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Soil physico-bio-chemical properties under different agroforestry systems in Terai region of the Garhwal Hiamalayas

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Abstract

An experiment was carried out to evaluate the effect of different agroforestry systems on soil physico-chemical and biological properties. The results revealed that different agroforestry tree species showed the positive impact on soil physico-chemical and biological properties in comparison to sole agriculture cropping system. At 0-15, 15-30 and 30-45 cm soil depth lowest soil bulk density (1.25, 1.27 and 1.28 g/cm³), particle density (2.62, 2.66 and 2.71 g/cm³) and pH (6.50, 6.90 and 6.80) was recorded under *Populus deltoids*, *Anthocephalus cadamba* and *Madhuca indica* based agroforestry systems respectively. Soil organic carbon content was recorded highest (1.06, 0.90 and 0.84%) under *Quercus leucotrichophora* at 0-15, 15-30 and 30-45 cm soil depth. Nitrogen, phosphorus and potassium content were also higher under *Q. leucotrichophora* based agroforestry system. In soil biological properties, the highest bacterial and fungal colony was also recorded under *Quercus leucotrichophora* based agroforestry system. So, in this study, *Quercus leucotrichophora* based agroforestry system found superior for the enhancement in soil fertility over the other tree based agroforestry systems as well as open cropping system.

Keywords: Soil, physico-bio-chemical properties, different agroforestry systems

Introduction

In an agroforestry system woody perennials (trees, shrubs, palms, bamboos, etc.) are deliberately combined with agricultural crops and/or animals in same land management unit, either in some form of spatial arrangement or temporal sequence. There are both ecological as well as economical interactions between the different components of agroforestry (Nair, 1993). The advantages of ecological interactions between trees and agricultural crops are *i.e.*, increase in soil fertility through nitrogen fixation, addition of organic matter production, recycling of nutrients (Young, 1986) [93], more biomass production per unit area, uptake of more water and nutrients (Huxley, 1983) and trees act as a protective barrier against soil erosion or as wind breaks (Wiersum, 1984) [89]. Apart from the effect of agroforestry in improving the soil physical properties through the improvement in the soil structure and porosity, it also influences the chemical properties of the soil. Trees add high amount of organic matter in the form of leaf litter, fine root biomass and pruning debris. They help in lowering down the pH and EC of soil through organic matter accumulation and addition of nitrogen, potassium and phosphorous. Soil microorganisms mineralize nutrients via organic matter decomposition. The living microbial cells comprises of 1% to 5% (w/w) of the total organic carbon, and 1% to 6% of the total organic nitrogen (Jankinson and Paulson, 1976). Soil organic matter decomposition by various microorganisms takes place through various enzymes which catalyze innumerable reactions necessary for the life processes of microorganisms, decomposition of organic residues, nutrient cycling, organic matter formation and soil structure (Dick, 1992) [19]. Most of the soils are not so rich to supply all the nutrients for its optimum growth and development. Furthermore, it is difficult to sustain the yield of the crop and soil health for longer duration without integrated use of organics and inorganics. The removal of nitrogen, phosphorus and potassium by crops is much more than their replenishment through mineral fertilizers, thereby leading to nutrient mining but this problem can be overcome through the agroforestry. Because agroforestry improve the soil fertility status by the addition of continuous organic matter in the soil in the form of leaves, twig and branches etc. At present, the level of N, P and K removal in India is about 28 million tonnes against addition of only 18 million tonnes, thus resulting in a negative balance of about 10 million tonnes (Rao and Srivastava, 1998) [66]. The removal of nutrient from the soil can also be minimize by the agroforestry because the tree root works as a binding agent against the soil

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Erosion and enhance the natural nutrient recycling into the soil. Thus, Present study was conducted for the assessment of soil health under different agroforestry tree species in Terai region of Udham Singh Nagar, Uttarakhand, at Agroforestry Research Centre, Pantnagar, and District U. S. Nagar.

Material and methods

Study site

The present investigation was carried out at Agroforestry Research Centre, Govind Ballabh Pant University of Agriculture and Technology, Pantnagar. The study site was located at 29° Latitude and 79°3' longitudes and at an altitude of 243.84 meters above the mean sea level in Terai region. The climate and weather of Pantnagar is humid sub-tropical with cold winters and hot dry summers. The maximum daily temperature in summer may reach up to 42°C and minimum temperature in winter may fall up to 0.5°C. Generally, south-west monsoon sets in the second or third week of June and continues up to the end of September. The mean annual rainfall is about 1450 mm, of which 80-90 per cent is received during the wet season (July to September). The soils of Terai region are developed from alluvial, medium to moderately

coarse textures materials under predominant influence of tall vegetation and moderate to well drain conditions. The soils are weakly developed with mollic epipedons and horizons are classified as Mollisols (Despandey *et al.*, 1971).

Experimental Details

The soil samples were collected from three depths 0-15cm, 15-30 cm and 30-45 from eight different agroforestry tree species and sole agriculture field (Open condition) as control during February to June 2018. The agroforestry systems were established in the year 2003-04. All the agroforestry tree species were regularly pruned up to five year to maintain as a single stem. First seven year wheat and soyabean crop were intercropped in all the agroforestry systems alternatively. Thereafter wheat-soyabean was replaced with turmeric and zinger due to drastic decrease in the yield of wheat and soyabean. In the present study we had selected eight agroforestry systems and one sole agriculture crop field. Each tree based agroforestry system and sole cropping system is considered as a treatment. Thus, there were nine treatments including control (Sole agriculture field). The details of the tree species are as following:

Table 1: Details of treatments

S. No.	Scientific name	Common name	Family	Spacing
1	<i>Eucalyptus spp.</i>	Eucalyptus	Myrtaceae	4×4
2	<i>Tectona grandis</i>	Teak	Verbanaceae	4×4
3	<i>Melia azedarach</i>	Baken	Melaceae	4×4
4	<i>Madhuca indica</i>	Mahua	Sapindaceae	4×4
5	<i>Anthocephalus cadamba</i>	Kadamba	Rubiaceae	4×4
6	<i>Shorea robusta</i>	Sal	Depterocarpaceae	4×4
7	<i>Populus deltoids</i>	Paplar	Salicaceae	4×4
8	<i>Quercus leucotrichophora</i>	Banj	Fagaceae	4×4
9	Sole Agriculture field (Control)	--	--	--

