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## Role of bio-fertilizers towards sustainable agricultural development: A review

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**Abstract**

The worldwide increase in human population every year raises a major threat to the food security of the people as the land for agriculture is restricted and even drastic reduction with time. Therefore, it is essential that agricultural productivity should be enhanced significantly within the next few decades to meet the large demand of food by emerging population. Not to point out too much dependence on chemical fertilizers for more crop productions certainly damages both ecosystem and human health with great severity. Bio-fertilizers are one of the greatest nature gifts of our agricultural science as a replacement to chemical fertilizers. Bio-fertilizer contains microorganisms which encourage the adequate supply of nutrients to the host plants and ensure their proper development of growth, development and regulation in their physiology. Living microorganisms are used in the preparation of bio-fertilizers which have specific functions to augment plant growth and reproduction. Bio-fertilizers being essential components of sustainable farming play vital role in maintaining long term soil fertility and sustainability of crop production.

**Keywords:** Biofertilizers, Microorganisms soil fertility, crop production

**1. Introduction**

The World population is now 7.6 Billion and India alone contains 1.35 Billion people, which is increasing day by day. This imparts pressure on the agricultural lands and other resources which are needed for food of this huge population. According to 15th Census of India in 2011, the population decadal growth of 17.64% was observed of which around 68.84% is rural population. This growing human population demands conventional agriculture to meet its needs of food which makes farmers to depend on usage of chemical fertilizers and pesticides for increased productivity (Santos *et al.* 2012) [49]. The chemical fertilizers are made in industries with known composition of various nutrients like nitrogen, phosphorus, potassium, sulfur *etc.* Harmful effects of usage of such fertilizers include weakening of roots of plants, increase of disease incidence, soil acidification (Chun-Li *et al.* 2014) [11] and eutrophication of ground water and other water bodies (Youssef and Eissa, 2014) [64]. Nutrients like nitrates leach to ground water and cause Blue Baby Syndrome also called 'acquired methemoglobinemia' (Knobeloch, *et al.* 2000). Such chemicals will have a great impact on the future generations. In this regard, eco-friendly approaches are gaining popularity with a view of bio-safety of which Biofertilizers play a major role in sustainable agriculture. The global market for biofertilizers is expected to exceed a market worth of USD 10.2 billion by 2018. European and Latin American countries are the leading consumers of biofertilizers, due to the stringent regulations imposed on the usage of chemical fertilizers, eventually getting replaced by biofertilizers (Raja, 2013) [45].

Biofertilizers may be defined as "substances which contain living microorganisms that colonize the rhizosphere or the interior of the plants and promote growth by increasing the supply or availability of primary nutrients to the target crops, when applied to soils, seeds or plant surfaces". (Mazid *et al.* 2011) [35]. They colonize in rhizosphere accompanying interior of the plant and stimulates growth by increasing the accessibility and uptake of mineral nutrients to the host plant (Malusa *et al.* 2012) [32]. They are the preparations containing cells of microorganisms which may be N fixers, P solubilizers, S-oxidisers or organic matter decomposers. They are "eco-friendly" agro-input of organic origin and work on conversion of unavailable essential elements to available form through their routine metabolic activities. (Vessey, 2003) [61]. Plant utilizes the only 10% - 40% of applied nutrients, the rest of 60-90% is lost in the form of immobilization, leaching, volatilization *etc.* Biofertilizers help in slow and continuous release of nutrients by their metabolism and thus form an important component of Integrated Nutrient Management (INM) system to sustaining agricultural productivity and a

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healthy environment (Adesemoye and Kloepper, 2009) <sup>[1]</sup>

**Table 1:** Different groups of biofertilizers (Barman *et al.* 2017) <sup>[5]</sup>

| Sl. No.  | Groups                 | Examples   |
|--|------------------------|--|
| <b>1. Nitrogen (N<sub>2</sub>) fixing Bio fertilizers</b>              |                        |  |
| i.   | Free living            | Azotobacter, Clostridium, Anabaena, Nostoc   |
| ii.  | Bio Symbiotic          | <i>Rhizobium</i> , <i>Frankia</i> , <i>Anabaena azollae</i>                              |
| iii.   | Associative-Symbiotic  | <i>Azospirillum</i>  |
| <b>2. P-Solubilizing Bio fertilizers</b>                               |                        |  |
| i.   | Bacteria               | <i>Bacillus subtilis</i><br><i>Pseudomonas striata</i>                                   |
| ii.  | Fungi                  | <i>Penicillium sp.</i> , <i>Aspergillus awamori</i>                                      |
| <b>3. P-Mobilizing Bio fertilizers</b>                                 |                        |  |
| i.   | Arbuscular Mycorrhizae | <i>Glomussp.</i> , <i>Gigasporasp.</i>   |
| ii.  | Ectomycorrhiza         | <i>Laccarai sp.</i> , <i>Pisolithus sp.</i> , <i>Boletus sp.</i> ,<br><i>Amanita sp.</i> |
| iii.   | Ericoid Mycorrhiza     | <i>Pezizellaericae</i>   |
| <b>4. K- solubilizers</b>  |                        |  |
| i.   | Bacteria               | <i>Frateuria aurantia</i>  |
| <b>5. Micronutrient solubilizers (Silicate and Zinc soloubilizers)</b> |                        |  |
| i.   | Bacteria               | <i>Bacillus sp.</i>  |
| <b>6. Plant Growth Promoting Rhizobacteria</b>                         |                        |  |
| i.   | Bacteria               | <i>Pseudomonas fluorescens</i>   |

