



E-ISSN: 2278-4136  
P-ISSN: 2349-8234  
JPP 2018; 7(4): 2788-2793  
Received: 14-05-2018  
Accepted: 18-06-2018

**Gunjan Guleria**  
Department of Agronomy,  
Forage and Grassland  
Management, College of  
Agriculture, CSK Himachal  
Pradesh Krishi  
Vishwavidyalaya, Palampur,  
Himachal Pradesh, India

**SS Rana**  
Department of Agronomy,  
Forage and Grassland  
Management, College of  
Agriculture, CSK Himachal  
Pradesh Krishi  
Vishwavidyalaya, Palampur,  
Himachal Pradesh, India

**Amit Kumar Singh**  
ICAR-Indian Grassland and  
Fodder Research Institute,  
Jhansi, Uttar Pradesh, India

## Surveillance and distribution of different weeds in transplanted rice as influenced by integrated plant nutrition system (IPNS) in rice-wheat cropping system in Hilly area

**Gunjan Guleria, SS Rana and Amit Kumar Singh**

### Abstract

The present investigation was undertaken in a long-term experiment on IPNS in rice-wheat cropping system. Twelve treatments viz., control (no fertilizer/manure), 50, 75 and 100% NPK each to rice and wheat through fertilizers, 50% NPK to rice and 100% NPK to wheat through fertilizers, 50% substitution of N through FYM, wheat straw and green manure in rice and 100% NPK through fertilizers in wheat; 25% substitution of N through FYM, wheat straw and green manure in rice and 75% NPK through fertilizers in wheat; and farmers' practice (40% NPK through fertilizers to each crop plus 5 t FYM/ha on dry weight basis to rice) were evaluated for a period of two years (2014 and 2015) with four replications. *Ammannia baccifera* was the most dominant weed constituting 40 and 35% of the total weed flora during kharif 2014 and 2015, respectively. This was followed by *Monochoria vaginalis*, *Eleocharis* sp., *Scirpus* sp., *Echinochloa* sp. and *Cyperus difformis* during 2014 and *Monochoria vaginalis*, *Brassica* sp., *Eleocharis* sp., *Scirpus* sp., *Cyperus difformis* and *Cynodon dactylon* during 2015. On an average *Ammannia baccifera* was the most dominant and problematic weed found in rice.

**Keywords:** rice, weed, importance weed index, integrated plant nutrition system

### Introduction

Of the 30 major cropping systems identified in India (Yadav and Prasad 1998)<sup>[28]</sup>, rice-based cropping system is the most predominant in India occupying around 10.5 mha area (Sharma 2009)<sup>[20]</sup>. Farmers realize much of their food security from this cropping system. Besides food security, the low production levels jeopardize farmers' economic security to a considerable extent. To strengthen the economic conditions of the farmers, it is imperative to sustain the productivity of this system. In India, total area under rice is 43.9 mha with production of 104.3 mt (Anonymous 2016)<sup>[3]</sup>. However, fertilizers are the kingpin in increasing crop productivity. But in case of intensive cultivation, growing of exhaustive crops like rice, use of unbalanced and inadequate fertilizers accompanied by restricted use of organic manures and biofertilizers have made the soils not only deficient in the nutrients, but also deterioration in its health resulting in decline in crop response to the recommended dose of N-fertilizer. Under such a situation, integrated nutrient management (INM) has assumed a great importance and has vital significance for the maintenance of soil productivity. The integrated use of organic manures and inorganic fertilizers can help to maintain optimum crop yields and long-term soil productivity (Puli *et al.* 2016)<sup>[17]</sup>.

Weeds are the serious constraints in rice-wheat cropping system. Weed infestation is one of the factors responsible for low productivity (Singh *et al.* 2015)<sup>[23]</sup>. They reduce the crop yield and deteriorate the quality of produce (Arif *et al.* 2006)<sup>[6]</sup>. If left uncontrolled, the weeds in many fields are capable of reducing yields by more than 80% (Karlen *et al.* 2002)<sup>[16]</sup>. In rice-wheat system, yield reduction in rice due to weeds has been reported to the extent of 45% depending upon the soil type and rainfall pattern of a particular area (De Datta 1981)<sup>[13]</sup>. Researchers believe that application of organic manures is more superior for boosting crop yield and soil fertility over the synthetic fertilizers (Arif *et al.* 2012)<sup>[5]</sup>. This concept was further strengthened by Jones *et al.* (2009)<sup>[15]</sup> who found that plants are able to utilize nitrogen in organic form as well. However, it was found that application of organic manure can increase weeds population (Arif *et al.* 2013)<sup>[4]</sup> as most of the time incorporation of organic manure such as FYM served as weed seeds store bank (Baig *et al.* 2001; Ali *et al.* 2015)<sup>[8, 1]</sup>.

