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Screening of anticancer medicinal plants for secondary metabolites

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Abstract

Anticancer medicinal plants contain cancer healing property due to their bioactive secondary metabolites. In the present work, 23 anticancer medicinal plant species representing 10 families of Metachlamydeae were considered for qualitative assessment of the secondary metabolites such as alkaloids, flavonoids, sterols, tannins, glycosides and saponins. All plant parts except the leaf of *Leucas aspera* gave positive responses to different alkaloid detecting reagents. For 6 other secondary metabolites 13, 14, 9, 15, 9 and 15 cases gave positive responses for flavonoid, sterol, resin, tannin, glycoside and saponin tests, respectively. Results of the screening work for secondary metabolites were discussed in relation to their abundance and distribution in different test plants and their organs.

Keywords: Anticancer, Metachlamydeae, secondary metabolites, screening work

1. Introduction

The use of medicinal plants has been an important component of health care in many countries and 80% of the world's rural people meet their needs of preliminary health care from medicinal plants because plant drugs are easily available and easily accessible to most people in the sense of price compared to modern allopathic synthetic drugs [1, 2]. More than 1000 species of medicinal plants grow in Bangladesh [3], but the majority of them have not yet undergone thorough phytochemical screening for their bioactive secondary metabolites. As per published reports, about 500-752 medicinal plant species of Bangladesh have so far been enlisted and out of them, about 64 plants of have been described to possess anticancer activity [4, 5]. Anticancer medicinal plants show cancer healing property due to their secondary metabolites [6-8]. The discovery and development of vinblastine and vincristine alkaloids from the *Catharanthus roseus* and several other compounds from other sources as efficacious anticancer agents provided convincing evidence that plant's secondary metabolites could be a potential source of anticancer agents and cancer chemopreventives [9, 10]. In the present work, 23 anticancer medicinal plant species of Metachlamydeae or Sympetalae were considered for phytochemical screening for their secondary metabolites.

2. Materials and Methods

Twenty three anticancer medicinal plant species representing 10 families of Metachlamydeae or Sympetalae, enlisted in different published literature [4, 5] were considered in the present work. These plants were collected from the hilly area of Chittagong University campus and around. They were considered in the phytochemical screening work for different secondary metabolites such as alkaloids, flavonoids, sterols, tannins, glycosides and saponins. For alkaloids, the modified method of Webb [11] was followed according to Amarasingham [12] *et al.* as well as Apline and Cannon [13]. Five alkaloid detecting reagents such as Dragendorff (D), Wagner (W), Mayer (M), Hager (H) and Tannic acid (T) were prepared following Cromwell [14] were used. Flavonoids were determined in the Et-OH extract following Wall [15] *et al.* and Farnsworth [16]. Sterols, tannins, glycosides and saponins were assessed following Bhattacharjee and Das [17], Wall [15] *et al.* and Eyjolfsson [18] respectively. The presence, absence and abundance of different secondary metabolites in the test samples were indicated by -, + and multiple of + signs, respectively. Each test was replicated thrice.

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3. Results and Discussion

Results of the qualitative analysis of different secondary metabolites of 23 anticancer medicinal plants and their parts are given in Table 1 and 2.

Table 1: Alkaloid contents in different parts of 23 anticancer medicinal plants.