### 1.1 Role of Bio fertilizers

Biofertilizers show plant growth promoting qualities and increase the yield by various mechanisms like nitrogen fixation, P-solubilization, P-mobilization, K solubilization, micronutrient solubilization, plant growth promotion, preventing the depletion of the soil organic matter (Jeyabal and Kupuswamy, 2001) <sup>[24]</sup> and maintenance of the natural habitat of the soil.

**Table 2:** Recommendations of different Bio-fertilizer for various crops

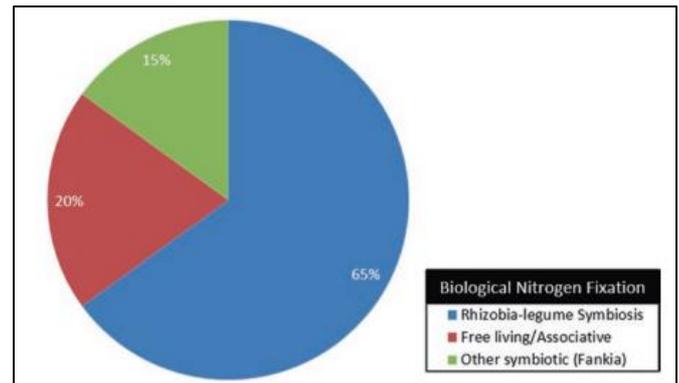
| Biofertilizer    | Recommended Crops   |
|------------------|---|
| Rizobium         | Pluses, Oilseeds, Fodders   |
| Azospirillum     | Rice, Wheat, millets, maize, sorghum, sugarcane                                       |
| Azotobacter      | Rice, Wheat, millets, other cereals, cotton, vegetables, sunflower, mustered, flowers |
| Azolla           | Submerged rice with maximum temperature   |
| Blue Green algae | Submerged rice  |
| PSM              | All crops   |

Source: FAI, 2006-07

#### 1.1.1 Nitrogen fixation

Certain bacteria and algae elemental, atmospheric nitrogen into plant usable forms like ammonical and nitrate forms. This process is called Biological nitrogen fixation (BNF) (Gothwal

*et al.* 2007) <sup>[18]</sup> This mechanism is exploited by utilizing such microorganisms in the form of potent biofertilizers which act as a substitute for chemical fertilizers, help in maintenance of soil nitrogen reserves and give large crop yields (Peoples and Craswell, 1992) <sup>[41]</sup> They can be grouped into free-living bacteria like *Azotobacter*, associative like *Azospirillum*, cyanobacteria and symbiotic organisms like *Rhizobium*, *Frankia* and *Azolla* (Gupta, 2004) <sup>[20]</sup> Symbiotic associations are the maximum contributors for BNF in nature (Fig. 1). Apart from *Azotobacter*, many other genera fix nitrogen. Latt and others (2018) isolated and identified six free living N-fixing bacteria belonging to the genera *Azotobacter*, *Alcaligenes* and *Azomonas* by 16s rRNA sequencing. Certain genera are discussed here.



**Fig 1:** Biological nitrogen fixation (Modified from Bouizgarne *et al.* 2015) <sup>[8]</sup>

##### 1.1.1.1 Azotobacter

*Azotobacter* is an aerobic, gram-negative, heterotrophic, free living nitrogen fixing bacteria. With the capacity of independent BNF, it has been found that some *Azotobacter* species are in association with some crops like cereals (Martyniuk and Martyniuk, 2003). <sup>[34]</sup> They are generally present in natural and alkaline soil occurring more in arable soils. The genus *Azotobacter* comprises of different species: *A. chroococcum*, *A. vinelandii*, *A. beijerinckii*, *A. paspali*, *A. armeniacus*, *A. nigricans* and *A. salinestri* (Gothandapani *et al.* 2017) <sup>[17]</sup>. *Azotobacter* spp. are motile and mesophilic with capability of fixing on an average 20 kg N/ha/per year (Rawia *et al.* 2009). *Azotobacter* spp. as biofertilizer has other roles to play like synthesis of antibiotics, plant growth hormones (Pandey *et al.* 1989), <sup>[40]</sup> vitamins, exopolysaccharides and pigment (Jimenez *et al.* 2011) <sup>[25]</sup> and antifungal activity (Sudhir *et al.* 1983). Table 3. Shown that the per cent increase in yield due to application of *Azotobacter* over chemical fertilizers in different crops.