Soil analysis

Soil physic-chemical analysis

The soil analysis was done at the laboratory of Forest Soil and Land Reclamation Division of Forest Research Institute of Dehradun. Bulk density was determined by using the core sampler technique from a 10.3 cm diameter core sampler (Blake and Hartge, 1986) [9]. To measure the particle density a measuring cylinder of 100 ml capacity was taken and filled exactly half (50 ml) with water. Then, the 10 g of soil was put into the measuring cylinder and the rise in water level was observed after 30 minutes and continues till the level became constant. The particle density was calculated by using the following formula as $PD = W/V$; where PD is particle density in g/cm^3 , W is the weight of dry soil and V is the volume of soil solution. Soil porosity was determined by the method described by (Danielson and Southerland, 1986) [16]. The pH of the soil was determined in 1:2 (soil: water) ratio after half an hour of equilibrium using glass electrode on a digital pH meter (Jackson, 1967) [36]. Electrical conductivity of the soil sample was measured in 1:2 (soil: water suspension) at 25° C using conductivity meter (Bower and Wilcox, 1965) [10]. The organic carbon content in soil was determined by modified (Walkley and Black 1934) [87] method as described by (Jackson 1967) [36]. Available nitrogen in soil was determined by alkaline potassium permanganate method (Subbiah and Asija, 1956) [78]. Available phosphorus was extracted by sodium bi-carbonate extractant (0.5 M $NaHCO_3$) adjusted to pH 8.5 as per the method of (Olsen *et al.*, 1954) [59] and developing the blue colour acid as described by (Murphy and Riley 1962) [51]. Available

Potassium in soil was determined by extraction with 1 N ammonium acetate (pH 7) and K concentration was determined by flame photometer (Perur *et al.*, 1973) [65].

Soil biological analysis

Freshly collected soil samples were kept in refrigerator at temperature $>4^{\circ}C$ till the analysis of some biological properties. The soil dilution and plate count techniques were adopted to enumerate micro flora of soil. 1g of soil was transferred to 10 ml of sterile water and serial dilutions were made. This stock solution was serially diluted to the concentrations up to 10^{-7} at which the desirable organisms showed optimum growth. This concentration was 10^{-3} to 10^{-5} for fungi, 10^{-5} to 10^{-7} for bacteria. 1 ml of the respective dilutions were spread evenly using a sterilized glass spreader on the plates. About 20-25 ml of the medium was poured in sterilized petri plates and allowed to solidify (Wollum, 1982) [90]. A nutrient medium were prepared separately according to the directions for respective micro-organisms, sterilized in autoclave at pressure 1.05 N cm^{-2} and temperature $120^{\circ}C$ for 15 minutes and was cooled up to $40^{\circ}C$. Martin's Rose Bengal Agar, Plated Count Agar and Kenkenight and Munir's medium are used for the fungi and Bacteria respectively. After the corresponding incubation period, total numbers of colonies formed were noted. Bacteria colony counts were taken after two days incubation, fungi after five days. The colony forming unit (cfu) per gram of dry soil was calculated.

Results and discussion

Soil Physical Properties

Soil bulk density

The bulk density was increased with increasing soil depth both under different agroforestry tree species as well as in the sole agriculture cropping field. The soil bulk density for all the soil depths (0-15 cm 15-30 cm as well as 30-45 cm) was significantly higher (1.41, 1.43, 1.45) in sole agriculture field as compared to under different tree species (Table 2). Among all the tree species the maximum bulk density for all the soil depths was observed in the soil collected from *Melia azedarach* (1.37, 1.40 and 1.44) based agroforestry field while lowest in *Populus deltoids* (1.25, 1.27 and 1.28). The reduction in soil bulk density under trees is attributed to the addition of organic matter through litter fall, fine root recycling, twigs etc. The findings of the present study showed the inverse relation between soil bulk density and soil organic carbon content which has been also reported earlier (Gupta and Sharma, 2008). Similarly, the significant reduction in soil

bulk density as compared to sole agricultural cropping has been reported as under the canopy of *Prosopis juliflora* (Nayak *et al.*, 2009) [55].

Soil particle density

The soil particle density was increased successively with increasing soil depth from 0-15 cm to 30-45 cm under different agroforestry tree species as well as sole agriculture cropping field. The significant reduction in soil particle density was observed in the depth of 0-15 cm only. The maximum particle density in 0-15 cm depth was observed in sole agriculture cropping field (2.85 g/cm³) while minimum under *Anthocephalus cadamba* (2.62). Similarly, the lower particle density was recorded under different agroforestry tree species as compared to the agriculture cropping field primarily due to the higher soil organic carbon content under tree species by addition of organic matter through leaf litter, twigs, pruning debris etc (Tandel *et al.*, 2009) [81].

Table 2: Soil physical properties under various tree based agroforestry systems

Treatments	Bulk Density (g/cm ³)			Particle Density (g/cm ³)			Soil porosity (%)		
	Soil Depth			Soil Depth			Soil Depth		
	0-15 cm	15-30 cm	30-45 cm	0-15 cm	15-30 cm	30-45 cm	0-15 cm	15-30 cm	30-45 cm
<i>Eucalyptus spp.</i>	1.31	1.33	2.65	2.75	2.78	2.787	55.20	53.50	53.20
<i>Tectona grandis</i>	1.30	1.35	2.73	2.77	2.78	2.78	54.46	54.03	53.06
<i>Melia azedarach</i>	1.37	1.40	2.74	2.78	2.82	2.82	55.10	52.83	52.10
<i>Madhuca indica</i>	1.36	1.40	2.74	2.76	2.8	2.8	55.80	53.83	52.43
<i>Anthocephalus cadamba</i>	1.28	1.30	2.62	2.667	2.713	2.713	56.46	55.40	53.43
<i>Shorea robusta</i>	1.27	1.33	2.69	2.72	2.76	2.76	54.90	53.70	52.83
<i>Populus deltoids</i>	1.25	1.27	2.66	2.693	2.76	2.76	55.23	54.50	52.20
<i>Quercus leucotrichophora</i>	1.27	1.32	2.72	2.75	2.797	2.797	57.60	56.86	54.60
Agriculture sole cropping	1.41	1.43	2.85	2.88	2.92	2.92	52.66	51.90	51.20
C.D.	0.062	0.062	0.099	NS	NS	N/A	2.26	1.673	NS
SEm±	0.021	0.02	0.033	0.044	0.046	0.046	0.747	0.553	0.654