**Correspondence**  
**Gunjan Guleria**  
Department of Agronomy,  
Forage and Grassland  
Management, College of  
Agriculture, CSK Himachal  
Pradesh Krishi  
Vishwavidyalaya, Palampur,  
(H.P.) India

## Materials and Methods

### Experimental site

Geographically, the experimental site is situated at 32° 6' N latitude, 76° 3' E longitude and 1223.7 m altitude. in North Western Himalaya in the Palam Valley of Kangra district of Himachal Pradesh. The present study was undertaken during 2014 and 2015 in an ongoing long - term experiment which was initiated during kharif 1991 with rice - wheat cropping system at the Bhadiarkhar farm of CSK HPKV Palampur university. Palampur represents the sub-temperate humid zone of Himachal Pradesh which is characterized by mild summers and cool winters. The area receives a very high rainfall during monsoon and medium to high rainfall with an occasional snowfall during winters. Agro-climatically, the experimental site falls in the sub-temperate zone in the mid-hills of Shivalik

ranges of Himalayas which is endowed with mild summers and cool winters along with high rainfall during south-west monsoons. Average rainfall at the experimental site is 2600 mm/annum, major portion of which (80%) is received during monsoon season (June to September).

The soil of the experimental site was silty clay loam in texture, acidic in reaction (pH 5.5), high in available nitrogen (675 kg/ha), medium in available P (22 kg/ha) and K (221 kg/ha) with CEC of 11.5 c mol (p±). Taxonomically the soils of the region are classified as 'Typic Hapludalf'. The field experiment was established with rice and wheat as test crops. In this field investigation, 12 treatments were evaluated in a randomized block design with four replications which are as follows (Table 1):

**Table 1:** Details of treatments in rice-wheat cropping system

Treatment	Kharif	Rabi
T <sub>1</sub>	Control (No fertilizer, no manures)	Control (No fertilizer, no manures)
T <sub>2</sub>	50% NPK* through fertilizer	50% NPK through fertilizer
T <sub>3</sub>	50% NPK through fertilizer	100% NPK through fertilizer
T <sub>4</sub>	75% NPK through fertilizer	75% NPK through fertilizer
T <sub>5</sub>	100% NPK through fertilizer	100% NPK through fertilizer
T <sub>6</sub>	50% NPK+50% N through farmyard manure (FYM)	100% NPK through fertilizer
T <sub>7</sub>	75% NPK+25% N through farmyard manure	75% NPK through fertilizer
T <sub>8</sub>	50% NPK+50% N through wheat cut straw (WCS)	100% NPK through fertilizer
T <sub>9</sub>	75% NPK+25% N through wheat cut straw	75% NPK through fertilizer
T <sub>10</sub>	50% NPK+50% N through green manure (GM)	100% NPK through fertilizer
T <sub>11</sub>	75% NPK+25% N through green Manure	75% NPK through fertilizer
T <sub>12</sub>	Farmers' practice (40% NPK+ 5t FYM/ha)	Farmers' practice (40% NPK through fertilizer)

\*NPK - Through chemical fertilizer

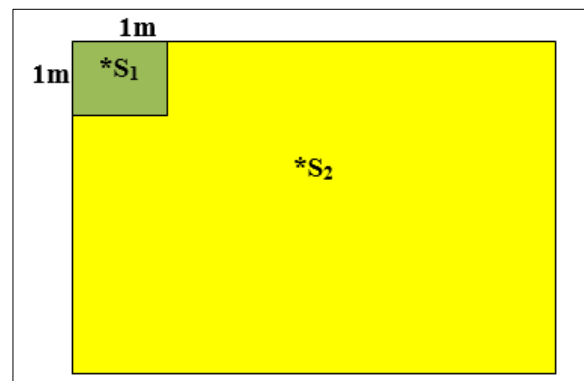
In farmers' practice, FYM 5 t/ha was applied along with 40% NPK to rice followed by 40% NPK to wheat. The recommended (100%) dose of nutrients in rice and wheat was 90:40:40 and 120:60:30 kg N, P<sub>2</sub>O<sub>5</sub> and K<sub>2</sub>O/ha, respectively. Samples of organic sources were analyzed for N, P and K contents as per the methods outlined before application in rice season and data have been reported in Table 2. Quantity of farmyard manure (FYM), wheat cut straw (WCS) and *ex-situ* green manure (GM) used in the experiment were worked out on field weight basis and incorporated before transplanting of rice.