	Scientific name	Family	Plant/plant parts	Alkaloid detecting reagents used				
				D	W	M	T	T
1	<i>Ageratum conyzoides</i> L.	Asteraceae	Flw-buds	4+	4+	3+	2+	4+
2	<i>Alstonia scholaris</i> R.Br.	Apocynaceae	Stem bark	3+	2+	3+	3+	3+
3	<i>Andrographis paniculata</i> (Burm. F) Wall	Acanthaceae	Leaf	3+	2+	2+	3+	3+
4	<i>Anthocephalus chinensis</i> (Lamk.) A. Rich.	Rubiaceae	Stem bark	4+	2+	2+	2+	3+
5	<i>Bacopa monniera</i> (L.) Pennel.	Scrophularaceae	Leaf	3+	3+	2+	2+	3+
6	<i>Calotropis gigantean</i> (L.) Alt. f.	Asclepiadaceae	Leaf	3+	2+	2+	2+	3+
7	<i>Capsicum frutescens</i> L.	Solanaceae	Fruit	3+	3+	2+	3+	3+
8	<i>Eclipta alba</i> (L.) Hassk.	Asteraceae	Leaf	2+	+	+	2+	2+
9	<i>Elephantopus scaber</i> L.	Asteraceae	Leaf	3+	2+	3+	2+	3+
10	<i>Enhydra fluctuens</i> Lour.	Asteraceae	Leaf	3+	2+	2+	2+	3+
11	<i>Eupatorium odoratum</i> L.	Asteraceae	Leaf	2+	+	+	+	2+
12	<i>Leucas aspera</i> Spreng.	Lamiaceae	Leaf	-	-	-	-	-
13	<i>Lycopersicum esculentum</i> Mill.	Solanaceae	Fruit	2+	+	+	+	2+
14	<i>Mikania cordata</i> (Burm. f.) Rob	Asteraceae	Leaf	3+	2+	+	-	+
15	<i>Momordica charantea</i> L.	Cucurbitaceae	Fruit	2+	+	3+	+	+
16	<i>Ocimum sanctum</i> L.	Lamiaceae	Leaf	+	+	+	+	+
17	<i>Rauwolfia serpentina</i> (L.) Benth. Ex Kurz	Apocynaceae	Root	4+	3+	3+	4+	4+
18	<i>Scoparia dulcis</i> L.	Scrophulariaceae	Leaf	3+	3+	3+	3+	3+
19	<i>Solanum surattense</i> Bum. f.	Solanaceae	Leaf	+	+	+	+	+
20	<i>Tagetes erecta</i> L.	Asteraceae	leaf	2+	+	+	+	2+
21	<i>Vitex negundo</i> L.	Verbenaceae	Leaf	+	2+	+	+	2+
22	<i>Wedelia chinensis</i> (Osbeck) Merr.	Asteraceae	Leaf	3+	3+	2+	2+	3+
23	<i>Withania somnifera</i> (L.) Dunal	Solanaceae	Leaf	3+	3+	2+	2+	3+

Table 2: Flavonoids, sterols, tannins, glycosides and saponins contents in different parts of 23 anticancer medicinal plants.

	Scientific name	Family	Plant/plant parts	Six secondary metabolites					
				Flavonoids	Sterols	Resins	Tanins	Glycosides	Saponins
1	<i>Ageratum conyzoides</i> L.	Asteraceae	Flw-buds	-	-	-	+	-	-
2	<i>Alstonia scholaris</i> R.Br.	Apocynaceae	Stem bark	+	-	-	+	-	+
3	<i>Andrographis paniculata</i> (Burm. F) Wall	Acanthaceae	Leaf	+	+	+	+	-	+
4	<i>Anthocephalus chinensis</i> (Lamk.) A. Rich.	Rubiaceae	Stem bark	+	-	-	+	-	+
5	<i>Bacopa monniera</i> (L.) Pennel.	Scrophularaceae	Leaf	+	+	+	-	+	+
6	<i>Calotropis gigantean</i> (L.) Alt. f.	Asclepiadaceae	Leaf	-	-	+	-	-	-
7	<i>Capsicum frutescens</i> L.	Solanaceae	Fruit	-	+	+	-	-	+
8	<i>Eclipta alba</i> (L.) Hassk.	Asteraceae	Leaf	+	-	+	+	+	-
9	<i>Elephantopus scaber</i> L.	Asteraceae	Leaf	-	-	-	+	-	-
10	<i>Enhydra fluctuens</i> Lour.	Asteraceae	Leaf	-	+	+	+	+	+
11	<i>Eupatorium odoratum</i> L.	Asteraceae	Leaf	+	-	-	+	-	-
12	<i>Leucas aspera</i> Spreng.	Lamiaceae	Leaf	-	+	-	+	+	+
13	<i>Lycopersicum esculentum</i> Mill.	Solanaceae	Fruit	+	-	+	+	-	+
14	<i>Mikania cordata</i> (Burm. f.) Rob	Asteraceae	Leaf	-	+	-	-	-	-
15	<i>Momordica charantea</i> L.	Cucurbitaceae	Fruit	+	+	-	-	+	+
16	<i>Ocimum sanctum</i> L.	Lamiaceae	Leaf	-	+	-	+	+	+
17	<i>Rauwolfia serpentina</i> (L.) Benth. Ex Kurz	Apocynaceae	Root	-	+	-	-	-	-
18	<i>Scoparia dulcis</i> L.	Scrophulariaceae	Leaf	+	+	+	+	+	+
19	<i>Solanum surattense</i> Bum. f.	Solanaceae	Leaf	+	-	-	+	+	+
20	<i>Tagetes erecta</i> L.	Asteraceae	leaf	+	+	-	-	+	+
21	<i>Vitex negundo</i> L.	Verbenaceae	Leaf	-	+	-	+	-	-
22	<i>Wedelia chinensis</i> (Osbeck) Merr.	Asteraceae	Leaf	+	+	+	+	-	+
23	<i>Withania somnifera</i> (L.) Dunal	Solanaceae	Leaf	+	+	-	-	-	+

