the rests, all metabolites except resin showed positive result.

Table 1: Qualitative assessment of different secondary metabolites in *C. grandis* leaf.

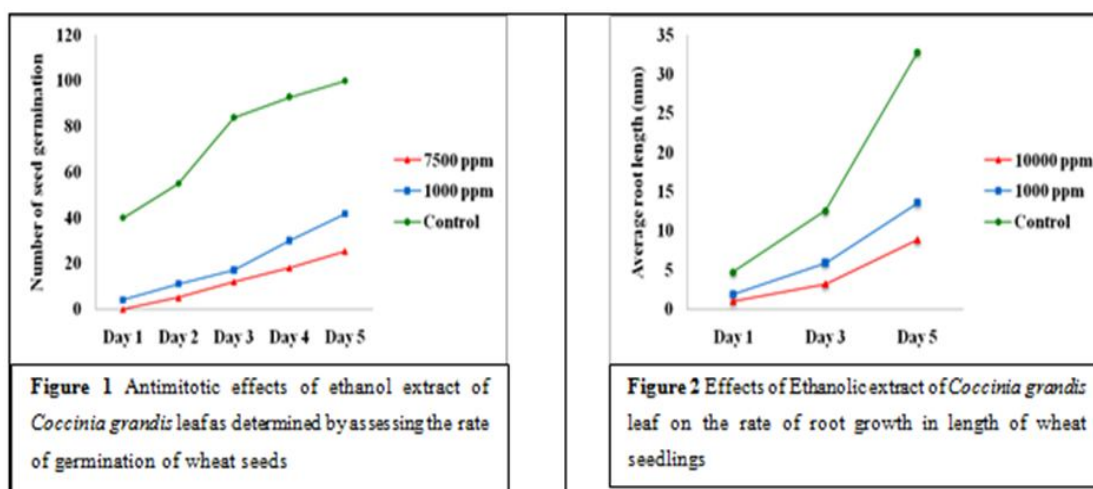
Secondary metabolites										
Alkaloid					Other secondary metabolites					
Reagents used					Flavonoid	Glycoside	Resin	Saponin	Sterol	Tanin
D	W	M	H	T						
3+	3+	3+	2+	3+	+	+	-	+	+	+

The occurrence of different secondary metabolites suggests a wide range of biological application of the plant [10, 11, 12]. Several alkaloids like vinblastine, vincristine, camptothecin, taxol etc. are successfully employed in cancer treatment [13]. Kunle and Egharevba [14] suggested to consider the presence of flavonoids in a plant as indication of its antioxidant, antiallergic, antiinflammatory, antimicrobial and anticancer properties. However, glycoside derivatives showed very promising activity *in vitro* and *in vivo* [15] and two of them, ethylidene derivative etoposide [16] and theylidene derivative teniposide [17] were developed as anticancer drugs. Terpenoids and steroids are capable of preventing cancer [18], because of their anticarcinogenic effects [19]. The presence of tannins shows that the plant is astringent as documented and suggests that it might have antiviral and antibacterial activities and can aid in wound healing and burns [20]. Recent reports show that tannins may have potential value as cytotoxic and/or antineoplastic agents [21]. Some researchers have also reported that some saponins have anticancer and immunomodulatory properties [14, 22]. Saponins and glycoside are also very important classes of secondary metabolites as some are

cardio-active and used in treatment of heart conditions [23].

Growth inhibition test provides such way of detecting the antimitotic activity of chemical compounds under laboratory conditions. Antimitotic chemical compounds such as vinblastine and podophyllotoxin have been shown to inhibit cell division [24, 25]. Inhibition of cell division is a measure of the antimitotic activity of chemical compounds.

