



E-ISSN: 2278-4136
P-ISSN: 2349-8234
JPP 2016; 5(4): 220-223
Received: 28-05-2016
Accepted: 29-06-2016

Dora Babu Neerugatti
Associate Professor, Department
of Pharmacognosy and
Phytochemistry, Santhiram
College of Pharmacy, Nandyal,
Kurnool, Andhra Pradesh, India-
518502.

Mallikarjuna Rao Talluri
Research Associate, MCGENE
labs, Visakhapatnam, Andhra
Pradesh, India-530013.

Ganga Rao Battu
Professor, Department of
Pharmacognosy and
Phytochemistry, Andhra
University, Visakhapatnam
Andhra Pradesh-530003.

Correspondence:
Dora Babu Neerugatti
Associate Professor, Department
of Pharmacognosy and
Phytochemistry, Santhiram
College of Pharmacy, Nandyal,
Kurnool, Andhra Pradesh, India-
518502.

Cytotoxicity activity of folklore medicinal plants of India

Dora Babu Neerugatti, Mallikarjuna Rao Talluri and Ganga Rao Battu

Abstract

The present study, cytotoxicity activity of the methanolic extracts of some folklore medicinal (*Buchanania axillaris* Desr, *Tamilnadia ulignosa* Retz, *Phaseolus semierectus* L and *Stylosanthes fruticosa* Retz) was evaluated on human cancer cell lines such as colon cancer (HT-29) and breast cancer (MCF-7 and MDA-MB) using MTT assay method. The selected plant extracts showed the dose dependent cytotoxicity activity on the tested cell lines. The cytotoxicity variations on different cell lines were also observed for selected plants extracts. The cytotoxicity of the extracts were increased as the concentration of them was increased. Among all tested plants extracts *Phaseolus semierectus* showed the better cytotoxicity activity on tested cell lines. The results support the folkloric usage of the studied plants and confirmed that the studied plant possesses the constituents with cytotoxic properties that can be useful for developing new anticancer agents.

Keywords: Cytotoxicity, Cancer, Cell lines and MTT assay

1. Introduction

Cancer is the second leading disease causing deaths around the world after cardiovascular disease. Cancer is the disease can affect any part of the body, in this the affected part produce the new cell abnormally rather than normal rate of growth and causes development of a lump, mass or tumor of the cells that are no benefit to the body. The cancerous cells can spread throughout the body from the infected part of the body through metastasis. The commonly affected parts are cervical, breast, stomach, oral, lungs, liver etc [1]. In these type of cancers breast, stomach cancers are leading diseases causing the deaths in India [2]. The healing of cancer consists of psychological support, surgery, radiotherapy and chemotherapy [3]. Presently, chemotherapy is the most commonly using treatment for cancer includes alkylating agents, antimetabolites, antitumor antibiotics, platinum analogs and natural anticancer agents [4]. However, due to the growing rate of mortality because of cancer and undesirable side effects of cancer treatment, the researchers are started investigation to discover new anticancer agents from nature [5], particularly plants because of their less side effects [6, 7] and they have been using in the treatment of diseases since the pre-historic time. In this pint of view the present study was carried out to evaluate the cytotoxic activity of four folklore medicinal plants of India [8, 9] on human cancer cell lines such as colon cancer and breast cancer.

2. Materials and Methods

2.1 Plant material collection and preparation of extracts

Buchanania axillaris Desr, *Tamilnadia ulignosa* Retz, *Phaseolus semierectus* L and *Stylosanthes fruticosa* Retz were collected from of the Thalakona region, Chittoor district, India. The plant specimen was authenticated by Dr. K. Madhava chetty, Department of Botany, Sri Venkateswara University, Thirupati. The plant materials were shade dried, then powdered in mill and extracted separately with methanol using soxhlet extraction process.

2.2 Cell lines

HT-29 cell lines for colon cancer, MCF-7 and MDA-MB cell lines for breast cancer were used for the present study.