Soil porosity

The soil porosity decreased with increasing soil depth from 0-15 to 30-45 cm. The soil porosity (% pore space) was significantly higher under the different agroforestry tree species as compared to the agriculture sole cropping field. Among the tree species, the highest soil porosity was observed under *Q. leucotrichophora* which was significantly higher over the other trees. The increase in soil porosity under tree species as compared to the agriculture might be due to the addition of organic matter through leaf litter and penetration of fine roots of trees in soil. Similar results were reported by (Tandel *et al.*, 2009) [81] who concluded after their studies that the soil porosity and water holding capacity improved under trees as compared to the agriculture field.

Soil chemical properties

Soil pH

The soil pH was significantly higher in sole agriculture field as compared to under tree species. Soil pH increased with increasing soil depth from 0-15 cm, 15-30 cm and 30-45 cm. The trend was common among all the tree species as well as under the agriculture (Table 3). The highest pH for the surface soil depths (0-15 cm, 15-30 cm and 30-45 cm) was under agriculture (7.06, 7.5 and 7.9), which was significantly higher over the other soil samples collected from the tree based agroforestry system. Among all the tree species, the highest pH range was observed under *T. grandis* (6.9, 7.1 to 7.4) for soil depths 0-15 cm, 15-30 cm and 30-45 cm, which was significantly higher than the other species for all depths while,

the lowest pH was recorded under *Madhuca indica*. Relatively lower pH was observed under 0-15 cm soil depth than other soil depths. This might be probably due to the leaching of salts down the soil profile and getting deposited into the deeper layers of soil. The significantly lower soil pH at 0-15 cm soil depth under different agroforestry trees than the agriculture might be due to substantial addition of organic matter to the surface soil under trees and the release of organic acid during decomposition of litter. This might also be due to the leaching of soluble salts from the surface to the deeper layers of soil. Similar results and trends of variation in soil pH under agroforestry systems in comparison to crop fields has been reported by (Prasadini and Sreemannarayana, 2007; Kumar *et al.*, 2008 and Newaj *et al.*, 2007) [69, 42, 15] also observed nominal changes in soil pH under white siris (*A. procera*) based agroforestry system after four years of experimentation as the pH value was lower compared to the initial value and it was also lower than the pH value of the Agriculture field.

Electrical Conductivity (EC)

The soil EC showed a decrease with successive soil depth. The soil EC was relatively higher under trees as compared to the agriculture field. However the difference in EC was significant for all the soil depths *i.e* for 0-15 cm 15-30 cm and 30-45 cm. Among the tree species, the maximum EC of soil was observed under *T. grandis* for all the soil depths (0.437, 0.36 and 0.32) which was significantly higher over other tree species and agriculture. The minimum EC value among all the

treatments was found in *Eucalyptus spp.*, which was significantly lower over other treatments. Soil electrical conductivity (EC) is a measurement correlating with soil properties that influence crop productivity, including soil texture, cation exchange capacity (CEC), drainage conditions, organic matter level, soil salinity and sub-soil characteristics. The EC was higher under *T. grandis* as compared to the agriculture field which could be due to enrichment of soil mineral basic salts through addition and decomposition of litter. Similar results and reasons have been reported by (Newaj *et al.*, 2007) ^[57] who also observed significantly higher values for soil EC under *A. procera* based agri-silvicultural system as compared to the agricultural field. Also contrary to this (Malik *et al.*, 1996) ^[47] observed reduction in

soil EC values by 10% under tree canopy as compared to the Agriculture area which was also observed in the present study in case of the other four species except teak for the surface soil depths.

Soil Organic Carbon (SOC)

The soil organic carbon (SOC) content was significantly higher under the tree species as compared to the agriculture field. Also, the SOC decreased with successive soil depths from 0-15 cm, 15-30 cm and 30-45 cm under all the trees. Among all the trees SOC carbon was found under *Q. leucotrichophora* (1.07, 0.90 and 0.84%) which was significantly higher over the other tree species, while lowest in *Shorea robusta* (0.92, 0.75 and 0.69%) in all soil depths.

Table 3

Treatments	Soil pH			Electric Conductivity EC (dS m ⁻¹)			Soil Organic Carbon SOC		
	Soil Depth (g/cm ³)			Soil Depth (g/cm ³)			Soil Depth (%)		
	0-15 cm	15-30 cm	30-45 cm	0-15 cm	15-30 cm	30-45 cm	0-15 cm	15-30 cm	30-45 cm
<i>Eucalyptus spp.</i>	6.70	7.03	7.20	0.26	0.21	0.20	0.93	0.77	0.71
<i>Tectona grandis</i>	6.90	7.13	7.40	0.43	0.36	0.32	1.03	0.90	0.82
<i>Melia azedarach</i>	6.86	7.13	7.30	0.35	0.30	0.27	0.97	0.83	0.78
<i>Madhuca indica</i>	6.50	6.90	6.80	0.29	0.24	0.21	0.92	0.79	0.67
<i>Anthocephalus cadamba</i>	6.86	7.00	7.00	0.36	0.34	0.29	0.96	0.80	0.74
<i>Shorea robusta</i>	6.80	6.93	7.23	0.33	0.26	0.23	0.92	0.75	0.67
<i>Populus deltoids</i>	6.60	6.73	7.00	0.33	0.28	0.24	1.02	0.85	0.73
<i>Quercus leucotrichophora</i>	6.70	6.93	7.06	0.35	0.31	0.27	1.07	0.90	0.84
Agriculture (Open)	7.06	7.56	7.90	0.25	0.21	0.19	0.75	0.63	0.60
C.D.	0.228	0.354	0.305	0.016	0.012	0.012	0.037	0.038	0.031
SEm±	0.075	0.117	0.101	0.005	0.004	0.004	0.012	0.013	0.01