**Table 2:** Nitrogen, phosphorus and potassium contents (%) of organics (dry wt. basis)

Organics	N	P	K
FYM (Cow dung manure)	1.20	0.225	1.013
Wheat cut straw	0.46	0.048	1.300
Green manure (Dhaincha)	2.40	0.163	1.556

### Weed studies

**Analysis of weed diversity:** Weeds are dynamic in nature. The crop(s), cropping systems and management practices mainly influence the weed shifts. Of the total losses caused by pests, weeds have a major share (30%). Studying the weed dynamics is helpful to understand the dominance or absence of a particular species in a crop/cropping system, devise means and ways to reduce their population, find out ways to delay or avoid the development of resistance by them against a herbicide, identify suitable crops for crop rotation / diversification and modify agronomic practices in favour of healthy crop growth. Manure is mainly used as a nutrient source and also in order to improve soil fertility (Baitilwake *et al.* 2011; Ali *et al.* 2012) [9, 2].



Two situations as shown above were established in each plot  
 \*(i) S<sub>1</sub> - No weed control/weedy without herbicide spray or hand weeding during both seasons and,  
 \*(ii) S<sub>2</sub> - Usual weed control both in *kharif* and *rabi*. Two samples from S<sub>2</sub> were taken randomly during each observation. From S<sub>1</sub>, only one sample was drawn each time. The important quantitative analysis such as density, frequency and abundance of weed species, was done as per Curtis and McIntosh (1950) [10].

### Density

Density is an expression of the numerical strength of a species where the total number of individuals of each species in all the quadrates is divided by the total number of quadrates studied. It is calculated by the equation as below:

$$\text{Density} = \frac{\text{Total number of individuals of a species in all quadrates}}{\text{Total number of quadrates studied}}$$

### Importance value index

This index is used to determine the overall importance of each species in the community structure. In calculating this index, the percentage values of the relative frequency, relative density and relative abundance are summed up together and this value is designated as the Importance Value Index or IVI of the species. This index is used to determine the overall importance of each species in the community structure (Curtis, 1959) [11].

### Summed dominance ratio (SDR)

It indicates the degree of dominance of a species over other species in a given sample plot. SDR of the weed species was computed using the following equation:

$$\text{SDR} = \frac{\text{Important Value Index}}{3}$$

### Similarity index

Similarity index is a measure of the relative abundance of the different species making up the richness of an area (Azizi 2016) [7].

$$\text{SI} = \frac{\text{Relative frequency of particular sp.}}{\text{Relative frequency of total of sp.}}$$

### Simpson's diversity index

The idea behind this index is that the diversity of a community is similar to the amount of information in a code or message. Simpson Diversity index is often used to quantify the biodiversity of a habitat (Simpson 1949) [21]. It takes into account the number of species present, as well as the large quantity of each species. It measures the probability that two individuals randomly selected from a sample will belong to same species. It can be measured with the following formula:

$$D = \frac{n(n-1)}{N(N-1)}$$

Where, n= total number of organisms of a particular species, N=total number of organisms of all species, Simpson's Diversity index = 1-D

As D increases, diversity decreases and Simpson's index was therefore usually expressed as  $1 - D$  or  $1/ D$ . Simpson's Diversity Index is a measure of diversity which takes into account the number of species present, as well as the relative abundance of each species. As species richness and evenness increase, so diversity increases. With this index, 1 represents infinite diversity and 0, no diversity.

## Results and Discussion

### Rice (Kharif)

#### Surveillance of weed flora

Weeds are dynamic over times and places and their competition depends on soil, climate and management practices. The weed flora in rice is very much diverse and consists of different species. In India, rice is grown under a wide variety of cultural practices in different agro-ecological conditions that is why weed diversity varied (Duary *et al.* 2015) [14]. The experimental field was monitored at different crop growth phases to look for the association of different weed species at a particular time. The occurrence of weed species noticed at a particular time of the crop growth. During a period of two years, 8 weed species were found growing in association with rice. This clearly indicated the greater diversity of weed flora that invaded the rice crop (Punia *et al.* 2007; Singh *et al.* 2012; Tiwari *et al.* 2013) [18, 22, 26]. *Monochoria vaginalis* and *Ammannia baccifera* have shown their presence at most of the stages of observation during both the years (Table 3). *Eleocharis* sp. and *Scirpus* sp. were also recorded during both the years. *Echinochloa* sp. was recorded at few observations during the first year only. *Brassica* sp. and *Cynodon dactylon* were recorded during 2015 only. It was indicated that weeds like *Brassica* sp. and *Cynodon dactylon* may likely be the menace of future in rice-wheat system (Singh *et al.* 2012) [22].