**Table 3:** Effect of *azotobacter* on crop yield (Dudeja *et al.* 1981) <sup>[14]</sup>

| Crop    | Increase in yield over yields obtained with chemical fertilizers (%) | Crop        | Increase in yield over yields obtained with bio fertilizers (%) |
|---------|--|-------------|---|
| Wheat   | 8-10   | Potato      | 16  |
| Rice    | 5  | Carrot      | 40  |
| Maize   | 15-20  | Cauliflower | 2-24  |
| Sorghum | 15-20  | Tomato      | 7-27  |
| Other   | 13   | Cotton      | 9-24  |

##### 1.1.1.2 Azospirillum

*Azospirillum* are gram-negative, non-nodule-forming, aerobic, nitrogen-fixing bacteria belonging to the family Spirillaceae. They are associated with roots of mainly C<sub>4</sub> crops having

dicarboxylic pathway (Hatch-Slack pathway) of photosynthesis because they grow and fix nitrogen on the organic salts of malic and aspartic acid (Mishra and Dash, 2014). <sup>[38]</sup> *Azospirillum* greatly affects root development and

exudation (Trabelsi and Mhamdi, 2013).<sup>[59]</sup> It is capable of fixing 20–40 kg nitrogen under micro-aerobic conditions. Important species of *Azospirillum* are *A. lipoferum*, *A. brasilense*, *A. amazonense*, *A. haloproferans* and *A. trakense*. Steenhoudt and Vanderleyden (2000)<sup>[60]</sup> reported that when *A. brasilense* sp. produced different phytohormones resulting in increase in maize growth and yield. Currently, *Azospirillum* is used as seed treatment just before sowing without the application in slurry with other products (i.e. micronutrients, fungicides, insecticides, polymers etc.). Apart from BNF, it has also been proved of phytohormone and/or siderophore production, phosphate solubilization (Puente *et al.* 2004),<sup>0</sup> biocontrol activity (Bashan and Bashan, 2010)<sup>[6]</sup> and protection of plants against stress like soil salinity or toxic compounds (Creus *et al.* 1997)<sup>[12]</sup>.

### 1.1.1.3 *Rhizobium*

*Rhizobium* is an example of symbiotic association colonizing legume roots and fixes the atmospheric nitrogen. Bacteria obtain products of photosynthesis as energy source from plant and fix nitrogen from the air for their host. The morphology and physiology of *Rhizobium* at free-living condition is different from bacteroid of nodules. They are the most efficient biofertilizer for legumes as far as the quantity of nitrogen fixation is concerned (Jehangir *et al.* 2017)<sup>[23]</sup> Cross-inoculation plays an important role in host specificity of rhizobium and host (Table 4). The *nod*, *nif* and *fix* genes control the nodulation and nitrogen fixation by the bacterium.

**Table 4:** Major inoculation groups with inoculant and host plants (Ponmurugan and Gopi, 2006)<sup>[42]</sup>

| Cross inoculation Group | <i>Rhizobium</i> species        | Host Legume                             |
|-------------------------|---------------------------------|---|
| Pea group               | <i>R. leguminosarum</i>         | Pea, sweet pea                          |
| Alfalfa group           | <i>R. meliloti</i>              | Sweet clover                            |
| Clover group            | <i>R. trifoli</i>               | Clover / berseem                        |
| Bean group              | <i>R. phaseoli</i>              | All beans                               |
| Soybean group           | <i>Bradyrhizobium japonicum</i> | Lupins                                  |
| Cowpea group            | <i>Rhizobium</i> sp.            | Cowpea, arhar, urd, moong and groundnut |

### 1.1.1.4 Cyanobacteria

Cyanobacteria (Blue Green Alga, BGA) are prokaryotes responsible for water blooms in stagnant waterbodies. They are also found in snow and hot springs. They are photosynthetic and can fix atmospheric nitrogen. They also promote plant growth by producing auxins (indole acetic acid), gibberellic acid and can fix around 20–30 kg N/ha in submerged rice fields. Under low land rice conditions, application of BGA + *Azospirillum* proved beneficial in improving LAI and all yield attributes (Mishra *et al.* 2013)<sup>[38]</sup> Examples of certain BGA are *Nostoc*, *Anabaena*, *Cylindrospermum*, *Gloetrichia Tolypothrix*, *Aulosira* and *Aphanotheca*. Application of BGA can enhance rice productivity from 15% to 38%. Beneficial effects of cyanobacterial inoculation were also reported on a number of other crops such as barley, oats, tomato, radish, cotton, sugarcane, maize, chilli and lettuce (Thajuddin and Subramanian, 2005)<sup>[58]</sup>