The soil enrichment in SOC content under tree based systems might be because of several factors such as addition of litter, annual recycling of fine root biomass and root exudates and its reduced oxidation of organic matter under tree shades (Gill and Burman, 2002) ^[25]. The results obtained in the present study are in conformity with the findings reported by (Pingale, 2009 and Ghimire, 2010) ^[68]. There was a decrease in SOC content of soil with increasing soil depth and the highest SOC was observed 0-15 cm soil depth for all the treatments. This may be attributed mainly to the contributions made by litter fall at the surface layer. Similar variation of SOC with increasing soil depth has been observed by some investigators (Swamy and Puri, 2005; Chauhan *et al.*, 2010 and Ghimire, 2010) ^[80, 14, 23]. Higher SOC content are observed under tree + crop systems not only in surface but in all the soil depths as compared to the Agriculture and uncultivated lands. This can be attributed to the recycling of organic matter through roots in the layers they occur. When the decomposition of a root residue takes place, they supply nutrients to the soil through the process of mineralization and also contribute to the addition of carbon in the soil through the humification process. Several researchers have reported that root biomass addition in an agroforestry system is generally higher than agriculture or agriculture fields (Sharma *et al.*, 2009) ^[14]. Also, trees generally have lignified cells in its plant parts such as litter, small branches, bark, roots etc. which may lead to the biochemical stabilization of organic carbon in the soil and hence leads to the improvement in SOC content of soil under agroforestry as concluded by (Six *et al.*, 2002) ^[77]. Therefore, one of the reasons for lower concentrations of SOC under both the Agriculture field

(without tree) is attributed to the lack of lignified cells in agricultural residues. Further, the large scale tillage and other Cultural operations could be a probable cause to reduce the SOC content of soil under Agriculture with full exposure to sunlight.

Available Nitrogen (N)

The available soil nitrogen (kg/ha) was significantly influenced by different tree species and soil depths (Table 4). The available soil nitrogen was significantly higher under tree species as compared to the Agriculture field. This showed that the availability of nitrogen in soil decreased with successive increase in soil depth from 0-15 cm, 15-30 cm and 30-45 cm under all the treatments. Highest N content in soil was recorded under *Q. leucotrichophora* which was significantly higher than other tree species while lowest under *Tectona grandis*. The available N content in soil increased in different tree species under agro-forestry over the Agriculture field which is attributed to the addition of organic matter in soil in the form of litter fall and fine root biomass. The release of nutrient into the soil through the process of mineralization of organic matter leads to an increase in the nutrient status of soil (Osman *et al.*, 2001) ^[60]. (Chaudhry *et al.*, 2007) ^[13] also reported similar results as obtained in the present study after his studies on poplar based agroforestry systems. The highest amount of available N was found in surface soil *i.e.* 0-15 cm due to more turn-over of the organic residues on the surface layer of soil which decreased with depth of soil. The lower most layer (60-90 cm) had the lowest available N content in soil. (Bhardwaj *et al.*, 2001) ^[7] also observed a decreasing trend in available N content in soil with an increase in soil depth under high density poplar plantation. Some other

researchers also observed similar trends during their studies on soil properties under poplar based agroforestry systems (Ghimire, 2010 and Swami *et al.*, 2006).

Available Phosphorus (P) in soil under different tree species and agriculture

The available phosphorus content (kg/ha) in soil showed a decreasing trend with increase in successive soil depth for all the treatments. The available P in soil was significantly higher under different tree species over the agriculture. The maximum available P content for soil depth 0-15 cm, 15-30 cm and 30-45 cm was observed under *Q. leucotrichophora* (26.56 kg/ha, 21.54 kg/ha and 17.59 kg/ha) respectively which was relatively higher as compared to the other tree species and agriculture followed by *T. grandis* (26.41 kg/ha, 20.96 kg/ha and 16.67 kg/ha). The minimum available P was observed under Agriculture (Open) in all the depths

(16.14kg/ha, 14.44 kg/ha and 12.10kg/ha) which was significantly lower as compared to the other treatments. The highest available P was observed in *Tectona grandis* for all the soil depths (26.41, 20.96 and 16.67 kg/ha) which was significantly higher than other tree species while lowest in agriculture sole cropping system (16.14, 14.44 and 12.10 kg/ha). This might be due to higher activity of acidic phosphatase enzyme at these soil depths under this species over the other treatments, as the organic anion exudation and acid phosphatase activity may lead to an increase in the mobilization of P in rhizosphere. The available P decreased with an increase in soil depth in the present investigation which is in conformity with the findings of (Swami *et al.*, 2006; Majumdar *et al.*, 2004 and Ghimire, 2010) [46, 23] who also observed similar trends on studying the soil properties under poplar based agroforestry system.

Table 4

Treatments	N (Kg/h)			P (Kg/h)			K (Kg/h)		
	Soil Depth			Soil Depth			Soil Depth		
	0-15 cm	15-30 cm	30-45 cm	0-15 cm	15-30 cm	30-45 cm	0-15 cm	15-30 cm	30-45 cm
<i>Eucalyptus spp.</i>	324.56	291.47	270.96	18.09	15.01	13.42	153.67	143.43	122.35
<i>Tectona grandis</i>	258.12	245.29	221.93	26.41	20.96	16.67	151.45	141.25	120.65
<i>Melia azedarach</i>	272.54	242.60	215.47	24.35	18.68	14.18	166.13	156.33	128.82
<i>Madhuca indica</i>	312.03	290.87	311.96	17.97	15.48	13.01	148.84	138.81	122.60
<i>Anthocephalus cadamba</i>	337.04	324.93	315.96	21.87	16.49	13.35	162.81	152.42	129.72
<i>Shorea robusta</i>	333.04	323.93	268.50	26.00	19.10	15.20	154.56	144.36	118.29
<i>Populus deltoids</i>	306.34	280.32	257.71	24.87	17.37	13.90	148.67	138.58	124.78
<i>Quercus leucotrichophora</i>	357.26	345.15	332.18	26.56	21.54	17.59	167.56	157.26	132.26
Agriculture (Open)	232.73	217.61	198.12	16.14	14.44	12.10	135.68	125.62	118.43
C.D.	17.54	8.67	13.27	1.129	0.88	0.65	7.65	5.88	6.15
SEm±	5.80	2.87	4.39	0.373	0.291	0.22	2.532	2.31	2.034