**Table 3:** Surveillance of weed flora in rice during 2014 and 2015

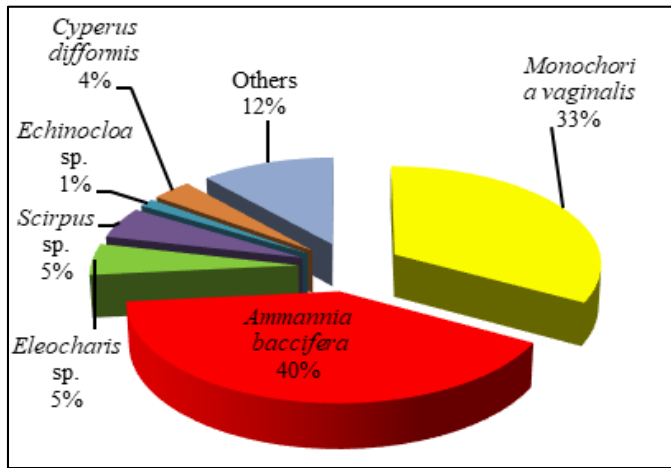
Weed species	2014				2015			
	30 DAT	60 DAT	90 DAT	Harvest	30 DAT	60 DAT	90 DAT	Harvest
<i>Monochoria vaginalis</i>	√	√	√	√	√	√	√	√
<i>Ammannia baccifera</i>	√	√	√	√	√	√	√	√
<i>Cyperus difformis</i>	√	√	-	-	-	-	√	-
<i>Echinochloa</i> sp.	√	-	-	√	-	-	-	-
<i>Eleocharis</i> sp.	-	√	√	-	√	√	√	-
<i>Scirpus</i> sp.	-	√	√	√	√	√	√	√
<i>Brassica</i> sp.	-	-	-	-	√	√	-	√
<i>Cynodon dactylon</i>	-	-	-	-	-	-	√	√
Others	-	-	√	√	√	√	√	√

√ - indicating presence

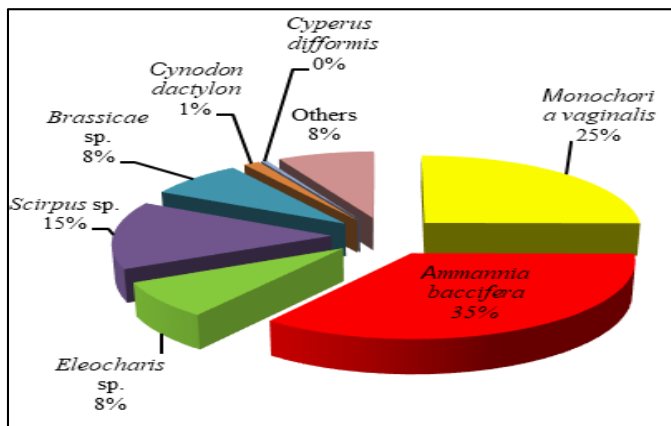
### Distribution of weed species

The year-wise overall distribution of different weeds associated in rice crop showed the *Ammannia baccifera* as most dominant weed constituting 40 and 35% of the total weed flora during *kharif* 2014 and 2015, respectively (Fig. 1a and 1b). This was followed by *Monochoria vaginalis* (33 and 25%), *Eleocharis* sp. (5 and 8%) and *Scirpus* sp. (5 and 15%). *Cyperus difformis* (4%) and *Echinochloa* sp. (1%) had shown their invasion in a negligible proportion during 2014. Similarly, *Brassica* sp. (8%) and *Cynodon dactylon* (1%) had

invaded rice during *kharif* 2015 only. Weed shift may be the reason of occurrence of new weed species. Weed shifts have occurred in response to changes in tillage (Tuesca *et al.* 2001) [27], irrigation, fertility (Davis *et al.* 2005) [12], crop rotation (Takim *et al.* 2014) [25] and herbicide use practices (Davis *et al.* 2005; Rana *et al.* 2016) [12]. Weed shifts occur when weed management practices do not control an entire weed community or population. The other unidentified weed species as a whole constituted 12 and 8% of the total weed flora during 2014 and 2015, respectively.