### 1.1.1.5 *Azolla*

*Azolla* is a biofertilizer used in rice cultivation extensively. It is an aquatic fern sheltering a cyanobacteria (*Anabaena azollae*). It is an example for symbiotic association. *Azolla* as

biofertilizer may be used as a partial substitute for synthetic fertilizer N due to its sustainable supplementation of N to rice crops and the associated improvement of soil fertility. Nitrogen fixing potential of *Azolla* varies between 30 and 50 kg N ha<sup>-1</sup> which designates *Azolla* as an important biological N source for agriculture and animal industry (Yao *et al.*, 2018).<sup>[63]</sup> Gupta (2004) stated that application of *Azolla* brought an impressive increase in rice yield by 0.5–2 t/ha. Sundaravarathan and Kannaiyan (2002)<sup>[57]</sup> showed that application of *Azolla microphylla* @ 15 t/ha increased grain yield by 29.2%.

### 1.1.1.6 *Gluconacetobacter diazotrophicus*

*Gluconacetobacter diazotrophicus* is a nitrogen fixing bacterium originally found in monocotyledon sugarcane plants in which the bacterium actively fixes atmospheric nitrogen and provides significant amounts of nitrogen to plants. This bacterium mainly colonizes intercellular spaces within the roots and stems of plants and does not require the formation of the complex root organ like nodule. The bacterium is less plant/crop specific and indeed *G. diazotrophicus* has been found in a number of unrelated plant species. Importantly, as the bacterium was of monocot plant origin, there exists a possibility that the nitrogen fixation feature of the bacterium may be used in many other monocot crops.

### 1.1.2 P-solubilization

Most of the phosphorus is insoluble and hence remain unavailable to promote plant growth as plants absorb it only in two soluble forms—monobasic and dibasic. Unavailable P is present in the form of organic matter (phosphodiester, inositol phosphate, apatite). Immobilization of soluble inorganic P fertilizers makes it unavailable for the plants. Phosphorus solubilizing bacteria (PSB) in soil like *Pseudomonas putida* and *Bacillus megaterium* and fungi like *Aspergillus* and *Penicillium* solubilize bound phosphate and fixed phosphorus to make it available for plants. 20–25% phosphorus requirement of plants is fulfilled by PSB and PSF. Thus, its use in agriculture not only reduces manufacturing costs of phosphate fertilizers but also mobilizes insoluble fertilizers in soil (Chang and Yang 2009).<sup>[9]</sup> Among the P-solubilizing microbial population in soil, bacteria (PSB) constitute 1 to 50%, while fungi (PSF) are only 0.1 to 0.5% in P solubilization potential (Chen *et al.* 2006).<sup>[10]</sup> Some actinomycetes are known for P-solubilization and are gaining popularity due to added advantages like capability of surviving in extreme environments like drought, production of antibiotics and phytohormone-like compounds. Hamdali *et al.* (2008) had stated that approximately 20% of actinomycetes can solubilize P, including those in the common genera *Streptomyces* and *Micromonospora*. Some of the P-solubilizing microorganisms are discussed here.

### 1.1.2.1 Bacteria

Also called as “Phosphobacterium”, *Bacillus megaterium* are Gram +ve, rod shaped bacteria reported to enhance mineral phosphorus (P) solubilisation (Lach *et al.* 1990).<sup>[30]</sup> The proposed mechanisms of P-solubilisation are by the release of organic and inorganic acids, excretion of protons that accompanies to the NH<sub>4</sub><sup>+</sup> assimilation and due to the release of phosphatase enzymes that mineralize organic P compounds (Stevenson, 1986). It also has the Zn, K, Fe and Mn solubilizing potential (Amalraj *et al.* 2012). It also enhances root proliferation due to release of growth-promoting

hormones. It supplies P to the plants to the extent of 10 to 15 kg / ha, saving up to 50% over the cost of P chemical fertilizer inputs. Modes of application of this biofertilizer can be seed treatment, root dipping for transplanting, soil application and drip system. *Pseudomonas* and *P. chlororaphis* were able to solubilize P sources strongly and HPLC analyses showed several organic acids (Diriba Muleta *et al.* 2013)<sup>[13]</sup>