Available Potassium (K) in soil under different tree species and agriculture

The available potassium (kg/ha) in soil decreased with increasing soil depth (Table 5) under all the tree species as well as agriculture. The available K content in soil was significantly higher under trees as compared to the agriculture sole cropping field. The depths (0-15 cm, 15-30 cm and 30-45 cm) available K was highest in *Q. leucotrichophora* (167.56, 167.56 and 132.26 kg/ha) respectively, which was significantly higher over the other tree species while, lowest sole agriculture crop field (135.68, 135.68 and 118.43 kg/ha). The surface soil (0-15 cm) had higher level of available K in soil in comparison to the sub-surface soil (15-30 and 30-45 cm) and decreased with successive increase in soil depth. This trend was found for all the tree species as well as agriculture. This was probably as a consequence of higher amount of organic matter at the surface layer due to higher litter fall and fine root turn over at the surface layer of soil than the sub-surface layer. Also, higher mobility of potassium at the surface layer could be another cause for higher K content in surface soil than sub-surface and deeper layers of soil. Similar decrease in soil available K content in soil with increase in soil depth have been observed by several investigators like (Bhardwaj *et al.*, 2001; Swamy *et al.*, 2006; Mishra and Swamy, 2007) [7, 51, 55, 21]. The available soil potassium was higher under alley cropping system as compared to the Agriculture field as a result of release of organic acids due to

organic matter accumulation under agroforestry and ultimately resulting in higher mineralization of potassium has been reported by (Miah *et al.*, 2001) [48]. Such findings have also been conferred by (Bajpai *et al.*, 2006) [4] who reported higher K content in soil as a result of higher organic matter. Availability of potassium is also increased under agroforestry as compared to treeless farming systems because of enhanced recycling of nutrients through bio-chemical process as reported by (Hasan and Ashraful Alam, 2006) [31].

Soil biological properties under different agroforestry species and agriculture

Among different tree species bacterial population was significantly highest (62.67×10^6 cfu g⁻¹ soil) in surface and sub-surface soil (33.33×10^6 cfu g⁻¹ soil) under *Q. leucotrichophora*. The minimum bacterial population was in both surface and sub-surface soil in Agriculture (35.00×10^6 cfu g⁻¹ soil) and (13.00×10^6 cfu g⁻¹ soil). The number of bacteria count in soil decreased with increasing soil depth under all the tree species as well as agriculture. All the tree species has significantly higher microbial population in the soil as compared to the agriculture field in surface as well as sub-surface soil horizon. Bacterial population in soil within range as reported by (Whitman *et al.*, 1998) [88]. They had reported that bacterial population in different soils ranged between 4-106 to 2-109 g⁻¹ dry soil.

Table 5: Soil bacteria (no of colony per g soil $\times 10^6$ cfu) and soil fungi (no of colony per g soil $\times 10^4$ cfu) under different tree species and agriculture field.

Treatments	Bacterial colony		Fungal Colony	
	Soil Depth		Soil Depth	
	0-15 cm	15-30 cm	0-15 cm	15-30 cm
<i>Eucalyptus spp.</i>	53.00	17.67	82.33	25.00
<i>Tectona grandis</i>	56.67	25.67	100.00	34.33
<i>Melia azedarach</i>	46.00	19.00	61.33	19.00
<i>Madhuca indica</i>	55.33	19.67	57.33	28.67
<i>Anthocephalus cadamba</i>	62.00	29.67	120.33	39.67
<i>Shorea robusta</i>	58.33	26.33	72.33	22.33
<i>Populus deltoids</i>	50.33	26.00	88.00	19.33
<i>Quercus leucotrichophora</i>	62.67	33.33	123.67	58.67
Agriculture (Open)	35.00	13.00	45.67	12.33
C.D.	1.98	1.16	1.94	1.39
SEm \pm	0.66	0.38	0.64	0.46

Fungi in soil under different tree species and agriculture

The fungal population in soil ranged between 58.67 to 123.67 $\times 10^4$ cfu. g⁻¹ soil and 12.33 to 45.67 $\times 10^4$ cfu g⁻¹ soil at the surface (0-15 cm) and sub-surface (15-30 cm) soil, respectively. The fungal population, irrespective of agroforestry system and agriculture, decreased significantly with soil depth. All the tree species has significantly higher fungal population as compared to the agriculture. This may be due to the high availability of fresh litter or/and root exudates at the soil surface to select for microbial communities that are able to rapidly utilize these labile carbon substrates. The microbial community residing in the deeper soil horizons are more severely resource limited than their surface-dwelling counterparts. Similar result was shown by (Taylor *et al.*, 2002) [82] who reported that the fungi were isolated from surface soils but were absent from the deep soil samples. Decrease in organic matter content in sub-soil sample caused the lower number of fungal population in sub-surface soil. The surface soil has the highest species richness whereas sub-surface soil of barren sand dunes shows lowest richness. The results obtained during this course work pertaining to soil microbial biomass carbon (MBC) got significant support from the findings of earlier workers (Kaur *et al.*, 2000) [39] who had also reported that the rate of mineralization of soil microbial biomass carbon increased appreciably under agroforestry systems. An increased proportion of microbial carbon in the total soil organic pool indicate higher nutrient availability to the plants in agroforestry systems as compared to sole cropping. Similarly (Yadav *et al.*, 2011) [14] observed that soil microbial biomass carbon was higher ranging from 262-320 $\mu\text{g g}^{-1}$ under agroforestry corresponding to lower soil microbial biomass C (186 $\mu\text{g g}^{-1}$) under treeless control. The probable reason being the different organic inputs from trees in the form of litter fall, recycling of fine root biomass and pruning debris contributed significantly towards organic matter pool under agroforestry enhancing the microbial population and mineralization rate of carbon. Other researchers (Campbell *et al.*, 1991; Munna *et al.*, 2007) [12, 50] observed similar trends of decrease in microbial biomass C in soil with increasing soil depths.