2.3 Cytotoxic assay

MTT assay method is a Colorimetric, nonradioactive, fast and economical assay widely used to quantify cell viability and proliferation of mammalian cells.

So, the cytotoxicity of the selected plants methanolic extracts were tested using MTT assay [10, 11]. The yellow tetrazolium MTT (3-(4, 5-dimethylthiazolyl-2)-2, 5-diphenyltetrazolium bromide) is reduced by metabolically active cells, in part by the action of dehydrogenase enzymes, to generate reducing equivalents such as NADH and NADPH. The resulting intracellular purple formazan can be solubilized and quantified by spectrophotometric means. Absorbance values that are lower than the control cells indicate a reduction in the rate of cell proliferation. Conversely a higher absorbance rate indicates an increase in cell proliferation. Evidence of cell death may be inferred from morphological changes.

3. Results and Discussion

Plants are the base for the treatment for various diseases before the modern medicine. The chemical drugs using in modern medicine for treatment of different diseases including cancer are have the roots from naturally isolated compounds, especially medicinal plants [12]. But the long term use of the synthetic drugs are causing severe side effects to humans [13]. Still natural products, mainly plants are significant source of new drugs [14]. In this point of view, the present study was done to identify the cytotoxicity (Anticancer) activity of *Buchanania axillaris*, *Tamilnadia ulignosa*, *Phaseolus semierectus* and *Stylosanthes fruticosa* on breast cancer (MCF-7 and MDA-MB) and colon cancer (HT-29) cell lines [15].

The selected plant extracts showed the dose dependent cytotoxicity activity on the tested cell lines (Fig. 1-3). The cytotoxicity variations on different cell lines were observed for selected plants extracts. The IC₅₀ values are showed in the table 1. The cytotoxicity of the extracts were increased as the concentration of them was increased. The selected plants *B. axillaris*, *T. ulignosa*, *P. semierectus* and *S. fruticosa* have been using by people around the southern parts of the India for treating the diseases [8, 9, 16, 17].

P. semierectus methanolic extract showed more cytotoxic activity on tested cell lines compared to other plants methanolic extracts. The IC₅₀ values are 25.50±4.64,

35.56±4.07, 122.06±3.45 on MCF-7, MDA-MB and HT-29 cell lines respectively. The variation of activity on breast cancer cell lines was observed because of their response of action on cells [15]. The *T. ulignosa*, *P. semierectus* and *S. fruticosa* extracts showed the more activity on the MCF-7 cell lines and HT-29 cell lines but surprisingly *B. axillaris* extract showed the better activity on MDA-MB cell lines (Table 1). Among all tested plants extracts *P. semierectus* showed the better cytotoxicity activity on tested cell lines. The cytotoxicity of the medicinal plants is may be due to the apoptosis or necrosis [18, 19]. Apoptosis include cell shrinkage, activation of caspases, DNA cleavage, chromatin condensation, and nuclear fragmentation. During apoptosis, activation of endonucleases causes double-strand breaks in DNA between nucleosomes leading to that DNA is fragmented into multiples less than 200 base pair pieces [20]. Necrotic cell death is an unregulated process resulting from severe damage, such as ATP depletion, hypoxia, various toxins and hyperthermia and characterized by cell swelling, lysis, and the release of intracellular contents associated with pathological tissue injury [21]. The cytotoxicity activity of the selected plants extracts is may be these process [5, 22]. Plants have different chemical constituents in them for their metabolic activities at the same time protection from their predators i.e. the compounds which are protect the plants are may be responsible for increasing the cancer cells mortality. The further studies are needed to isolate the pure compounds and their derivatives from the selected plants which are responsible cytotoxicity.

Table 1: IC₅₀ values for test extracts (*B. axillaris*, *T. ulignosa*, *P. semierectus* and *S. fruticosa*) after performing cytotoxicity assay (MTT assay) for 24h on MCF-7, MDA-MB and HT-29 cell lines.