### 1.1.2.2 Fungi

Fungi like *Aspergillus* and *Penicillium* are potent P-solubilizing fungi (Vassilev *et al.* 2007; Oleivera *et al.* 2009). Gunes *et al.* (2009)<sup>[19]</sup> reported that commonly found arable soil fungi such as *Penicillium* sp., *Mucor* sp. and *Aspergillus*

sp. have shown to increase plant growth by 5–20%. Kucey and Paul (1982)<sup>[29]</sup> recognized and reported the ability of several species of *Aspergillus* and *Penicillium* to solubilize insoluble phosphate. *A. niger* increased dry biomass of chickpea plants by 22–33% compared with non-inoculated control (Kapri and Tewari 2010).<sup>[26]</sup> PSF gave significantly higher (by 12.6%) grain yield than the control alone by increasing solubility of the unavailable P forms in soil. The study showed potential of using PSF (*Penicillium bilaii*) as bio-inoculants along with 50% of recommended P fertilizer dose that produced wheat yield similar to 100% P when no PSF was used (Ram *et al.* 2015).<sup>[46]</sup>

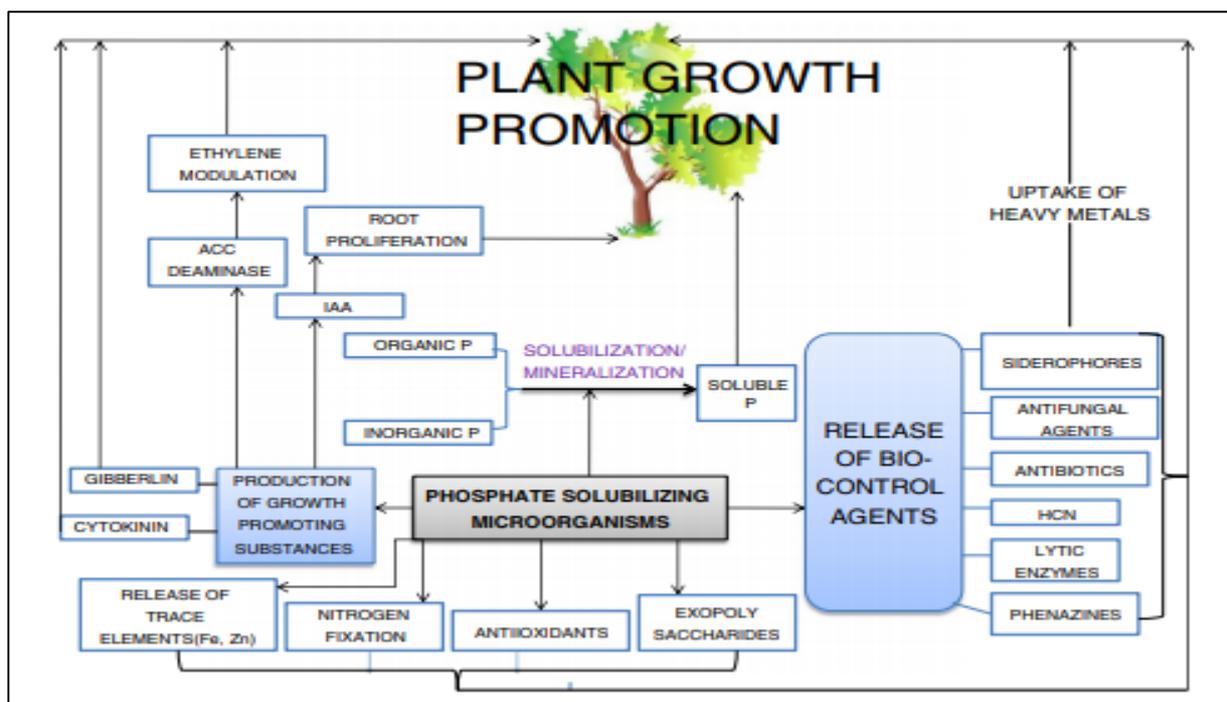


Fig 2: Possible Mechanisms involved in plant growth promotion by PSM (Sharma *et al.* 2013)<sup>[51]</sup>

### 1.1.3 P-mobilizers

Certain microorganisms increase the uptake of P by mobilizing it from a P rich environment to plants instead of solubilizing P. Such microorganisms are called P- mobilizers. While arbuscularmycorrhizae fungi are known to increase fertilizer P use efficiency by increasing its mobility in soil and reducing fixation, these fungi have only an advantage at low P availability in soil (Ghorbanian *et al.* 2012)<sup>[16]</sup> *Pseudomonas letioli* on mobilization of P and growth of Ligol (young apple trees) grown in a pot, significantly increased the total shoot length and solubilised insoluble P compounds (Ewa *et al.* 2013)<sup>[15]</sup>. While Mycorrhizae are the major contributors for P-mobilization, they are being discussed.