References

- Amponsah I, Meyer W. Soil characteristics in teak plantations and natural forests in Ashanti region, Ghana. Communications in Soil Science and Plant Analysis. 2000; 31:355-373.
- Annon, Soil and Water Conservation Research, 1956-71. ICAR, New Delhi, 1976.
- Aweto AO, Dikinya O. The beneficial effects of two tree species on soil properties in a semi-arid savannah rangeland in Botswana. Land Contamination & Reclamation. 2003; 11(3):339-344.
- Bajpai RK, Chitale S, Upadhyay SK, Upkumar JS. Long term studies on soil physic-chemical properties and productivity of rice-wheat system as influenced by integrated nutrient management in inceptisols of Chattisgarh. Journal of Indian Society of Soil Science. 2006; 54(1):24-29.
- Balagopalan M, Jose AI. Soil chemical characteristics in a natural forest and adjacent exotic plantations in Kerala, India. Journal of Tropical Forest Science. 1995; 8:161-166.
- Balagopalan M, Thomas TP, Mary MV, Sankar S, Alexander TG. Soil properties in teak, bombax and eucalyptus plantations of Thrissur liorest Division, Kerala. Journal of Tropical Forest Science. 1992; 5:35-43.
- Bhardwaj SD, Panwar P, Gautam S. Biomass production potential and nutrient dynamics of *Populus deltoides* under high density plantations. Indian Forester. 2001; 127(2):144-153.
- Biswas S, Ghoshal SK, Sahoo SS, Mukherjee D. Some soil properties under agroforestry in Gangetic alluvial tract of West Bengal. Environment and Ecology. 2003; 21(3):562-567.
- Blake GR, Hartge KH. Methods of soil analysis. American Society of Agronomy. Inc. Publisher, Madison, U.S.A, 1986.
- Bower CA, Wilcox LV. Methods of soil analysis. American Society of Agronomy. Inc. Publisher, Madison, Wisconsin, USA, 1965.
- Brookes PC. The soil microbial biomass: concept, measurement and applications in soil ecosystem research. Microbes Environ. 2001; 16:31-140.
- Campbell CA, Beiderbeck BO, Zenter RP, lafond GP. Effect of crop rotations and cultural practices on soil organic matter, soil microbial biomass and respiration in a thin Black Chernozom. Canadian Journal of Soil Science. 1991; 71:363-376.
- Chaudhry AK, Khan GS, Ahmad I. Effect of poplar tree intercropping at various densities on the post-harvest soil nutrient contents. Pakistani Journal of Agricultural Sciences. 2007; 44(2):468-472.
- Chauhan SK, Sharma SC, Beri V, Ritu Yadav S, Gupta N. Yield and carbon sequestration potential of wheat (*Triticum aestivum*)-poplar (*Populus deltoides*) based