In the present study, model of radish seed phytotoxicity assay described by Turker and Camper [7] was used to evaluate growth stimulation or inhibition properties of the ethanolic extracts of *C. grandis* leaf. The seed germination counts taken on the 5th day were 41.66 ± 0.88 and 25.33 ± 0.88 at 1000 ppm and 7500 ppm, respectively. (Figure 1). The root length measured on the 5th day of experiment was $13.56 \text{ mm} \pm 0.34$ at 1000 ppm extract and $8.83 \text{ mm} \pm 0.37$ at 10,000 ppm extract (Figure 2). This indicates that higher doses were much more effective in inhibiting both seed germination and root growth in length. Similar result was observed by Inayatullah *et al.* [26] and Islam *et al.* [27].



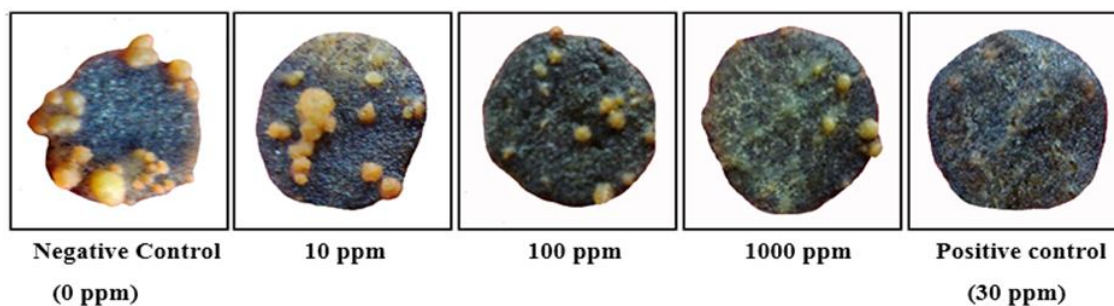


Fig 3: Effects of ethanolic extract of *C. grandis* leaf on crown gall tumor formation at different concentrations.

In the present work, *C. grandis* leaf extract showed LC_{50} value of 24.20 $\mu\text{g/ml}$ with brine shrimp nauplii. The cytotoxicity of plant material would indicate the presence of antitumor compounds in plant extract [28]. Crude extracts resulting in LC_{50} values of less than 250 $\mu\text{g/ml}$ could be considered significantly active and potential for further investigation [29]. The leaf extract in the present work, had shown LC_{50} value of less than 250 $\mu\text{g/ml}$ and this might be the potential for inhibiting tumor or cancer cells. According to Coker *et al.* [30] antitumor agents might inhibit the initiation and growth of tumors in both plant and animal systems.

The crown gall tumor bioassay has helped in the short listing of plants with anticancer activity and in the discovery of novel compounds from plants [9, 31, 32, 33, 34]. It is a simple, inexpensive, rapid and sensitive assay for chemicals that disrupt the cell cycle (mitosis, S phase, etc.) regardless of their mode of action [30]. Inhibition of crown gall formation on potato discs by the leaf extract shows similar result against the P388 (3PS) leukemia in mice [32].

Table 2: Effect of *C. grandis* leaf extract on tumor formation activities on potato discs

Concentration	Mean number of Tumor \pm SE	% of Inhibition
Negative control (Without extract or suppressor)	25 \pm 0.7071	
10 ppm	19.6 \pm 1.0770	21.60
100 ppm	14 \pm 0.9129	44.00
1000 ppm	8.5 \pm 0.6455	66.00

The leaf extract of *C. grandis* of the present work exhibited a dose dependent manner of inhibitory action. The % inhibition of tumor (Photograph-1) was compared with the standard tumor suppressor drug, camptothecin (positive control), which completely inhibited the growth of gall tumor on potato discs. Highly significant tumor inhibition e.g. 44% and 66% was observed at 100 ppm and 1000 ppm concentration (Table 2) of the leaf extract respectively because plant extracts showing more than 20% tumor inhibition activity are considered significant [31]. Similar findings were also reported by others [28, 35, 36, 37] with other plants.