#### 1.1.3.1 Mycorrhiza

Mycorrhiza is group of fungi in symbiotic association with plant roots in rhizosphere. They belong to phylum *Glomeromycota* and greatly enhance the uptake of relatively immobile soil nutrients, especially P, by plant root systems (Bolan, 1991)<sup>[7]</sup> They are known for exploitation of a larger soil volume and thus increase P uptake by the extensive external hyphae, Jakobsen *et al.* (1992)<sup>[22]</sup> showed that different amounts of external mycelium produced by AM Fungi supplied different amounts of P to the host plant. In a study conducted by Yao and others (2001),<sup>[63]</sup> the results proved that AMF gave increased contact with phosphates in

the soil by mobilizing phosphates under conditions of low P availability and also solubilized P by production of organic acids. The small diameter of hyphae (2-20  $\mu$ m) makes P acquisition easier as compared to that of new roots formation. Transfer from fungus to plant occurs principally at the arbuscule interface, but likely also at the site of hyphal coils (Karandashov *et al.* 2004)<sup>[27]</sup>.

### 1.1.4 K Solubilizing Bacteria

Potassium (K) is considered as an essential nutrient for plants. Naturally, soils harbour K in larger amounts than any other nutrients but in unavailable form for plant uptake. Only 1 to 2% of this element is available to plants (Sparks and Huang, 1985)<sup>[55]</sup> Many groups of microorganisms like bacteria, fungi and actinomycetes are reported for solubilization of soil K of which, bacteria solubilize by production of inorganic and organic acids, acidolysis, polysaccharides, complexolysis, chelation, polysaccharides, and exchange reactions (Archana *et al.* 2013; Meena *et al.* 2015).<sup>[4]</sup> In rice fields, *Bacillus licheniformis* and *Pseudomonas azotoformans* had higher K-solubilizing ability than other isolated rhizobacteria (Saha *et al.* 2016).<sup>[48]</sup> Prajapati and Modi (2016)<sup>[43]</sup> reported that K and chlorophyll content in cucumber was remarkably enhanced by *Enterobacter hormoechei*, which had the ability of K- solubilization. (Subhashini, 2015)<sup>[55]</sup> observed that soluble potassium and inoculated with strain *Frateuria*

*aurantia* shown higher potassium content of the leaf was increased by 39%. K solubilization is carried out by a large number of bacteria such as *Bacillus mucilaginosus*, *B. edaphicus*, *B. circulans*, *Pseudomonas*, *Burkholderia*, *Acidithiobacillus ferrooxidans*, and *Paenibacillus* spp. The production and management of biological fertilizers containing KSB can be an effective alternative to chemical fertilizers.

### 1.1.5 Plant Growth Promoting Rhizobacteria (PGPR)

The group of bacteria that colonize roots or rhizosphere soil and beneficial to crops are referred to as plant growth promoting rhizobacteria (PGPR). They are also called as microbial pesticides e.g. *Bacillus* spp. and *Pseudomonas fluorescens*. PGPR inoculants currently commercialized that seem to promote growth through at least one mechanism; suppression of plant disease (termed Bio-protectants), improved nutrient acquisition (termed Bio-fertilizers), or phyto-hormone production (termed Bio-stimulants). Species of *Pseudomonas* and *Bacillus* can produce as yet not well characterized phyto-hormones or growth regulators that cause crops to have greater amounts of fine roots which have the effect of increasing the absorptive surface of plant roots for uptake of water and nutrients. These PGPR are referred to as Bio-stimulants and the phyto-hormones they produce include indole-acetic acid, cytokinins, gibberellins and inhibitors of ethylene production Sudheer *et al.*, 1983. [56]. Rhizobacteria induce resistance through the salicylic acid-dependent SAR pathway, or require jasmonic acid and ethylene perception from the plant for ISR. Rhizobacteria belonging to the genera *Pseudomonas* and *Bacillus* are well known for their antagonistic effects and their ability to trigger ISR. Resistance-inducing and antagonistic rhizobacteria might be useful in formulating new inoculants with combinations of different mechanisms of action, leading to a more efficient use for biocontrol strategies to improve cropping systems. Olivera *et al.*, 2009. [39]

### 1.1.6 Zinc solubilizers

The zinc can be solubilized by microorganism's viz., *B. subtilis*, *Thiobacillus thiooxidans* and *Saccharomyces* sp. These microorganisms can be used as bio-fertilizers for solubilization of fixed micronutrients like zinc. The results have shown that a *Bacillus* sp. (Zn solubilizing bacteria) can be used as bio-fertilizer for zinc or in soils where native zinc is higher or in conjunction with insoluble cheaper zinc compounds like zinc oxide (ZnO), zinc carbonate (ZnCO<sub>3</sub>) and zinc sulphide (ZnS) instead of costly zinc sulphate. (Mahdi *et al.* 2010) [33].