- agri-silvicultural system. *Indian Journal of Agricultural Sciences*. 2010; 80(2):129-135.
15. Chavan SB, Keerthika A, Dhyani SK, Handa AK, Newaj R, Rajarajan K. National Agroforestry Policy in India: a low hanging fruit. *Current Science*. 2015; 10:1826-1834.
 16. Danielson RE, Southerland PL. Porosity In: *Methods of soil analysis Part 1*. American Society of Agronomy and Soil Science Society of America. Madison, Wisconsin, U.S.A, 1986.
 17. Deshpandey SB, Feheronbacher JB, Roy BW. Mollisol of tarai region of Uttar Pradesh, Northern India, 2. Genetics and classification. *Geoderma*. 1971; 6:195-211.
 18. Dhyani SK, Handa AK. Area under agroforestry in India: an assessment for present status and future perspective. *Indian Journal of Agroforestry*. 2013; 15:1-10.
 19. Dick RP. A review: Long-term effects of agricultural systems on soil biochemical and microbial parameters. *Agricultural Ecosystems*. 1992; 4:25-36.
 20. Don- Scott H. *Soil physics: Agricultural environmental application*. Iowa: Iowa State University Press, 2000.
 21. Dwivedi AP. *A Textbook of Silviculture International Book Distributors, Dehradun, India, 2004, 146-148.*
 22. FSI. *India State of Forest Report 2013, Forest Survey of India, (Ministry of Environment & Forests), Dehradun, India, 2013.*
 23. Ghimire TB. Effect of fertility levels on mustard (*Brassica juncea L.*) productivity under varyinf poplar tree densities. Ph.D. Thesis. G.B. Pant University of Agriculture and Technology, Pantnagar- 263 145, Uttarakhand, 2010, 306.
 24. Ghosh RC. The protective role of forestry to the land. 18th Commonwealth Forestry Conference, London, 1974.
 25. Gill AS, Burman D. Production management of field crops in agroforestry systems. In: *Recent advances in Agronomy*. (Singh, G., Kolar, J.S.W. and Sekhon, H.S. Eds.) New Delhi, Indian Society of Agronomy, 2002, 523-542.
 26. Gomez KA, Gomez AA. *Statistical procedure for agricultural research (2nd ed.)*. John Wiley and Sons. Inc. New York, 1984, 80.
 27. Gupta MK. Soil Organic Carbon Pool under Different Land Uses in Haridwar District of Uttarakhand. *Indian Forester*. 2011; 137(1):105-112.
 28. Gupta MK, Sharma SD. Effect of tree plantation on soil properties, profile morphology and productivity index in Uttarakhand. *Annals of Forestry*. 2008; 16(2):209-224.
 29. Gupta N, Kukal SS, Bawa Dhaliwal GS. Soil organic carbon and aggregation under poplar based agroforestry system in relation to tree age and soil type. *Agroforestry Systems*. 2009; 76A:27-35.
 30. Hanway JJ, Heidel H. *Soil analysis methods as used in Iowa State College of Soil Testing Laboratory*. Iowa Agriculture. 1952; 57:1-31.
 31. Hasan MK, Ashrafal Alam AKM. Land Degradation Situation in Bangladesh and Role of Agroforestry. *Journal of Agriculture and Rural Development*. 2006; 4(1 & 2):19-25.
 32. Hosur GC, Dasog GS. Effect of tree species on soil properties. *Journal of the Indian Society of Soil Science*. 1995; 43(2):256-259.
 33. ICRAF. *The Future of Agroforestry*. World Agroforestry Centre, Nairobi Kenya, 2006.
 34. Imoro ZA, Tom-Dery D, Kwadwo KA. Assessment of soil quality improvement under Teak and Albizia. *Journal of Soil Science and Environmental Management*. 2012; 3(4):91-96.
 35. Jackinson DS, Powlson DS. The effects of biological treatments on metabolism in soil. A method for measuring soil biomass. *Soil Biological Function*. 1976; 8:209-213.
 36. Jackson ML. *Soil chemical analysis*. Prentice Hall Pvt. Ltd. New Delhi, India, 1967, 498.
 37. Jaiycoha IA. Changes in soil properties related to different land uses in part of the Nigerian semi-arid savannah. *Soil Use and Management*. 1995; 1(1):84-89.
 38. Joshi G, Negi CGS. Physic-chemical properties along soil profiles of two dominant forest types in Western Himalaya. *Research Communications*. 2015; 1:798-803.
 39. Kaur B, Gupta SR, Singh G. Soil carbon, microbial activity and nitrogen availability in agroforestry systems on moderately alkaline soils in Northern India. *Applied Soil Ecology*. 2000; 15:283-294.
 40. Kitteredge J. *Forest influences*. McGraw-Hill Book Company, New York, 1948.
 41. Kumar BM. Agroforestry: the new old paradigm for Asian food security. *Journal of Tropical Agriculture*. 2006; 44(1-2):1-14.
 42. Kumar K, Laik R, Das DK, Chaturvedi OP. Soil microbial biomass and respirationin afforested calciorthent. *Indian Journal of Agroforestry*. 2008; 10(2):75-83.
 43. Laik R, Koushendra K, Das DK. Organic carbon and nutrient build-up in calciorthant soil under 6 forest tree species. *Forests, Trees and Livelihood*. 2009; 19(1): 81-92.
 44. Leakey R. Definition of agroforestry ICRAF, Nairobi (Kenya) International Centre for Research in Agroforestry (ICRAF), Nairobi (Kenya), 1996.
 45. Leakey RRB. Definition of agroforestry revisited. *Agroforestry Today*. 1996; 8(1):5-7.
 46. Majumdar B, Kumar K, Venkatesh MS, Patriam Bhatt BP. Effect of different agroforestry systems on soil properties in acidic Alfisols of Meghalaya. *Journal of Hill Research*. 2004; 17(1):1-5.
 47. Malik KPS, Prakash O, Kohli RK, Arya KS. Change in some physical and chemical properties of soil in an agri-silvicultural system with some multipurpose tree spp. in Tarai region of Uttar Pradesh. In: *Proc. IUFRO-DNAES International Meeting: Resource Inventory Techniques to Support Agroforestry and Environment (Eds)*, 1996, 249-254.
 48. Miah MG, Rahman MA, Haque MM, Ahmed NU, Islam MH. Crop productivity and soil fertility in Alley ropping system at different nitrogen levels in upland ecosystem. In "National workshop on Agroforestry research and development in Bangladesh" (M. F. Haq, M. K. Hasan, S. M. Asaduzzaman, and M. Y. Ali (Eds.), held on September, BARI, Gazipur, 2001.
 49. Mishra A, Swamy S. Eco-physiology, productivity and nutrient uptake of soyabean under *Populus deltoides* based agri-silviculture system. *Journal of Soils and Crops*. 2007; 17(2):217-224.
 50. Munna R, Anirudha Y, Yadav RS. Effect of Anjan (*Hardwickia binnata* Roxb.) based agroforestry system on soil fertility. *Range Management and Agroforsetry*. 2007; 28(2A):95-97.
 51. Murphy J, Riley JP. A modified single solution method for determination of phosphorous in natural waters. *Annals Chem. Acta*. 1962; 27:31-36.