Out of the total 115 tests done for alkaloids, 109 tests gave positive responses in different degrees (+ to 4+) with 5 different reagents in the following proportion: D-26, W-20, M-19, H-19 and T-25 (Table 1). On the basis of such responses of the reagents to plant extracts, the relative efficacies of the

reagents for alkaloid detection appeared as: D> T> W>M>H. Pasha ^[19] reported positive response for alkaloids in 48 plant species out of 102 species of medicinal plants while Tariq ^[20] *et al.* noted 32 positive responses out of 42 they examined. In the present work, 3 plant species gave 4+, 14 gave 3+ 19 gave

2+ and 9 gave + responses. Kapoor ^[21] *et al.* noted weak positive response for alkaloids while others ^[19, 22] observed strong positive reactions (3+, 4+) for alkaloids in a few plant species. The relative abundance of alkaloids, as found in the present work, was higher in leaf, bark and rhizome than other organs of the test plants. According to Viji and Murugesan ^[23] as well as Pascaline ^[24] *et al.* the leaf and stem; and the leaf and root of medicinal plants, respectively contained a broad spectrum of secondary metabolites including alkaloids. It appears that the distribution of alkaloids is uneven and sporadic within and among different medicinal plants of the present work. Chhetri ^[25] *et al.* in a phytochemical screening for alkaloids and other bioactive chemicals, noted differences in the pattern of distribution of chemicals in different plant species and their parts. Pascaline ^[24] *et al.*, however, reported the presence of alkaloids and some other metabolites in all 10 plants species they examined.

Results of the analysis of 6 other secondary metabolites presented in Table 2 show that out of the total 138 tests there were 13, 14, 9, 15, 9 and 15 positive responses for flavonoids, sterols, resins, tannins, glycosides and saponins respectively. Pasha ^[19] reported the presence of saponins in 14 plant species while Tariq ^[20] *et al.* noted the presence of flavonoids in 21, sterols in 22, tannins in 20 and saponins only in 4 species among the lot of 23 species of Compositae. In the present work, *A. paniculata*, *B. monniera*, *E. fluctuans*, *S. dulcis* and *W. chinensis* gave maximum positive responses to spot tests done for these secondary metabolites. In a phytochemical screening work Ambil ^[26] *et al.* reported the presence of steroid glycosides and flavonoids along with alkaloids in *Citrullus* seeds while Ayoola ^[27] *et al.* noted the presence of flavonoids, terpenoids, saponins, tannins and reducing sugars in *C. papaya*, *M. indica*, *P. guajava* and *V. amygdalina*, but cardiac glycosides and alkaloids in *M. indica* as well as alkaloids and anthraquinones in *P. guajava* and anthraquinones in *V. amygdalina* were absent. Sivasankari ^[28] *et al.* while examining the major metabolites like carbohydrates, tannins, saponins, flavonoids, alkaloids, betacyanins, quinones, terpenoids, phenols, glycosides and cardiac glycosides in *Caesalpinia pulcherrima* (a domesticated shrub) and *Caesalpinia bonduc* (a wild shrub) leaf extracts prepared with 4 different solvents reported their uneven distribution in the plant species and the wild plants contributed high values for the secondary metabolites than the domesticated. The occurrence of different secondary metabolites suggests a wide range of therapeutic application of medicinal plants as they show anti-oxidant, antiallergic, anti-inflammatory, antimicrobial, anticancer activities ^[29-33].

