

Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



E-ISSN: 2278-4136 P-ISSN: 2349-8234 JPP 2018; 7(4): 3168-3171 Received: 14-05-2018 Accepted: 18-06-2018

#### **TSSK Patro**

Acharya N.G. Ranga Agricultural University, Agricultural Research Station, Vizianagaram, Andhra Pradesh, India

#### A Meena

Acharya N.G. Ranga Agricultural University, Agricultural Research Station, Vizianagaram, Andhra Pradesh, India

#### M Divya

Acharya N.G. Ranga Agricultural University, Agricultural Research Station, Vizianagaram, Andhra Pradesh, India

#### N Anuradha

Acharya N.G. Ranga Agricultural University, Agricultural Research Station, Vizianagaram, Andhra Pradesh, India

Correspondence TSSK Patro Acharya N.G. Ranga Agricultural University, Agricultural Research Station, Vizianagaram, Andhra Pradesh, India

# Eco-friendly management of incitant *Rhizoctonia solani*, the causing sheath blight in Kodo millet

# TSSK Patro, A Meena, M Divya and N Anuradha

#### Abstract

Sheath blight caused by *Rhizoctonia solani* is one of the most serious diseases worldwide. The disease is currently managed only by the excessive application of chemical fungicides which are toxic and not environmentally friendly. Therefore, greater emphasis should be given to biological control as being both safe and effective. A field experiment was conducted during *kharif* 2017, at Agricultural Research Station, Vizianagaram for the management of banded sheath blight disease in kodo millet by using potential biocontrol agents *viz.*, *Bacillus subtilis*, *Pseudomonas fluorescens* and *Trichoderma asperellum*. Lowest sheath blight intensity (49.33%) was recorded in T<sub>7</sub> *i.e.* soil application of value added *P*. *fluorescens* + *T. asperellum* + *B. subtilis* (one kg talc formulation mixed in 25 kg FYM or vermicompost, incubated for 15 days) followed by 53.33% in T<sub>1</sub> (*i.e.*, Seed treatment with *Trichoderma asperellum* @ 10 g/kg) and the highest (61.33%) in T<sub>6</sub> whereas it was 76.00% in the control. And high grain (1274.07 kg/ha) and fodder yield (2233.33 kg/ha) was found in T<sub>7</sub> *whereas*, it was 459.26 kg/ha and 1390.30 kg/ha in the control respectively. In mean of all locations the lowest sheath blight intensity (22.11) was recorded in T<sub>7</sub> *i.e.* soil application of value added *P. fluorescens* + *T. asperellum* + *B. subtilis* (one kg talc formulation mixed in 25 kg FYM or vermicompost, incubated for 15 days) and the highest (31.31%) in T<sub>3</sub> whereas it was 41.24% in the control.

Keywords: Kodo millet, bio control, R. solani

## Introduction

Kodo millet (*Paspalum scrobiculatum*) is one of the hardiest crops grown in Madhya Pradesh, Maharashtra, and Uttar Pradesh and various other parts of India. Minor millets like kodo is also described as nutritious millet and has received far less research and development attention than other crops with regard to crop improvement and utilization. It is the main source of protein and minerals in the daily diets of tribal and weaker section living in remote rural areas. Millets are nutritionally superior than other cereals. The millet contains a high proportion of complex carbohydrate and dietary fiber which helps in prevention of constipation and slow release of glucose to the blood stream. Glycemic index is an important tool used in treating people with diabetes, cardiovascular disease management and weight regulation programs. Millets including Kodo contain water soluble fiber and this property may be utilized for maintaining or lowering blood glucose response among diabetic and CVD patients. Glycemic load (GL) representing both quality and quantity of carbohydrate in a food and allows comparison of the likely glycemic effect of realistic portion of the different foods (Neelam, *et al.*).