52. Musoz C, Zagal E, Ovalle C. Influence of trees on soil organic matter in Mediterranean agroforestry systems: an example from the Espinal of central Chile. *European Journal of Soil Science*. 2007; 58:728-735.
53. Nair PKR. *Soil Productivity Aspects of Agroforestry*. ICRAF, Nairobi, 1984.
54. Nair PKR. *An Introduction to Agroforestry*. Springer (India) Pvt. Ltd., New Delhi, 2008.
55. Nayak AK, Khan U, Sharma DK, Mishra VK, Sharma CJ, Singh R, Singh G. Spatial variability of soil physico-chemical properties under *Prosopis juliflora* and *Terminalia arjuna* in sodic soil of Indo-gangetic plains. *Journal of Indian Society of Soil Science*. 2009; 57(1):31-38.
56. Nazir T, Samweel N. Soil Nutrient Status under Pine and Oak forests in temperate valley slopes of Garhwal Himalayas, India. *International Journal of Current Research*. 2013; 5(12):4309-4315.
57. Newaj R, Dar SA, Bhagwan MK, Yadav RS. Effect of management practices on growth of white siris (*Albizia procera*), grain yield of intercrops, weed population and soil fertility changes in agri-silvicultural system in semi-arid India. *Indian Journal of Agricultural Sciences*. 2007; 77(7):403-407.
58. Okoro P, Aighewi IT, Osagie CO. Effects of selected monoculture plantation species on the humid tropical soils of Southern Nigeria. *Nigerian Journal of Forestry*. 1999; 29:73-79.
59. Olsen SR, Cole CV, Waterabe FS, Dean LA. Estimation of available phosphorus in soil by extraction with sodium carbonate. In: Black, C.A. (Ed) *Methods of soil analysis*, part 2. American society of Agronomy Inc. Publisher, Medison, Wisconsin, USA, 1954, 1044-1046.
60. Osman KT, Rahman MM, Barua P. Effects of some forest tree species on soil properties in Chittagong University Campus, Bangladesh. *Indian Forester*. 2001; 127(4):431-442.
61. Pande PK. Temporal variations in soil nutrients under tropical plantations. *Annals of Forestry* 2004; 12:29-37.
62. Pandey CB, Singh AK, Sharma DK. Soil properties under *Acacia nilotica* trees in a traditional agroforestry system in central India. *Agroforestry Systems*. 2000; 49:53-61.
63. Panse VG, Sukhatme PV. *Statistical methods for agricultural workers*, 3rd ed., ICAR publications, New Delhi, 1978, 347.
64. Pansu M, Gaatheyrou J, Loyer JY. *Soil analysis-sampling, instrumentation and quality control*. Balkema, Lisse, Abington, Exton, Tokiyo, 2001, 489.
65. Perur N, Mehar G, Roy H. Soil fertility evaluation to serve Indian farmers (Bulletin). Department of Agriculture, Mysore, 1973.
66. Rao SA, Srivastava S. Role of plant nutrition in increasing crop productivity. *Fertilizer News*. 1998; 43:65-75.
67. Patra AK. *Agroforestry Principles and Practices*. New India Publishing Agency, New Delhi, India, 2013..
68. Pingale BN. Studies on Carbon Sequestration in poplar (*Populus deltoides* Bartz. Ex. Marsh) based agroforestry system with varying tree density. M.Sc. Ag. (Agroforestry) Thesis. G.B. Pant University of Agriculture and Technology, Pantnagar- 263 145, Uttarakhand, 2009, 119.
69. Prasadini P, Sreemannarayana B. Impact of agroforestry systems on nutritional status and biological activity on rain fed and sandy loam soils. *Indian Forester*. 2007; 133(11):1519-1525.
70. Richards AE, Dalal RC, Schmidt S. Soil carbon turnover and sequestration in native subtropical tree plantations. *Soil Ecology and Biochemistry*. 2007; 39:2078-2091.
71. Sanchez PA. *Science in agroforestry*. *Agroforestry Systems*. 1995; 30:5-55.
72. Satyawali K. Studies on biomass accumulation, carbon storage and nutrient budgeting in high density *Eucalyptus* and *Melia* plantation. M.Sc. Thesis. G.B. Pant University of Agriculture and Technology, Pantnagar- 263 145, Uttarakhand, 2014, 158.
73. Sharma KL, Raju KR, Das SK, Rao Prasad BRC, Kulkarni BS, *et al.* Soil Fertility and Quality Assessment under Tree-crop and Pasture-Based Land-Use Systems in a Rainfed Environment. *Communications in Soil Science and Plant Analysis*. 2009; 40(9-10):1436-1461.
74. Shukla AK, Misra PN. Improvement of sodic soil under tree cover. *Indian forester*. 1993; 119:43-52.
75. Singh EN, Nungchim A, Singh SS, Tiwari SC. Influence of *Tectona grandis* and *Dalbergia grandiflora* on soil properties in humid tropics of Arunachal Pradesh. *Indian Journal of Forestry*. 2001; 24(2):135-142.
76. Singh RR, Hooda MS. Growth of multipurpose tree species in Haryana, India. *Nitrogen Fixing Tree Research Reports*. 1992; 10:57-58.
77. Six J, Conant RT, Paul EA, Paustan K. Ssailization mechanism of soil organic matter, implication for carbon saturation in soil. *Plant and Soil*. 2002; 241:155-176.
78. Subbiah BV, Asija GL. A rapid procedure for estimation of available N in soil. *Current Science*. 1956; 25:259-260.
79. Swamy SL, Mishra A, Puri S. Comparison of growth, biomass and nutrient distribution in five promising clones of *Populus deltoides* under an agri-silvicultural system. *Bioresource Technology*. 2006; 97:57-68.
80. Swamy SL, Puri S. Biomass production and C-sequestration of *Gmelina arborea* in plantation and agroforestry system in India. *Agroforestry systems*. 2005; 64(3):181-195.
81. Tandel MB, Kukaldia BN, Kolambe Jadeja DB. Influence of tree cover on physical properties of soil. *Indian Forester*. 2009; 135(5):420-424.
82. Taylor JP, Wilson B, Mills MS, Burns RG. Comparison of microbial numbers and enzymatic activities in surface soils and sub soils using various techniques. *Soil Biology and Biochemistry*. 2002; 34(3):387-401.
83. Toky OP, Bisht RP, Singh SP. Growth and biomass allocation in six multipurpose nitrogen fixing trees of arid regions of India. *Nitrogen Fixing Tree Research Reports*. 1992; 10:59-61.
84. Tornquist CG, Hons FMF, eagley SE, Hagger J. Agroforestry system effects on soil characteristics of the Sarapiquõ region of Costa Rica. *Agriculture, Ecosystems and Environment*. 1999; 73:19-28.
85. Troop RS. *Silviculture of Indian Trees*. Vol. III. Clarendon Press, Oxford, U.K, 1921.
86. Voroney RP, Winter JP, Beyert RP. Soil microbial biomass C and N. In M. R. Carter (*et al.*) *Soil sampling and methods of analysis*. Lewis, Chelses, 1993, 277-286.
87. Walkley A, Black IA. An examination of Degtjareff method for determining soil organic matter and a proposed modification of the chromic acid titration method. *Soil Science*. 1934; 37:29-38.

88. Whitman WB, Coleman DC, Wiebe WJ. Prokaryotes: the unseen majority. *Proceedings of the National Academy of Sciences*. 1998; 95(12):6578-6583.
89. Wiersum KF. Surface erosion under various agroforestry systems. In *Proc. Symp. Effects of forest land use on erosion and slope stability*. Environment & Policy Institute East-West Center, Honolulu, 1984, 231-239.
90. Wollum AG. Cultural Methods for Soil Microorganisms 1. Methods of soil analysis. Part 2. Chemical and microbiological properties, (method sofsolan2), 1982, 781-802.
91. Yadav R, Yadav B, Chhipa B, Dhyani S, Ram Munna. Soil biological properties under different tree based traditional agroforestry systems in a semi- arid region of Rajasthan, India. *Agroforestry Systems*. 2011; 81(3):195-202.
92. Yan T, Yang L, Campbell CD. Microbial biomass and metabolic quotient of soils under different land use in the Three Gorges Reservoir area. *Geoderma*. 2003; 115:129-138.
93. Young A. *Agroforestry for soil management*. CAB International, Walling Ford, U.K, 2005.