	MCF-7	MDA-MB	HT-29
Bam	350.13±27.03	118.97±0.99	NA
Tum	42.75±4.89	98.24±2.20	138.38±7.33
Psm	25.50±4.64	35.56±4.07	122.06±3.45
Sfm	81.46±21.05	95.07±1.21	142.54±2.86

Values are mean ±SD, n= triplicate experiment.

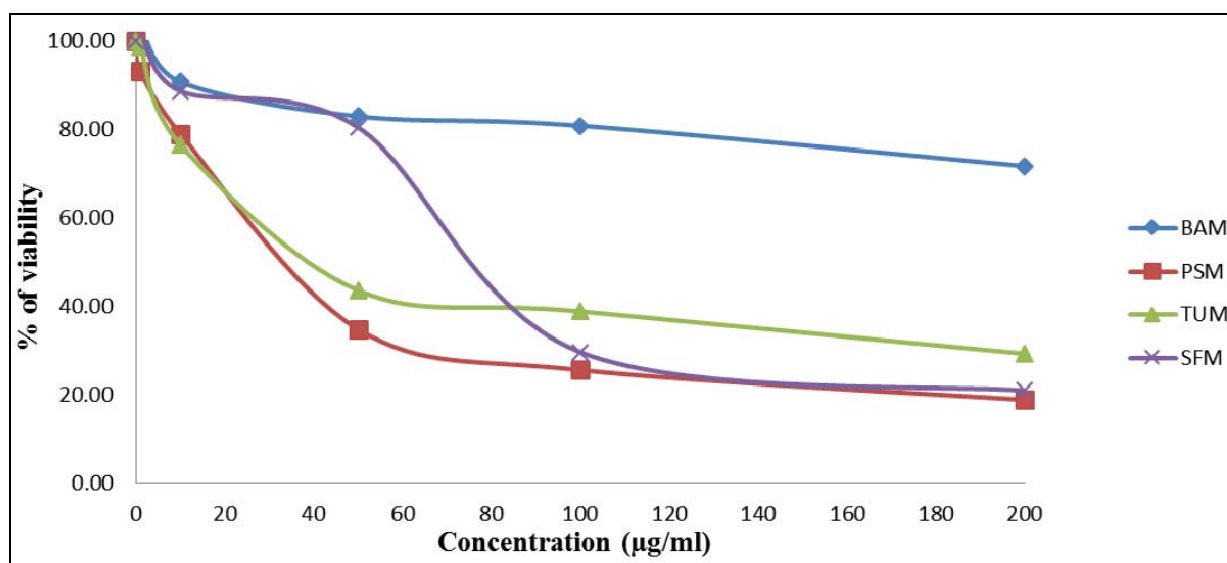


Fig 1: Graphical representation of Cytotoxicity of the test extracts (*B. axillaris*, *T. ulignosa*, *P. semierectus* and *S. fruticosa*) on MCF-7 cell line. All values are mean ± SD, n= triplicate experiment after 24h exposure