Banded blight of kodo millet incited by Rhizoctonia solani (Kuhn.) (Basidial stage: Thanatephorus cucumeris (Fr.) Donk) is one of the emerging malady in it's successful cultivation. Lalu Das and Girija (1989) for the first time reported as sheath blight of ragi from Vellayani in Kerala, where it occurred in a severe form. The disease was observed in severe form at the Agricultural Research Station in Vizianagaram, The widespread adoption of new, susceptible, high-yielding cultivars with large numbers of tillers, and the changes in cultural practices associated with these cultivars, favor the development of sheath blight and contribute greatly to the rapid increase in the incidence and severity of this disease in rice-producing areas throughout the world (Groth et al. 1991; Rush and Lee, 1992) [8, 19]. Furthermore, environmental conditions such as low light, cloudy days, high temperature and high relative humidity also favor the disease (Ou, 1985) <sup>[14]</sup>. The pathogen overwinters as soil-borne sclerotia and mycelium in plant debris; these constitute the primary inoculums. The disease is characterized by oval to irregular, light grey to dark brown lesions on the lower leaf sheath. In advanced stages, the lesions enlarge rapidly and coalesce to cover large portions of the sheath and leaf lamina. At this stage, the disease symptom is characterized by a series of copper or brown color bands across the leaves giving a very characteristic banded appearance.

Control of the pathogen is difficult because of its ecological behavior, its extremely broad host

range and the high survival rate of sclerotia under various environmental conditions (Groth *et al.* 2006) <sup>[9]</sup>. In the absence of a desired level of host resistance, the disease is currently managed by excessive application of chemical fungicides, which have drastic effects on the soil biota, pollute the atmosphere, and are environmentally harmful. Some potentially effective fungicides are highly phytotoxic to the crop and, if the disease is not severe, these fungicides may reduce yield (Groth *et al.* 1990) <sup>[7]</sup>. It is difficult to achieve control through host resistance or fungicides, therefore, biological control may be effective in minimizing the incidence of sheath blight (Das and Hazarika, 2000) <sup>[3]</sup>. So an experiment was conducted at Agricultural Research Station, Vizianagaram during *Kharif* 2017.

# **Materials and Methods**

A field experiment was conducted during *kharif* 2017, at Agricultural Research Station, Vizianagaram for the management of banded blight disease in finger millet by using potential biocontrol agents like *Bacillus subtilis*,

*Pseudomonas fluorescens* and *Trichoderma asperellum*. These isolates were collected from Department of Biological control, Vizianagaram. The experiment was laid out in randomized block design (RBD) with three replications at spacing of  $22.5 \times 10$  cm with  $3 \times 3$  m plot size. Standard agronomic practices of NPK-50kg, 40kg, 25kg were followed at the time of crop growth period. A susceptible variety (Co 3) was used in this experiment by imposing the following treatments: (Table 1)

The disease severity and yield were recorded and the data was statistically analysed by following the standard procedures (Gomez and Gomez, 1984)<sup>[5]</sup>. Banded blight (Anon, 1996)<sup>[1]</sup> was recorded by using 0 to 9 scale (Table 2). The percent disease index (PDI) was calculated by using the following formula:

## Sum of all the numerical ratings

- × 100

Number of observations × Maximum disease grade

PDI=-

| T1 | Seed treatment with Trichoderma asperellum @ 10 g/kg  |  |  |  |  |  |
|----|---|--|--|--|--|--|
| T2 | Seed treatment with Pseudomonas fluorescens @ 10 g/kg   |  |  |  |  |  |
| T3 | Seed treatment with Bacillus subtilis @ 10 g/kg   |  |  |  |  |  |
| T4 | Soil application of value added P.f. (one kg talc formulation mixed in 25 kg FYM or vermicompost, incubated for 15 days)      |  |  |  |  |  |
|    | applied over an acre at the time of sowing  |  |  |  |  |  |
| T5 | Soil application of value added T.a. (one kg talc formulation mixed in 25 kg FYM or vermicompost, incubated for 15 days)      |  |  |  |  |  |
|    | applied over an acre at the time of sowing  |  |  |  |  |  |
| T6 | Soil application of value added B.s. (one kg talc formulation mixed in 25 kg FYM or vermicompost, incubated for 15 days)      |  |  |  |  |  |
|    | applied over an acre at the time of sowing  |  |  |  |  |  |
| T7 | Soil application of value added P.f. + T.a. + B.s. (one kg talc formulation mixed in 25 kg FYM or vermicompost, incubated for |  |  |  |  |  |
|    | 15 days) applied over an acre at the time of sowing   |  |  |  |  |  |
| T8 | Control   |  |  |  |  |  |