## 1.2 Rejuvenation Strategies of Biofertilizers for Sustainable Agriculture Economic Development

The following strategies can be used for rejuvenating sustainable agriculture economic development: •Identifying and selecting efficient locations, crop, soil, and specific strains for N-fixing, P, Zn solubilizing and absorbing (mycorrhizal) to suit different agro climatic conditions. Strain improvement through biotechnological methods. Exchanging cultures between countries of similar climatic conditions and evaluating their performance for a better strain for particular crop as well as checking the activity of cultures during storage to avoid natural mutants • Developing suitable alternative formulations viz., liquid inoculants / granular formulations for all bioinoculants, to carrier based inoculants, standardizing the media, method of inoculation etc., for the new

formulations. •Employing microbiologists in production units to monitor the production • Developing cold storage facilities in production centers •Technical training on the production and quality control to the producers and rendering technical advice and projects to manufacturers • Organizational training to the extension workers and farmers to popularize the technology •Disseminating information through mass media, publications and bulletins. (Gupta and Sen, 2013)

## 1.3 Conclusion and future prospects

As a boon for farmers, Bio-fertilizers being essential components of sustainable farming play vital role in maintaining long term soil fertility and sustainability. Bio-fertilizers would be the viable option for farmers to increase productivity per unit area in organic farming for an era of prosperity and clean environment. Currently there is a gap of ten million tons of plant nutrients between removal of crops and supply through chemical fertilizers. In context of both the cost and environmental impact of chemical fertilizers, excessive reliance on the chemical fertilizers is not viable strategy in long run because of the cost, both in domestic resources and foreign exchange, involved in setting up of fertilizer plants and sustaining the production. Biofertilizers are products that are likely to be commercially promising in the long run once adequate information becomes available to producers and farmers. The use of biofertilizers in India will not only have an impact on sustainable agriculture's economic development but it will also contribute to a sustainable ecosystem and the holistic well-being of the country.

## 2. References

1. Adesemoye AO, Kloepper JW. Plant-microbe's interactions in enhanced fertilizer use efficiency. *Applied Microbiology Biotechnology*. 2009; 1:1-12.
2. Amalraj EDL, maiyappan S, John peter A. *In vivo* and *in vitro* studies of *Bacillus megaterium* var. phosphaticum on nutrient mobilization, antagonism and plant growth promoting traits. *Journal of Eco biotechnology*. 2012; 1:35-42.
3. Anelise Beneduzi, Adriana Ambrosini, Luciane MP, Passaglia. Plant growth-promoting rhizobacteria (PGPR): Their potential as antagonists and biocontrol agents. *Genetics and Molecular Biology*. 2012; 35:41044-1051
4. Archana D, Nandish M, Savalagi V, Alagawadi A. Characterization of potassium solubilizing bacteria (KSB) from rhizosphere soil. *Bioinfollet- A Quarterly J Life Sci*. 2013; 10:248-257.
5. Barman M, Paul S, Choudhury AG, Roy P, Sen J. Biofertilizer as Prospective Input for Sustainable Agriculture in India. *Int. J Curr. Microbiol. App. Sci*. 2017; 11:1177-1186.
6. Bashan Y, De-bashan L. How the plant growth-promoting bacterium *Azospirillum* promotes plant growth. A critical assessment. *Advances in Agronomy*. 2010; 108:77-136.
7. Bolan NS. A critical review on the role of mycorrhizal fungi in the uptake of phosphorus by plants. *Plant Soil*. 1991; 134:189-207.
8. Bouizgarne B, Oufdou K, Ouhdouch Y. Actinorhizal and rhizobial-legume symbioses for alleviation of abiotic stresses. In: Arora NK (Ed) *Plant Microbes symbiosis: applied facets*. Springer, New Delhi. 2015, 273-295.
9. Chang CH, Yang SS. Thermo-tolerant phosphate-solubilizing microbes for multi-functional biofertilizer preparation. *Bioresour Technol*. 2009; 100:1648-1658.