**Fig 1(a):** Distribution (%) of weed flora during kharif 2014 (rice) season



**Fig 1(b):** Distribution (%) of weed flora during kharif 2015 (rice) season

**Phytosociology of rice weeds and their interpretation**

Phytosociological study of weeds, which provides knowledge of the dynamics and relative importance of species in a particular phytosociety or across phytosocieties assume enough relevance in crop-weed ecosystem. It gives an appraisal of species through quantitative characters which allow effective weed management decisions (Sinha and Banerjee 2016) [24]. The treatment-wise phytosociological analysis has been presented in Table 4 (Important Value Index), 5 (Summed Dominance Ratio), 6 (Similarity Index) and 7 (Shimpson Diversity Index). The critical perusal of the data in indicated that *Ammannia baccifera* was the most important weed having highest IVI (Important Value Index), SDR (Summed Dominance Ratio), SI (Similarity Index) and lowest SDI (Shimpson Diversity Index) in all the treatments except T<sub>2</sub> (50% NPK through fertilizer to both rice and wheat), T<sub>6</sub> (50% NPK through fertilizers and 50% N through FYM to rice and 100% NPK through chemical fertilizers to wheat) and T<sub>10</sub> (50% NPK through fertilizers and 50% N through WCS (wheat cut straw) to rice and 100% NPK through fertilizers to wheat during 2015.

Under the treatments T<sub>2</sub> (50% NPK through fertilizer to both rice and wheat), T<sub>5</sub> (100% NPK through fertilizer to both rice and wheat), T<sub>6</sub> (50% NPK through fertilizers and 50% N through FYM to rice and 100% NPK through chemical fertilizers to wheat) and T<sub>9</sub> (75% NPK through fertilizers and 25% N through WCS to rice and 75% NPK through fertilizers to wheat) and T<sub>12</sub> (Farmers’ practice). *Monochoria vaginalis* had higher value of IVI, SDR, SI and lowest value of SDI followed by *Ammannia baccifera* during 2014. While, *Brassica* sp. had highest value of IVI, SDR, SI and lowest SDI in T<sub>2</sub> (50% NPK through chemical fertilizer to both crops) during 2015 than other weeds. These indices shows the diversity and prevalence of weed species in rice –wheat cropping system.

**Table 4:** Effect of fertility treatments on IVI (Important Value Index) of weeds in rice

Weed species	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	T <sub>8</sub>	T <sub>9</sub>	T <sub>10</sub>	T <sub>11</sub>	T <sub>12</sub>
<b>2014</b>												
<i>Monochoria vaginalis</i>	28.4	99.7	68.2	67.4	91.6	138.6	79.5	69.4	84.4	96.4	92.3	84.6
<i>Ammannia baccifera</i>	134.0	88.2	130.2	88.4	59.9	80.4	82.7	112.0	82.1	110.2	121.0	82.9
<i>Eleocharis</i> sp.	23.6	75.4	23.2	19.6	22.4	15.0	11.2	17.0	25.6	19.9	14.5	22.5
<i>Scirpus</i> sp.	36.1	15.4	18.4	33.8	16.5	18.8	26.5	26.2	36.2	34.3	20.4	24.3
<i>Echinochloa</i> sp.	5.7	11.9	13.9	9.9	12.9	9.9	12.0	11.3	12.2	17.1	10.0	4.1
<i>Cyperus difformis</i>	36.8	9.5	16.6	15.8	21.1	11.6	18.3	26.5	23.8	8.9	12.8	25.8
Others	35.3	0.0	29.5	65.1	75.6	25.7	69.8	37.6	35.7	13.3	29.1	55.9
<b>2015</b>												
<i>Monochoria vaginalis</i>	55.9	54.1	66.1	54.2	57.0	78.5	72.6	75.7	60.1	100.9	77.0	66.1
<i>Ammannia baccifera</i>	105.7	52.7	86.5	110.9	80.9	58.3	86.7	77.2	90.5	52.9	91.3	104.2
<i>Eleocharis</i> sp.	24.4	18.4	35.8	43.7	33.4	38.1	21.7	29.7	21.8	42.9	39.9	26.4
<i>Scirpus</i> sp.	66.6	42.0	41.4	49.6	34.8	33.2	46.7	54.7	48.9	41.5	45.3	52.6
<i>Brassica</i> sp.	0.0	82.2	28.9	0.0	42.9	25.2	43.6	28.6	45.1	22.0	29.6	17.9
<i>Cynodon dactylon</i>	16.1	11.7	0.0	8.7	6.2	23.3	3.2	14.2	3.0	8.5	5.8	10.3
<i>Cyperus difformis</i>	8.1	8.4	5.9	15.8	0.0	6.1	0.0	4.6	7.3	8.5	4.1	9.9
Others	23.2	30.5	35.3	17.2	44.9	37.3	25.4	15.4	23.3	22.6	7.0	12.7