## **Table 2:** Standard Evaluation System (SES) scale for sheath blight disease

| Score | Description  | Reaction      |
|-------|--|---------------|
| 0     | No incidence   | No disease/HR |
| 1     | Vertical spread of the lesions up to 20% of plant height     | R             |
| 3     | Vertical spread of the lesions up to 21-30% of plant height  | MR            |
| 5     | Vertical spread of the lesions up to 31-45% of plant height  | MS            |
| 7     | Vertical spread of the lesions up to 46-65% of plant height  | S             |
| 9     | Vertical spread of the lesions up to 66-100% of plant height | HS            |

**Statistical analysis:** The data was analyzed by applying statistical tools of ANOVA (Analysis of variance) technique for drawing conclusions from the data. Critical difference (C.D) was calculated to see the significant and non-significant difference between the mean values of sheath blight PDI in all the treatments.

## **Results and Discussion**

All the treatments were found significantly superior over check in controlling the disease. Among all the treatments

tested, the lowest sheath blight intensity (49.33%) was recorded in  $T_7$  *i.e.* soil application of value added *P*. *fluorescens* + *T. asperellum* + *B. subtilis* (one kg talc formulation mixed in 25 kg FYM or vermicompost, incubated for 15 days) followed by 53.33% in  $T_1$  (*i.e.*, Seed treatment with *Trichoderma asperellum* @ 10 g/kg) and the highest (61.33%) in  $T_6$  whereas it was 76.00% in the control. And high grain (1274.07 kg/ha) and fodder yield (2233.33 kg/ha) was found in  $T_7$  whereas, it was 459.26 kg/ha and 1390.30 kg/ha in the control respectively (Table 3).

Table 3: Management of banded sheath blight in Kodo Millet

| Treatments               | Sheath blight<br>(Vizianagaram) | Grain Yield (Kg/ha) | Fodder Yield (Kg/ha) | Sheath blight<br>(Mean of all centres) | Grain Yield (Kg/ha) | Fodder Yield (Kg/ha) |
|--------------------------|---------------------------------|---------------------|----------------------|--|---------------------|----------------------|
| T1                       | 53.33                           | 1255.56             | 2131.44              | 25.87                                  | 1662.47             | 2291.85              |
| $T_2$                    | 59.33                           | 992.59              | 1638.22              | 28.22                                  | 1688.03             | 2135.00              |
| T <sub>3</sub>           | 65.33                           | 751.85              | 1506.38              | 31.31                                  | 1682.84             | 2140.93              |
| $T_4$                    | 58.67                           | 1033.33             | 1659.26              | 26.41                                  | 1608.77             | 2101.11              |
| T <sub>5</sub>           | 54.67                           | 1225.93             | 2040.74              | 24.00                                  | 1604.57             | 2425.37              |
| T <sub>6</sub>           | 61.33                           | 840.74              | 1588.89              | 29.89                                  | 1442.47             | 1942.59              |
| T <sub>7</sub>           | 49.33                           | 1274.07             | 2233.33              | 22.11                                  | 1671.36             | 2695.37              |
| T <sub>8</sub> (Control) | 76.00                           | 459.26              | 1390.30              | 41.24                                  | 1141.98             | 1794.07              |

Journal of Pharmacognosy and Phytochemistry

| Mean | 59.75 | 979.17 | 1773.61 | 28.63 | 1562.81 | 2190.79 |
|------|-------|--------|---------|-------|---------|---------|
| CD   | 10.27 | 233.26 | 365.87  | 7.42  | 293.54  | 502.54  |
| CV % | 9.82  | 13.60  | 11.78   | 14.79 | 10.73   | 9.70    |

In mean of all locations the lowest sheath blight intensity (22.11) was recorded in  $T_7$  *i.e.* soil application of value added *P. fluorescens* + *T. asperellum* + *B. subtilis* (one kg talc formulation mixed in 25 kg FYM or vermicompost, incubated for 15 days) and the highest (31.31%) in  $T_3$  whereas it was 41.24% in the control.