4. References

- Yadav G, Mishra A, Tiwari A. Medical properties of ivy gourd (*Cephalandra indica*). A review IJPRD 2010; 2(9):92-98.
- Cromwell BT. Modern method of plant analysis. Springer-Verlag, Berlin, 1955, 373-374.
- Aplin TEH, Cannon JR. Distribution of alkaloids in some western Australian plants. Eco Bot 1971; 25(4):366-380.
- Wall ME, Krider MM, Krewson CF, Eddy CR, Willaman JJ, Corell SS *et al.* Steroidal sapogenins. Survey of plants for steroidal sapogenins and other constituents. J Pharm 1954; 1-7.
- Farnsworth NR. Biological and physiochemical screening of plants. J Pharm 1966; 55:225-276.
- Bhattacharjee AK, Das AK. Phytochemical screening of few Mysore plants. Econ Bot 1969; 23(3):204-206.
- Turker AU, Camper ND. Biological activity of common mullein, a medicinal plant. J Ethnopharmacol 2002; 82:117-125.
- Meyer BN, Ferrigni NR, Putnam JE, Jacobson LB, Nichols DE, McLaughlin JL. Brine shrimp, a convenient general bioassay for active plant constituents. Planta Medica 1982; 45: 31-34.
- Galsky AB, Wilsey JP, Powell RG. Crown-gall tumor disc bioassay, a possible aid in the detection of compounds with antitumor activity. Plant Physiol. 1980; 65:184-185.
- Tanrisever N, Fischer H, Williamson GB. Methofurans from *Calamintha ashei*, Effects on *Schizachyrium scoparium* and *Lactuca sativa*. Phytochemistry 1988; 27(8):2523-2526.
- Tashiro T, Sano S, Mori K. Synthesis of Optically Active 3-methyl-Alphahimachelene, sex pheromone of Sandfly, *Lutzomyia longipalpis* Nippon Kagakai Koen Yokoshu. 2000; 78(2):900-908.
- Ramzi AA, Mothana SA, Faisal AS, Lindquist A. Antimicrobial, Antioxidant and cytotoxic activities and phytochemical screening of some Yemeni Medicinal plants. Evidence-based Complementary and Alternative Medicine 2008; 7(3):323-330.
- Syrovets TY, Laumonnier. Project 1, Pentacyclic triterpenoids from the family of boswellic acids as antitumor agents, identification of effective molecular structures and molecular characterization of cytotoxic mechanisms. 2009 http://www.uni-ulm.de/klinik/nhk/dept/research/projekte_Syrovets_neu.htm.
- Kunle OF, Egharevba HO. Preliminary studies on *Vernonia ambigua*, phytochemistry and antimicrobial screening of whole plant. Ethnobotanical Leaflets. 2009; 13:1216-1221
- Keller-Juslén C, Kuhn M, Von-Wartburg A, Stähelin H. J. Synthesis and antimitotic activity of glycosidic lignan