**Table 5:** Effect of fertility treatments on SDR (Summed Dominance Ratio) of weeds in rice

Weed species	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	T <sub>8</sub>	T <sub>9</sub>	T <sub>10</sub>	T <sub>11</sub>	T <sub>12</sub>
<b>2014</b>												
<i>Monochoria vaginalis</i>	9.5	33.2	22.7	22.5	30.5	46.2	26.5	23.1	28.1	32.1	30.8	28.2
<i>Ammannia baccifera</i>	44.7	29.4	43.4	29.5	20.0	26.8	27.6	37.3	27.4	36.7	40.3	27.6
<i>Eleocharis</i> sp.	7.9	25.1	7.7	6.5	7.5	5.0	3.7	5.7	8.5	6.6	4.8	7.5
<i>Scirpus</i> sp.	12.0	5.1	6.1	11.3	5.5	6.3	8.8	8.7	12.1	11.4	6.8	8.1
<i>Echinochloa</i> sp.	1.9	4.0	4.6	3.3	4.3	3.3	4.0	3.8	4.1	5.7	3.3	1.4
<i>Cyperus difformis</i>	12.3	3.2	5.5	5.3	7.0	3.9	6.1	8.8	7.9	3.0	4.3	8.6
Others	11.8	0.0	9.8	21.7	25.2	8.6	23.3	12.5	11.9	4.4	9.7	18.6

2015												
<i>Monochoria vaginalis</i>	18.6	18.0	22.0	18.1	19.0	26.2	24.2	25.2	20.0	33.6	25.7	22.0
<i>Ammannia baccifera</i>	35.2	17.6	28.8	37.0	27.0	19.4	28.9	25.7	30.2	17.6	30.4	34.7
<i>Eleocharis sp.</i>	8.1	6.1	11.9	14.6	11.1	12.7	7.2	9.9	7.3	14.3	13.3	8.8
<i>Scirpus sp.</i>	22.2	14.0	13.8	16.5	11.6	11.1	15.6	18.2	16.3	13.8	15.1	17.5
<i>Brassica sp.</i>	0.0	27.4	9.6	0.0	14.3	8.4	14.5	9.5	15.0	7.3	9.9	6.0
<i>Cynodon dactylon</i>	5.4	3.9	0.0	2.9	2.1	7.8	1.1	4.7	1.0	2.8	1.9	3.4
<i>Cyperus difformis</i>	2.7	2.8	2.0	5.3	0.0	2.0	0.0	1.5	2.4	2.8	1.4	3.3
Others	7.7	10.2	11.8	5.7	15.0	12.4	8.5	5.1	7.8	7.5	2.3	4.2

Table 6: Effect of fertility treatments on SI (Similarity Index) of weeds in rice

Weed species	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	T <sub>8</sub>	T <sub>9</sub>	T <sub>10</sub>	T <sub>11</sub>	T <sub>12</sub>
2014												
<i>Monochoria vaginalis</i>	16.7	33.8	14.8	23.9	25.2	38.2	25.6	26.2	25.7	36.8	28.9	27.7
<i>Ammannia baccifera</i>	30.0	27.5	13.2	28.9	23.3	21.1	26.7	28.2	25.7	32.9	30.9	24.8
<i>Eleocharis sp.</i>	5.6	12.5	14.3	9.0	9.7	6.6	7.8	9.7	9.9	3.9	9.3	7.9
<i>Scirpus sp.</i>	20.0	10.0	15.9	13.9	10.7	13.2	13.3	14.6	13.9	7.9	12.4	13.9
<i>Echinocloa sp.</i>	4.4	8.8	12.6	6.5	7.8	6.6	5.6	6.8	6.9	7.9	4.1	2.0
<i>Cyperus difformis</i>	10.0	7.5	14.8	6.0	10.7	6.6	7.8	6.8	5.9	5.3	6.2	8.9
Others	13.3	0.0	14.3	11.9	12.6	7.9	13.3	7.8	11.9	5.3	8.2	14.9
2015												
<i>Monochoria vaginalis</i>	17.2	21.8	23.1	18.3	24.3	28.4	23.2	26.4	22.5	31.7	26.1	20.0
<i>Ammannia baccifera</i>	26.7	21.8	25.6	29.8	25.2	20.6	25.9	26.4	24.2	17.8	26.1	27.3
<i>Eleocharis sp.</i>	12.1	6.7	12.4	13.5	13.1	8.8	8.0	10.9	8.3	9.9	11.8	10.9
<i>Scirpus sp.</i>	23.3	17.6	21.5	22.1	13.1	12.7	18.8	19.1	22.5	13.9	21.8	20.9
<i>Brassica sp.</i>	0.0	9.2	7.4	0.0	7.5	2.9	10.7	1.8	8.3	7.9	4.2	5.5
<i>Cynodon dactylon</i>	7.8	6.7	0.0	4.8	3.7	6.9	2.7	4.5	0.8	4.0	3.4	6.4
<i>Cyperus difformis</i>	4.3	2.5	3.3	1.0	0.0	2.9	0.0	2.7	2.5	4.0	1.7	0.9
Others	8.6	13.4	6.6	10.6	13.1	16.7	10.7	8.2	10.8	10.9	5.0	8.2