Patro and Madhuri (2014) reported that P. fluorescens + T. harzianum followed by P. fluorescens alone and T. harzianum alone are effective against R. solani. Pal et al. (2015) revealed that seed treatment + 3 spraying with T. viride @ 1% was the most effective bio control treatment recording 10.93% pooled PDI against 34.41% in control plot and its performance was at par with the standard fungicide propiconazole @ 1%. The treatment also exhibited maximum increase in all the yield attributing factors recorded and gave a yield increase of 41.1% over control. The interaction between host and pathogen resulted significant changes in morphological, phenological parameters, which influence the yield and yield traits adversely, there was significant reduction in grain yield plant and fodder yield plant ranging from 2.1 to 18.5% and 8.5 to 26.6%, respectively was recorded in Rhizoctonia solani affected plants of little millet (Shailendra Singh Chouhan, 2014) <sup>[20]</sup>. Srinivas et al. (2013) <sup>[22]</sup> depicts that all the bioagents stopped the growth of R. solani after contact. The order of percent inhibition of Trichoderma viride (72.65%)> Penicillium notatum (64.07%)> T. atroviride (62.51%)>T. harzianum (42.18%)> T. longibrachiatum (38.29%)> T. koninzii (3.14%)> Aspergillus niger (1.57%). T. harzianum (ThF2-1) gave the maximum inhibition of R. solani 618 (Montealegre et al. 2014)<sup>[11]</sup>. Huang et al (2012)<sup>[10]</sup> reported that B. pumilus SQR-N43 is a potent antagonist against R. solani Q1. T. harzianum (Jn14) and T. hamatum (T36) were the most effective isolates to inhibit R. solani mycelial growth (Barakhat et al. 2007)<sup>[2]</sup>. Trichoderma strains were effective both in vitro and in vivo was reported by Das and Hazarika (2000)<sup>[3]</sup> and Tewari and Singh (2005)<sup>[23]</sup> who all found that T. harzianum was an effective BCA in controlling rice sheath blight. Patro et al. (2017)<sup>[4]</sup> results revealed that KAVT 5, KAVT 20 and KAVT 22 were found as resistant genotypes. Patro (2016) <sup>[15]</sup> recorded resistant varieties in Kodo millet under in-vivo conditions.

It is also possible to state that the signs that BCAs will be able to control sheath blight are good. Supplementing biological control with other, non-chemical control methods will improve disease control still more. On the other hand, biological control with the antagonists will lower the dependency on synthetic will it is hoped lead to a cleaner environment and healthier foods.

## References

- 1. Anonymous. Standard evaluation system for rice. International Rice Testing programme. International Rice Research Institute Report, Philippines, 1996.
- Barakat MR, Al-Mahareeq F, Ali –Shtayeh MS, AL-Masri MI. Biological Control of *Rhizoctonia solani* by Indigenous *Trichoderma* spp. isolates from Palestine. Hebron University Research Journal, 2007; 3(1):1-15.
- 3. Das BC, Hazarika DK. Biological management of sheath blight of rice. Indian Phytopathology. 2000; 53:433-435.
- 4. Divya M, Patro TSSK, Sandhya Rani Y, Triveni U, Anuradha N. Reaction of *Eleusine coracana* (L.) Gaertn.

Early and medium duration varieties against major maladies. Progressive Research-An International Journal, 2017; 12(1):30-32.