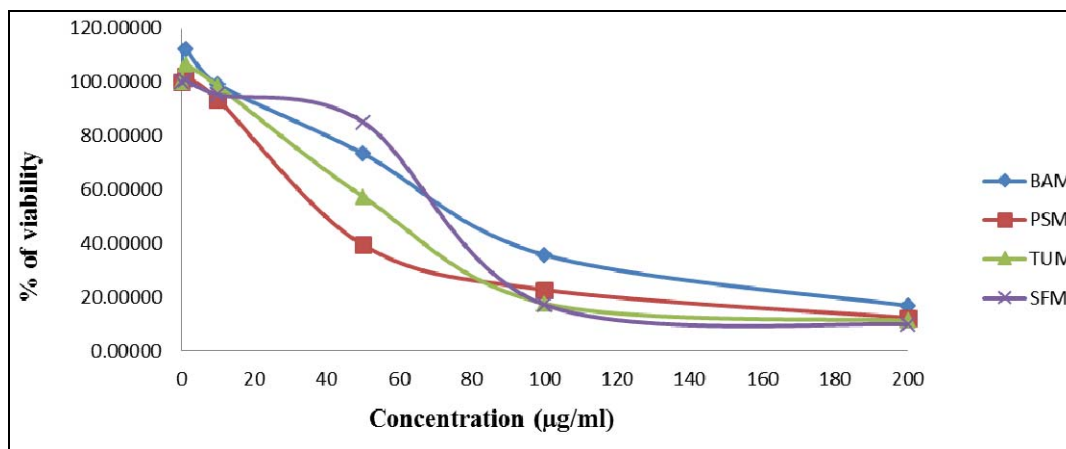


Fig 2: Graphical representation of Cytotoxicity of the test extracts (*B. axillaris*, *T. ulignosa*, *P. semierectus* and *S. fruticosa*) on MDA-MB cell line. All values are mean \pm SD, n = triplicate experiment after 24h exposure

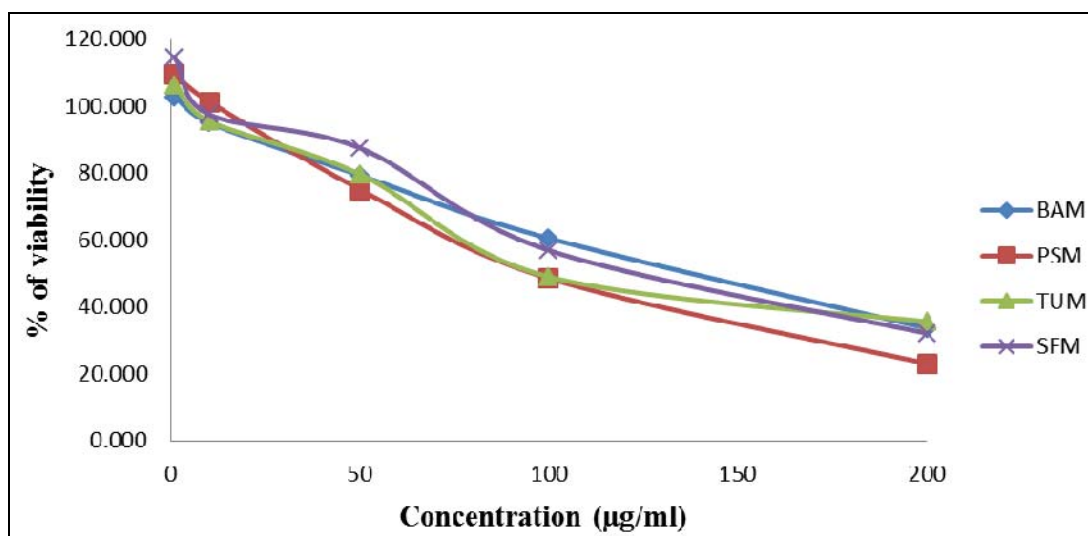


Fig 3: Graphical representation of Cytotoxicity of the test extracts (*B. axillaris*, *T. ulignosa*, *P. semierectus* and *S. fruticosa*) on HT-29 cell line. All values are mean \pm SD, n = triplicate experiment after 24h exposure

4. Acknowledgement

The authors would like to thank the TATA memorial centre, Advanced Centre for Treatment, Research and Education in Cancer (ACTREC), Kharghar, Mumbai, India for providing the cancer cell lines and doing the anticancer activity successfully.