4. Conclusion

The phytochemical screening of the medicinal plants showed the presence of various secondary metabolites which is the primary confirmation of anticancer plants. This increased the prospect for the efficient therapeutic principles of cancer treatment as most of the anticancer drugs are secondary metabolites in nature. So, further bioassay guided screening is necessary to isolate and identify the anticancer drug principals of these plants which may also be useful against other diseases.

5. References

- Sarker S. Medicinal plants and the law. Centre for Environmental law, WWF, India, 1996.
- WHO. Traditional Medicines Strategy. WHO 2002-2005, World Health Organization, Geneva. Switzerland, 2002.
- Mia AW, Ghani A. IN: Ghani, A. (ed), Traditional Medicine, Jahangirnagar University, 1990, 10-12.
- Ghani A. Medicinal plants of Bangladesh with chemical constituents and uses. Asiatic Society of Bangladesh, Dhaka. 2003, 196-197.
- Yusuf M, Chowdhury JU, Hoque MN, Begum J. Medicinal plants of Bangladesh. BCSRI, Chittagong-4220, Bangladesh, 2009.
- Syrovets T, Laumonnier Y. Pentacyclic triterpenoids from the family of boswellic acids as antitumor agents: identification of effective molecular structures and molecular characterization of cytotoxic mechanisms, 2009, 1. http://www.uni-ulm.de/klinik/nhk/dept/research/projekte_Syrovets_neu.htm.
- Kainsa S, Kumar P, Rani P. Medicinal Plants of Asian Origin Having Anticancer Potential: Short Review Asian Jof Biomed Pharma Scis. 2012; 2(10):01-07.
- Korkina L, Kostyuk V. Biotechnologically produced secondary plant metabolites for cancer treatment and prevention. Current Pharmaceutical Biotech. 2012; 13(1):265-275.
- Cragg GM, Simon JE, Jato JG, Snader KM. Drug discovery and development at the National Cancer Institute: Potential for New Pharmaceutical Crops. In: Janick J (ed) Progress in New Crops, ASHS Press, Arlington, VA. 1996, 554-560.
- Kinghorn AD. Plant secondary metabolites as potential anticancer agents and cancer chemopreventives. Molecules 2000; 5:285-288.
- Webb LJ. An Australian phytochemical survey. I. Alkaloids and cyanogenic compounds in the Queensland plants. C.S.I.R.D. Bull. 260, Melbourne, 1949.
- Amarasingham RD, Bisset NG, Millard AH, Woods MC. A phytochemical survey of Malaya plants: III. Alkaloids and saponins. Eco. Bot. 1965; 18:290-295.
- Apline THE, Cannon JR. Distribution of alkaloids in some western Australian plants. Eco. Bot. 1971; 25(4):366-380.
- Cromwell BT. Modern method of plant analysis. Springer – verlag, berlin, 1955, 373-374.
- Wall ME, Krider MM, Krewson CF, Eddy CR, Willaman JJ, Corell SS, Gentry HS. Steroidal saponins. Survey of plants for steroidal saponins and other constituents. J Pharm. 1954, 1-7.
- Farnsworth NR. Biological and Phytochemical Screening of Plants. J Pharm. Sci. 1964; 55(3):225-276.
- Bhattacharjee AK, Das AK. Phytochemical screening of few Mysore plants. Econ. Bot. 1969; 23(3):204-206.
- Eyolfsson R. Recent advances in the chemistry of cyanogenic glycosides. Frog. Org. Nat. Pro. 1970; 27:75-108.
- Pasha MK. Phytochemical survey of Bangladesh Plants, I. A. Preliminary report. Bangladesh. Journal Bot. 1977; 6(1, 2):57-64.
- Tariq M, Mossa JS, Al-Yahya MA, Al-Meshal IA, Al-Badar AA. Phytochemical and biological screening of