Table 7: Effect of fertility treatments on SDI (Shimpson Diversity Index) of weeds in rice

Weed species	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	T <sub>8</sub>	T <sub>9</sub>	T <sub>10</sub>	T <sub>11</sub>	T <sub>12</sub>
2014												
<i>Monochoria vaginalis</i>	1.0	0.6	0.8	0.8	0.4	0.2	0.6	0.8	0.5	0.6	0.7	0.6
<i>Ammannia baccifera</i>	0.1	0.7	0.2	0.5	0.8	0.8	0.6	0.2	0.6	0.4	0.3	0.6
<i>Eleocharis sp.</i>	1.0	0.9	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
<i>Scirpus sp.</i>	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
<i>Echinocloa sp.</i>	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
<i>Cyperus difformis</i>	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
Others	1.0	0.7	1.0	0.8	0.7	1.0	0.8	1.0	1.0	1.0	1.0	0.9
2015												
<i>Monochoria vaginalis</i>	0.9	0.8	0.7	0.9	0.8	0.4	0.7	0.6	0.8	0.3	0.7	0.8
<i>Ammannia baccifera</i>	0.3	0.8	0.4	0.2	0.4	0.8	0.5	0.6	0.4	0.9	0.5	0.3
<i>Eleocharis sp.</i>	1.0	1.0	1.0	1.0	1.0	0.9	1.0	1.0	1.0	1.0	1.0	1.0
<i>Scirpus sp.</i>	0.8	0.9	0.9	0.9	1.0	1.0	0.9	0.8	0.9	0.9	0.9	0.9
<i>Brassica sp.</i>	1.0	0.5	1.0	1.0	0.9	1.0	0.9	1.0	0.9	1.0	1.0	1.0
<i>Cynodon dactylon</i>	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
<i>Cyperus difformis</i>	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
Others	1.0	1.0	1.0	1.0	0.9	0.9	0.9	1.0	1.0	0.9	1.0	1.0

## Conclusion

The overview of this experiment throws light on the impact of IPNS (Integrated plant nutrition system) on composition and abundance of weed species in rice field is helpful in understanding the dynamics of particular weed species, in terms of numbers and diversity. IPNS can contribute to the suppression of weeds. Although weeds create problems in economic gains but simultaneously conserving weeds biodiversity as an integral part of maintaining balanced agro ecosystems.

## References

- Ali K, Arif M, Ullah W, Abdullah, Ahmad W, Khan MR *et al.* Influence of organic and inorganic amendments on weeds density and chemical composition. Pakistan Journal of Weed Science Research 2015; 21:47-57.
- Ali KF, Munsif I, Uddin A, Khan, Khan N. Maize phenology as affected by tillage practices and nitrogen sources. Agriculture Science Research Journal. 2012; 2:453-458.
- Anonymous. Area and production of Rice and Wheat. Directorate of Economics and Statistics (Department of Agriculture and Cooperation, Ministry of Agriculture. Government of India) [http://eands.dacnet.nic.in/PDF/Agricultural\\_Statistics\\_2016.pdf](http://eands.dacnet.nic.in/PDF/Agricultural_Statistics_2016.pdf), 2016
- Arif M, Ali K, Haq MS, Khan Z. Biochar, FYM and nitrogen increases weed Infestation in wheat. Pakistan Journal of Weed Science Research. 2013; 19:411-418.
- Arif M, Ali K, Munsif F, Ahmad W, Ahmad A, Naveed K. Effect of biochar, FYM and nitrogen on weeds and maize phenology. Pakistan Journal of Weed Science Research. 2012; 18:475-484.
- Arif M, Khan MA, Akbar HS, Ali S. Prospects of wheat as a dual-purpose crop and its impact on weeds. Pakistan Journal of Weed Science and Research. 2006; 12:13-17.
- Azizi E. Evaluation of Weed Species Richness and Similarity of Iran's Provinces in the Fields of Some Species of Poaceae Family. International Journal of Life Science and Engineering. 2016; 2(2):8-20.
- Baig MK, Nanjappa HV, Ramachandrapa BK. Weed dynamics due to different organic sources of nutrients and their effect on growth and yield of maize. Research Crops 2001; 2:283-288.
- Baitilwake MA, Bolle SD, Salomez J, Mrema JP, Neve SD. Effects of manure nitrogen on vegetables yield and nitrogen efficiency in Tanzania. International Journal of Plant Production. 2011; 5:417-430.
- Curtis JT, McIntosh RP. The interrelations of certain analytic and synthetic phytosociological characters. Ecology. 1950; 31:434-455.

