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JA Sawale
RKDF University, Airport
Bypass Road, Gandhi Nagar,
Bhopal, Madhya Pradesh, India

JR Patel
RKDF University, Airport
Bypass Road, Gandhi Nagar,
Bhopal, Madhya Pradesh, India

ML Kori
RKDF University, Airport
Bypass Road, Gandhi Nagar,
Bhopal, Madhya Pradesh, India

Isolation and characterization of oleonic acid and lupeol from *Vitex negundo* leaves

JA Sawale, JR Patel and ML Kori

Abstract

Vitex negundo is commonly known as nirgundi belonging to family Verbanaceae and is found throughout India. It has been traditionally reported for the treatment of depression, malaria, venereal diseases, asthma, wounds, skin diseases, anti-inflammatory, analgesic, ulcers and snake bite. The secondary metabolites in a pure form such as flavonoids, iridoids, sesquiterpene, diterpenes, lignans and plant steroids have been isolated and identified previously. In the present research work a triterpenoids i.e. oleonic acid and lupeol isolated and characterized for the first time from *Vitex negundo* leaves.

Keywords: Triterpene, lupeol, nirgundi, Vitex

Introduction

Vitex negundo is herb which is found throughout India, locally known as nirgundi and belongs to family Verbenaceae. In Sanskrit language, the word "nirgundi" is used for plant or any substance that protects the body from the disease(s). The usefulness of the nirgundi has been written in De Materia Medica as well as in the fundamental texts of Ayurveda and Charaka Samhita [1, 2]. It has been reported the use of *Vitex negundo* in traditional medicine for the treatment of depression, venereal diseases, malaria, asthma, allergy, wounds, skin diseases, anti-inflammatory, analgesic, ulcers and snake bite [3]. Various pure compounds such as flavonoids [4] iridoids [5], sesquiterpene [6], diterpenes [7], lignans [8] and plant steroids [9] have been isolated and identified.

Upon phytochemical investigations of about 30 species from the genus *Vitex* have revealed that the major secondary metabolites of these plants are diterpenoids, ecdysteroids, flavonoids and iridoid glycosides. Other components, such as lignans, phenylpropanoids, sesquiterpenoids and triterpenoids have been also found in some *Vitex* plants, but with less frequency [10].

But these studies are not enough to identify and characterize the phytochemicals present in the plant. Hence the present study initiated to isolate and characterize the constituents from *Vitex negundo* leaves.

Materials and Methods

Collection and Authentication

Leaves of *Vitex negundo* belonging to family verbanaceae were collected in the month of august from local region of Bhopal district, Madhya Pradesh (India) and were authenticated from Dr Vinayak Naik, Senior Research Scientist, Piramal Life Sciences India Ltd. Goregaon (E), Mumbai. A voucher specimen of the plant (No. NPIL/PLS/08-689) has been deposited for future reference.

Extraction and isolation

All solvents used for extraction were of technical grade, which were distilled and dried before use. Solvents used for Column Chromatography and preparative TLC were of Analytical Reagent grade. Adsorbent for column chromatography was silica gel G 60-120 (Merck).

Procedure for Extraction and Isolation

The air dried powdered leaves (1000g) were defatted with petroleum ether (60-80°C) and remaining marc was extracted with ethanol (70%v/v) and concentrated in rotary evaporator under reduced pressure to get ethanolic extract (55g). Ethanolic extract was dissolved in ethanol and water (1:2 v/v) and partitioned with ethyl acetate and n butanol in 50mL portion for several times till complete extraction takes place. Resulting ethyl acetate fraction was concentrated under reduced pressure (22g) and was chromatographed on silica gel column (70cmX15cm, 60-120mesh, 2kg) chromatography and preparative TLC [11].

Correspondence

JA Sawale
RKDF University, Airport
Bypass Road, Gandhi Nagar,
Bhopal, Madhya Pradesh, India

Column was first eluted with chloroform, then polarity of mobile phase was gradually increased by adding methanol in different concentrations (100:0, 95:5, 90:10, 85: 15, 80:20, 70:30, 60:40v/v).

Total 232 fractions each of 50 mL were collected and TLC was performed of each fraction individually. Fractions were pooled on the basis of their TLC profile, pooled fractions (36-40) and (55-75) were selected for the isolation of constituents. Further purification was performed by preparative TLC of isolated constituents.

Detection of Phytosterols

Salkowski test

A few crystals of compound A and B were dissolved in chloroform, to this solution few drops of concentrated sulphuric acid was added. Reddish colour produces shows the presence of phytosterols [12].

LiebermannBurchard test

A few crystals of compound A and B were dissolved in chloroform, to this solution few drops of concentrated sulphuric acid followed by few drops of diluted acetic acid, 3mL of acetic anhydride. A bluish green colour formation indicates the presence of phytosterols [13].

Acetylation

The isolated pure compounds were acetylated by refluxing with acetic anhydride and previously distilled pyridine under dry conditions. The reaction mixture was allowed to stay overnight at room temperature and the mixture was concentrated to dryness under reduced pressure [14].

Analytical methods

TLC was performed on silica gel GF₂₅₄ precoated (Merck) plates. IR spectra was recorded with FTIR (Shimadzu), ¹H and ¹³C spectra recorded on Bruker (300MHz and 75.4MHz) in CDCl₃ used TMS as internal standard. ESIMS were measured using a Q-TOF micro mass spectrometer (Waters, USA).

Results and Discussion

Ethanol extract obtained was 5.5% w/w. ethyl acetate extract obtained after partitioned of ethanolic extract was 22g upon column chromatography of ethyl acetate fraction (36-40) yielded (12mg) and fraction (55-75) yielded (10mg) of pure compounds by preparative TLC.