- Saudi medicinal plants. *Int. J Crude Drug Res.* 1987; 25(1):17-25.
21. Affandi H, Nuryadin A, Prayogo SB. Medicinal herbs of Pasir Mayang, Jambi: Ethnopharmacy and toxicity screening. *Biotropia* 2004; 22:40-58.
 22. Viji M, Murugesan S. Phytochemical analysis and antibacterial activity of medicinal plant *Cardiospermum halicacabum* Linn. *J Phytol.* 2010; 2(1):68-77.
 23. Pascaline J, Charles M, Lukhoba C, George O. Phytochemical constituents of some medicinal plants used by the Nandis of South Nandi district, Kenya. *Journal of Animal & Plant Sciences.* 2011; 9(3):1201-1210.
 24. Chhetri HP, Yogol NS, Sherchan J, Anupa KC, Mansoor S, Thapa P. Phytochemical and antimicrobial evaluations of some medicinal plants of Nepal. *Kathmandu Univ. J. Sci. Eng. & Technol.* 2008; 1(5):49-54.
 25. Ambil AA, Abdurrahman EM, Sule MI, Pateh UU, Abdurrahman YR, Ibrahim NDG. Phytochemical screening and histopathological studies of *Colosynthis citrullus* in albinprats. *Nig. Journ. Pharm. Sci.*, 2007; 6(2):7-13.
 26. Ayoola GA, Coker HAB, Adesegun SA, Adepoju-bello AA, Obaweya K, Ezennia EC *et al.* Phytochemical Screening and Antioxidant Activities of Some Selected Medicinal Plants Used for Malaria Therapy in Southwestern Nigeria. *Tropical J Pharmaceutical Research* 2008; 7(3):1019-1024.
 27. Sivasankari K, Janaky S, Sekar T. Evaluation of phytochemicals in selected medicinal plants of the *Caesalpinia species*. *Indian J Sci. Technol.* 2010; 3(12):1118-1121.
 28. Tanrisever N, Fischer H, Williamson GB. Methofurans from *Calamintha ashei*: Effects on *Schizachyrium scoparium* and *Lactuca sativa*, *Phytochemistry.* 1988; 27(8):2523- 2526.
 29. Tashiro T, Sano S, Mori K. Synthesis of Optically Active 3-methyl-Alphahimachelene, sex pheromone of Sandfly, *Lutzomyia longipalpis* Nippon Kaggakai Koen Yokoshu. 2000; 78(2):900-908.
 30. Raju J, Patlolla J, Swamy M, Rao C. Diosgenin, a steroid of *Trigonella foenum graecum* (Fenugreek), inhibits azoxymethaneinduced aberrant crypt foci formation in F344 rats and induces apoptosis in HT-29 human colon cancer cells. *Cancer Epidemiol Biomarkers Prev.* 2004; 13:1392-1398.
 31. Oloyode OI. Chemical profile of unripe pulp of *Carica papaya*. *Pakistan Journal of Nutrition.* 2005; 4(6):379-381.
 32. Ramzi AA, Mothana SA, Faisal AS, Lindquist A. Antimicrobial, Antioxidant and cytotoxic activities and phytochemical screening of some Yemeni Medicinal plants. *Evidence-based Complementary and Alternative Medicine.*, 2008; 7(3):323-330.
 33. Kunle OF, Egharevba HO. Preliminary studies on *Vernonia ambigua*: phytochemistry and antimicrobial screening of whole plant. *Ethnobotanical Leaflets.* 2009; 13:1216-1221.