- derivatives related to podophyllotoxin. *Med Chem* 1971; 14:963-968.
16. Nakanomyo H, Hiraoka M, Shiraya M. Mutagenicity tests of etoposide and teniposide. *J toxicol Sci* 1986; 11:301-310.
 17. Van-den BH. *Eur J Clin Nutr* 1997; 51:64. Van den Berg H, Wouter S, Behrendt H 1997. Treatment of Hodgkin's disease in children with alternating mechlorethamine, vincristine, procarbazine and prednisolone (MOPP) and adriamycin, bleomycin, vinblastine, and dacarbazine (ABVD) courses without radio-therapy. *Med Pediatr Oncol* 29, 23-27.
 18. Raju J, Patlolla, Swamy JM, Rao C. Diosgenin, a steroid of *Trigonella foenum graecum* (Fenugreek), inhibits azoxymethane-induced aberrant crypt foci formation in F344 rats and induces apoptosis in HT-29 human colon cancer cells. *Cancer Epidemiol Biomarkers Prev* 2004; 13:1392-1398.
 19. Yun K, Lee Y, Kwon H, Choi K. Saponin contents and anticarcinogenic effects of ginseng depending on types and ages in mice. *Zhongguo Yao Li Xue Bao* 1996; 17:293-298.
 20. Haslem E. *Plant polyphenols, Vegetable tannins revisited chemistry and pharmacology of natural products*. Cambridge University Press, Cambridge 1989; 169.
 21. Aguinaldo AM, Espeso EI, Guevara BQ, Nonato MG. Phytochemistry. In: Guevara BQ (ed.) *A Guidebook to Plant Screening, Phytochemical and Biological*. University of Santo Tomas, 2005.
 22. Evans WC. *Trease and Evans Pharmacognosy*. Ed 15, Sanders WB, London, 2002; 183-184, 191-393.
 23. Oloyode OI. Chemical profile of unripe pulp of *Carica papaya*. *Pakistan Journal of Nutrition*, 2005; 4(6):379-381.
 24. Jacobs RS, White S, Wilson L. Selective compounds derived from marine organisms: effects on cell division in fertilized sea urchin eggs. *Fed Proc*. 1981; 40(1):26-29.
 25. White SJ, Jacobs RS. Inhibition of cell division and of microtubule assembly by elatone, a halogenated sesquiterpene. *Mol Pharmacol* 1981; 20:614-620.
 26. Inayatullah S, Irum R, Ateeq-ur-Rehman M, Chaudhury F, Mirza B. Biological evaluation of some selected plant species of Pakistan. *Pharmaceut Biol* 2007; 45(5):397-407.
 27. Islam MS, Rahman MM, Rahman MA, Qayum MA, Alam MF. *In vitro* evaluation of *Croton bonplandianum* Baill. as potential antitumor properties using *Agrobacterium tumefaciens*. *Journal of Agricultural Technology* 2010; 6(1):79-86
 28. Martin-Cordero G, Saenz MT, Ayuso MJ. Cytotoxic activity of *Retama spaeocarpha*. *Fitoterapia* 1995; 16:495-498,
 29. Rieser MJ, Gu ZM, Fang XP, Zeng L, Wood KV, McLaughlin JL. Five novel mono-tetrahydrofuran ring acetogenins from the seeds of *Annona muricata*. *J Natur Products* 1996; 59:100-108.
 30. Coker PS, Radecke J, Guy C, Camper ND. Potato tumor induction assay, a multiple mode of action drug assay. *Phytomedicine* 2003; 10:133-138.
 31. Ferrigini NR, Putnam JE, Anderson B, Jacobsen LB, Nichols DE, Moore DS, McLaughlin JL. Modification and evaluation of the potato disc assay and antitumor screening of Euphorbiaceae seeds. *J Nat Prod* 1982; 45:679-686.
 32. McLaughlin JL, Chang CJ, Smith DL. Simple bench-top bioassay (brine shrimp and potato discs) for the discovery of plant antitumor compounds. In: Kinghorn AD, Balandrin MF (eds) *Human Medicinal Agents from Plants*. ACS Symposium. Washington, D.C., American Chemical Society 1993; 534:112-137.
 33. Lellau TF, Liebezeit G. Cytotoxic and antitumor activities of ethanolic extracts of salt marsh plants from the lower saxonian wadden sea, Southern North Sea. *Pharmaceut Biol* 2003; 41:293-300.
 34. Ahsan U, Ansari FL, Ihsan-ul-Haq, Nazir S, Mirza B. Combinatorial synthesis, lead identification, and antitumor study of a chalcone- based positional-scanning library. *Chemistry and Biodiversity* 2007; 4:203-214.
 35. Hussain A, Zia M, Mirza B. Cytotoxic and antitumor potential of *Fagonia cretica* L. *Turk. J Biol* 2007; 31:19-24.
 36. Islam MS, Akhtar MM, Rahman MM, Rahman MA, Sarker KK, Alam MF. Antitumor and phytotoxic activities of leaf methanol extract of *Oldenlandia diffusa* (Willd.) Roxb. *Global J Pharmacology* 2009; 2:99-106,
 37. Mazid MA, Nahar L, Datta BK, Bashar SAMK, Sarker SD. Potential antitumor activity of two *polygonum* species. *Arch. Biol. Sci. Belgrade* 2011; 63(2): 465-468.