Upon qualitative test performed on pure compounds indicated, it is of triterpenoidal nature.

Structural elucidation of compound A

It was white amorphous powder. m.p.:172-173°C. Mass spectrum showed molecular ion peak m/z at 515(M+1) which corresponds to molecular formula C₃₀H₄₈O₃

IR spectrum: An intensely broad band at 3443 cm⁻¹ showed presence of OH stretching, 2923, 1730 (carbonyl stretch), 1230 (C-O stretch, ether).

¹H NMR (300 MHz, CDCl₃, δ, TMS=0): 0.67(3H, S), 0.76(3H, S), 0.80 (3H, S), 0.83(3H, S), 0.85(3H, S), 0.91(3H, S), 1.09(3H, S), 2.04(9H, S), 2.82(2H, d, J=10.8Hz), 3.69(1H, d, J=11.7Hz), 3.87(1H, d, J=11.7Hz), 4.79(1H, m), 5.34(1H, d, J=15.3Hz)

¹³C NMR (75.4Hz, CDCl₃, δ, TMS=0): 143.62, 80.94, 77.43, 76.59, 74.54, 46.52, 45.85, 41.05, 40.52, 39.30, 38.11, 37.70, 36.79, 33.05, 32.29, 31.92, 31.23, 30.67, 30.20, 29.69, 28.04, 27.62, 25.88, 23.57, 22.46, and 22.69

The Compound A gave a positive Liebermann-Burckhardt test. It showed a molecular ion at m/z 515 corresponding to C₃₀H₄₈O₃. The ¹HNMR spectrum of compound A showed seven tertiary methyl groups at δ 0.67, 0.76, 0.80, 0.83, 0.85, 0.91 and 1.09 on an oleanane skeleton. A doublet of one proton at δ 3.69 and a doublet of one vinyl proton at δ 5.34 were assigned to H-18 and H-12, respectively, suggesting an olea-12-ene skeleton. In ¹³C-NMR spectrum, the signal corresponding to the carboxyl C-28 appeared at δ 143.62. The spectral data were similar to the ones reported for oleanolic acid.

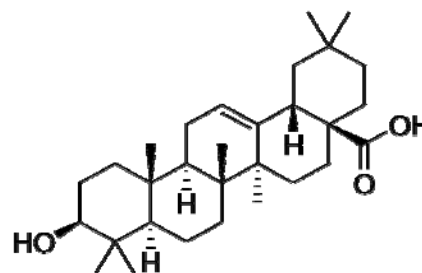


Fig 1: Chemical structure of Oleonolic acid

Characterization of compound B

It was yellowish amorphous powder. m.p: 217-219°C. Mass spectrum of isolated compound showed molecular ion m/z 271.3[M+1] corresponding to the molecular formula C₃₀H₅₀O.

IR Spectrum: Intensely broad band at 3454 cm⁻¹, moderate intense band at 1194 cm⁻¹ observed for the OH bond vibration of hydroxyl group. Carbonyl stretch was observed at 1696 cm⁻¹.The corresponding C=C vibrations was shown at 1453 cm⁻¹ was weakly intense band.

¹H NMR (300 MHz, CDCl₃, δ, TMS=0): 0.83(3H, S), 0.88(3H, S), 0.93(3H, S), 0.94(3H, S),1.00(3H, S),1.06(3H, S),1.18(3H, S), 2.21(1H, S), 3.18(1H, m), 5.05(2H, S), 5.11(2H, S).

¹³C NMR (75.4Hz, CDCl₃, δ, TMS=0): 79.05, 77.43, 64.39, 59.07, 55.19, 47.72, 47.24, 46.83, 42.08, 41.72, 40.02, 39.80, 38.78, 37.15, 36.90, 34.74, 33.75, 32.94, 31.93, 29.70, 28.75, 27.28, 26.94, 26.62, 25.99, 23.69, 23.53, 23.37, 23.27 and 22.69,

¹H NMR spectrum, revealed the presence of seven tertiary methyl protons at δ 0.83, δ 0.88, δ 0.93, δ 0.94, δ 1.00, δ 1.06, and δ 1.18. A sextet of one proton at δ 2.21 corresponds to 19 β -H is characteristic of lupeol. H-3 proton appeared as multiplet at δ 3.18 while two broad singlets at δ 5.05 and δ 5.11 due to two exomethylene protons attached at C29.

¹³C NMR spectrum, showed seven methyl groups at δ 28.75 (C- 23), δ 27.28 (C-28), δ 26.62 (C-25), δ 25.99 (C-26), δ 23.69(C-24), δ 23.37(C-27), and δ 22.69 (C-30). The deshielded signals at δ 79.05 was due to presence of hydroxyl group at C-3. The comparison with the spectral data with reported led us to propose the structure as lupeol.

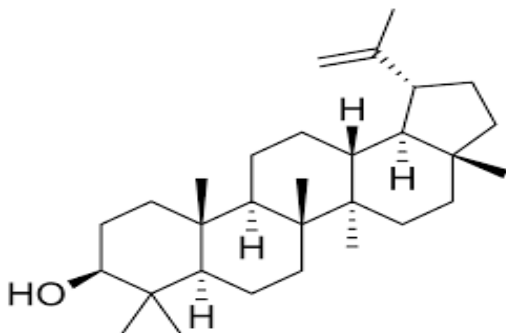


Fig 2: Chemical structure of Lupeol

Conclusion

The physical, chemical and spectral evidence of compound A and compound B confirms that the given constituents are oleonic acid and lupeol respectively.

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