11. Curtis JT. The vegetation of Wisconsin: An ordination of plant communities. Madison, WI: University of Wisconsin Press, 1959.
12. Davis AS, Renner KA, Gross KL. Weed seed bank and community shifts in a long-term cropping systems experiment. *Weed Science*. 2005; 53:296-306.
13. De Datta SK. Principles and practices of rice production. John Willey and sons, New York. 1981, 618.
14. Duary B, Mishra MM, Dash R, Teja KC. Weed management in lowland rice. *Indian Journal of Weed Science*. 2015; 47:224-232.
15. Jones DL, Kielland K, Sinclair FL, Dahlgren RA, Newsham KK, Farrar JF *et al.* Soil organic nitrogen mineralization across a global latitudinal gradient. *Global Biogeochemical Cycles*. 2009; 23:10-16.
16. Karlen LD, Buhler DD, Ellusbury MM, Andrew SS. Soil, weeds and insect management strategies for sustainable agriculture. *Journal of Biological Science*. 2002; 2:58-62.
17. Puli MR, Prasad PRK, Ravindra PB, Jayalakshmi M, Burla SR. Effect of organic and inorganic sources of nutrients on rice crop. *Oryza* 2016; 53:151-159.
18. Punia GR, Yadav SS, Malik DB. Weed flora of wheat in Panipat district of Haryana. *Haryana Journal of Agronomy*. 2007; 23:120-121.
19. Rana SS, Kumar R, Sharma N, Badiyala D. Effect of continuous use of herbicides on weed shifts in rice-wheat system: Technical Bulletin No-1. Department of Agronomy, College of Agriculture, CSK Himachal Pradesh Krishi Vishwavidyalaya, Palampur, HP, India, 2016.
20. Sharma R. Effect of long-term integrated nutrient management system on soil and crop productivity in rice-wheat crop sequence Ph.D. thesis, Department of Agronomy, CSK Himachal Pradesh Krishi Vishwavidyalaya, Palampur, India, 2009.
21. Simpson EH. Measurement of diversity. *Nature*. 1949; 163:688.
22. Singh A, Kaur K, Kang JS, Singh G. Weed dynamics in rice-wheat cropping system. *Global Journal of Biology, Agriculture and Health Sciences*. 2012; 1:7-16.
23. Singh P, Singh P, Singh L, Qayoom S, Lone BA, Kanth RH *et al.* Phyto-sociological Association of Weeds in Summer-Kharif Crops of Kashmir Valley Under Different Eco-Situations. *Journal of Agri Search*. 2015; 2:183-188.
24. Sinha MK, Banerjee A. Studies on weed diversity and its associated phytosociology under direct dry seeded rice systems. *Communications in Plant Sciences*. 2016; 6:47-54.
25. Takim FO, Fadayomi O, Ekeleme F. Influence of cropping system and weed management practice on emergence, growth of weeds, yield of maize (*Zea mays* L.) and cowpea (*Vigna unguiculata* L.). *Poljoprivreda Agriculture*. 2014; 20:10-15.
26. Tiwari RB, Pandey TD, Sharma G, Chaure NK. Effect of weed-control measures on yield, weed control, economics and energetics of rice (*Oryza sativa*)-wheat (*Triticum aestivum*) cropping system. *Indian Journal of Agronomy*. 2013; 58:465-468.
27. Tuesca D, Puricelli E, Papa JC. A long-term study of weed flora shifts in different tillage systems. *Weed Research*. 2001; 41:369-382.
28. Yadav RL, Prasad K. In: Annual Report. 1997-98. PDCSR, Modipuram, 1998, 30-49.