5. References

1. Cancer Research UK and International Agency for Research on Cancer, Cancer stats, Cancer Worldwide, 2011.
2. Krishnan Nair M, Cherian Varghese R. Swaminathan. Cancer: Current scenario, intervention strategies and projections for 2015. NCMH Background Papers; Burden of Disease in India. 2015, 219-225.
3. Cancers, NMH Facts sheet. World Health Organization, 2010.
4. Carol E, DeSantis MPH, Chun Chieh Lin, Angela B, Mariotto Rebecca L, Siegel Kevin D *et al.* Cancer Treatment and Survivorship Statistics, 2014. CA CANCER J CLIN. 2014; 64:252-271.
5. Cragg GM, Newman DJ. Plants as a source of anti-cancer agents, Journal of Ethnopharmacology. 2005; 100:72-79.
6. Mallikarjuna Rao T, Ganga Rao B, Venkateswara Rao Y. Antioxidant activity of *Spilanthes acmella* extracts. International Journal of Phytopharmacology. 2012; 3(2):216-220.
7. Srikanth M, Ganga rao B, Mallikarjuna Rao T. Anticancer activity of various extracts of *Musa rosacea*, *Avicennia marina* and *Bombex ceiba*, International Journal of Pharmacy and Pharmaceutical Sciences. 2013; 5(4):553-555.
8. Madhava CK, Sivaji K, Tulasi RK. Flowering plants of Chittoor district, Andhra predesh, 2008; 1:101.
9. Venkata KR, Venkata RRR. Traditional Medicine Used by the Adivasis of Eastern Ghats, Andhra Pradesh – For Bone Fractures. Ethnobotanical Leaflets. 2008; 12:19-22.
10. Mosmann T. Rapid colorimetric assay for cell growth and survival: application to proliferation and cytotoxicity assay J Immunol. Methods. 1983; (65):55-63.
11. Denizot F, Lang R. Rapid colorimetric assay for cell growth and survival. Modification to the tetrazolium dye procedure giving improved sensitivity and reliability J. Immunol. Methods. 1986; (89):271-277.
12. Ganga Rao B, Venkateswara Rao Y, Mallikarjuna Rao T. Hepatoprotective and antioxidant capacity of Melochia

- corchorifolia Extracts, Asian Pacific Journal of Tropical Medicine. 2013; 6(7):412-420.
13. Brayand F, Moller B. Predicting the future burden of cancer. Nat Rev Cancer. 2006; 6:63-74.
 14. Abhishek Bhanot, Rohini Sharma, Malleshappa. Natural sources as potential anti-cancer agents: A review. International Journal of Phytomedicine. 2011; 3:09-26.
 15. Soule HD, Vasquez J, Long A, Albert S, Brennan M. A human cell line from a pleural effusion derived from a breast carcinoma. J Natl Cancer Inst. 1973; 51:1409-1413.
 16. Sakthivel K, Palani S, Santhosh R. Phytoconstituents Analysis By Gc-Ms, Cardioprotective And Antioxidant Activity of *Buchanania Axillaris* Against Doxorubicin-Induced Cardio Toxicity in Albino Rats. International Journal of Pharmaceutical Studies and Research. 2000; 1(1):34-48.
 17. Chuakul W, Saralamp P, Boonpleng A. Medicinal plants used in the Kutchum District, Yasothon Province, Thailand. Thai Journal of Phytopharmacy. 2011; 9(1):22-49.
 18. Wyllie AH. The biology of cell death in tumors. Anticancer Res. 1985; 5(1):131-136.
 19. Cocco RE, Ucker DS. Distinct modes of macrophage recognition for apoptotic and necrotic cells are not specified exclusively by phosphatidylserine exposure. Mol Biol Cell. 2001; 12(4):919-930.
 20. Igney FH, Krammer PH. Death and anti-death: tumour resistance to apoptosis. Nat Rev Cancer, 2002; 2(4):277-288.
 21. Martinou JC, Green DR. Breaking the mitochondrial barrier. Nat Rev Mol Cell Biol. 2001; 2(1):63-67.
 22. Newman DJ, Cragg GM, Snader KM. Natural products as sources of new drugs over the period 1981-2002. J Nat Prod. 2003; 66(16):1022-37.