- Gomez KA, Gomez AA. Statistical procedures for Agricultural research. 2<sup>nd</sup> Edition. John Wiley & Sons Publications, New York, 1984, 680.
- Gowda KTK, Gowda J, Ashok EG, Nagaraja A, Jagadish PS, Sashidhar VR *et al.* Technology for increasing finger millet and other small millets production in India. Project Coordinating Cell, ICAR, University of Agricultural Sciences, Gandhi Krishi Vigyan Kendra Campus, Bangalore, 2006, 41.
- Groth DE, Rush MC, Lindberg GD. Foliar fungicides for control of rice diseases in the United States. In: *Pest Management in Rice* (BT Grayson, MB Green, LG Copping, ed.), Elsevier, London, UK, 1990, 31-52.
- Groth DE, Rush MC, Hollier CA. Rice diseases and disorders in Louisiana. Louisiana State University, Agricultural Center, Louisiana Agricultural Experiment Station, Bulletin No. 828. Baton Rouge, LA, USA, 1991.
- 9. Groth DE, Bond JA. Initiation of rice sheath blight epidemics and effects of application timing of azoxystrobin on disease incidence, severity, yield and milling yields. Plant Disease. 2006; 90:1073-1078.
- Huang K, Zhang N, Yong X, Yang X, Shen Q. Biocontrol of *Rhizoctonia solani* damping-off disease in cucumber with *Bacillus pumilus* SQR-N43. Microbial Research. 2012; 167(3):135-143.
- 11. Montealegre JR, Ochoa F, Besoain X, Herrera R, Pérez LM. *In vitro* and glasshouse biocontrol of *Rhizoctonia solani* with improved strains of *Trichoderma* spp. Ciencia E Investigacion Agraria. 2014; 41(2):197-206.
- 12. Nagaraja A, Kumar J, Jain AK, Narasimhudu Y, Raghuchander T, Kumar B *et al.* Compendium of small millets diseases. Project Coordination Cell, All India Coordinated Small Millets Improvement Project, University of Agricultural Sciences, Gandhi Krishi Vigyan Kendra Campus, Bangalore, 2007, 80.
- Neelam Yadav, Chaudhary Kanchan, Singh Alka, Gupta Alka. Evaluation of hypoglycemic properties of Kodo millet based food products in healthy subjects. Iosr Journal of Pharmacy. 2013; 3(2):14-20.
- 14. Ou SH. Rice Diseases. 2nd ed. Commonwealth Mycological Institute, Kew, UK, 1985, 380.
- Patro TSSK, Divya M, Ashok S. Identification of resistant sources for major diseases in finger millet. Progressive Research-An International Journal, 2016; 11(4):299-301.
- 16. Patro TSSK. Studies on kodo millet entries for their resistance to banded blight under *in-vivo*. Lead paper presented at International Conference and Innovative approach in applied science and technologies at Kasetsart Universit, Bangkok, Thailand, 2016.
- Patro TSSK, Divya M, Sandhya Rani Y, Triveni U, Anuradha N. Detection of resistant sources for major diseases in finger millet (*Eleusine coracana* (L.) Gaertn.). Frontiers of Crop Improvement Journal, 2017; 5(1):43-45.
- 18. Patro TSSK, Divya M, Sandhya Rani Y, Triveni U, Anuradha N. Resistance of Kodo millet (*Paspalum scrobiculatum*) varieties against *Rhizoctonia solani* the

incitant of banded sheath blight disease. Frontiers of Crop Improvement Journal, 2017; 5(1):78-79.

- 19. Rush MC, Lee FN. Sheath blight. In: *Compendium of Rice Diseases* (RK Webster, PS Gunnell, ed.), APS Press, St. Paul, MN, USA, 1992; 22-23.
- 20. Shailendra Singh Chouhan. Studies on banded leaf and sheath blight of little millet (*Panicum sumatrense*) caused by *Rhizoctonia solani* Kuhn. Thesis, Jawaharlal Nehru Krishi Vishwa Vidyalaya, Jabalpur (Madhya Pradesh), 2014.
- 21. Sinha AP, Upadhyay JP. Millet ke rog. (In Hindi), Directorate of Publication, GB Pant University of Agriculture and Technology, Pantnagar. 1997; 263:145-180.
- 22. Srinivas P, Ved Ratan, Narayan Reddy P, Bindu Madhavi G. *In-vitro* evaluation of fungicides, biocontrol agents and plant extracts against rice sheath blight pathogen Rhizoctonia solani. International Journal of Applied Biology and Pharmaceutical Technology. 2013; 5(1):121-126.
- 23. Tewari L, Singh R. Biological control of sheath blight of rice by *Trichoderma harzianum* using different delivery systems. Indian Phytopathology. 2005; 58:35-40.