10. Chen YP, Rekha PD, Arunshen AB, Lai WA, Young CC. Phosphate solubilizing bacteria from subtropical soil and their tricalcium phosphate solubilizing abilities. *Appl. Soil Ecol.* 2006; 34:33-41.
11. Chun-li W, Shiuan-yuh C, Chiu-chung Y. Present situation and future perspective of bio-fertilizer for environmentally friendly agriculture. *Annual Reports*, 2014, 1-5.
12. Creus C, Sueldo R, Barassi C. Shoot growth and water status in *Azospirillum* inoculated wheat seedlings grown under osmotic and salt stresses. *Plant Physiology and Biochemistry.* 1997; 35:939-944.
13. Diriba M, Fassil A, Elisabet B, Granhall UF. Phosphate solubilising rhizobacteria associated with *Coffea arabica* L. in natural coffee forests of southwestern Ethiopia. *J Saudi Soc. Agri. Sci.* 2013; 12:73-84.
14. Dudeja SS, Khurana AL, Kundu BS. Effect of rhizobium and phosphorus- micro-organisms on yield and nutrient uptake in chickpea. *Current Science.* 1981; 50:503-505.
15. Ewak, Ewa O, Piotr S, Anna S, Jolanta JS. Effect of *Pseudomonas luteola* on mobilization of phosphorus and growth of young apple trees (Ligol)-Pot experiment. *Scientia Horticulturae.* 2013; 164:270-276.
16. Ghorbanian D, Harutyunyan S, Mazaheri D, Rasoli V, Mohebi A. Influence of *Arbuscular mycorrhizal* fungi and different levels of phosphorus on the growth of corn in water stress conditions. *African J Agricu. Res.* 2012, 2575-2580.
17. Gothandapani S, Soundarapandian S, Jasdeep CP. *Azotobacter chroococcum*: Utilization and potential use for agricultural crop production: An overview. *Int. J. Adv. Res. Biol. Sci.* 2017; 4:35-42.
18. Gothwal RK, Nigam VK, Mohan MK, Sasmal D, Ghosh P. Screening of nitrogen fixers from rhizospheric bacterial isolates associated with important desert plants. *Appl. Ecol. Environ. Res.* 2007; 6:101-109.
19. Gunes A, Ataoglu N, Turan M, Esitken A, Ketterings QM. Effects of phosphate-solubilizing microorganisms on strawberry yield and nutrient concentrations. *J. Plant Nutr. Soil Sci.* 2009; 172:385-392.
20. Gupta AK. The complete technology book on biofertilizers and organic farming. *Natio. Insti. Of Indus. Res. Pres. India*, 2004.
21. Hamdali H, Bouizgarne B, Hafidi M, Lebrihi A, Virolle MJ, Ouhdouch Y. Screening for rock phosphate solubilizing Actinomycetes from Moroccan phosphate mines. *Appl. Soil. Ecol.* 2008; 38:12-19.
22. Jakobsen I, Abbott LK, Robson AD. External hyphae of vesicular-arbuscular mycorrhizal fungi associated with *Trifolium subterraneum* L. I. Spread of hyphae and phosphorus inflow into roots. *New Phytol.* 1992; 120:371-380
23. Jehangir IA, Mir MA, Bhat MA, Ahangar MA. Biofertilizers an Approach to Sustainability in Agriculture: A Review, *Int. J Pure App. Bio sci.* 2017; 5:327-334.
24. Jeyabal A, Kupuswamy G. Recycling of organic wastes for the production of vermicompost and its response in rice legume cropping system and soil fertility. *European J Agronomy.* 2001; 15:153-170.
25. Jimenez DJ, Jose SM, Maria MM. Characterization of free nitrogen fixing bacteria of the genus *Azotobacter* in organic vegetable-grown Colombian soils. *Brazilian J Micro.* 2011; 42:846-858.
26. Kapri A, Tewari L. Phosphate solubilization potential and phosphatase activity of rhizospheric *Trichoderma* spp. *Brazilian J Micro.* 2010; 41:787-79.
27. Karandashov V, Nagy R, Wegmuller S, Amrhein N, Bucher M. Evolutionary conservation of a phosphate transporter in the arbuscular mycorrhizal symbiosis. *Proceedings of the National Academy of Sciences of the United States of America.* 2004; 101:6285-6290.
28. Knobeloch L, Salna B, Hogan A, Postle J, Anderson H. Blue babies and nitrate- contaminated well water. *Environment Health perspective.* 2000; 108:675-678.
29. Kucey RMN, Paul FA. Carbon flux, photosynthesis and nitrogen fixation in mycorrhizal and nodulated faba beans (*Vicia faba* L.) *Soil Biol. Bio chem.* 1982; 14:407-412.
30. Lach D, Sharma VK, Vary PS. Isolation and characterization of a unique division mutant of *Bacillus megaterium*. *J Gen. Microbiol.* 1990; 3:545-553.
31. Latt ZK, Yu SS, Kyaw EP, Lynn TM, New MT. Isolation, Evaluation and Characterization of Free Living Nitrogen Fixing Bacteria from Agricultural Soils in Myanmar for Biofertilizer Formulation. *Int. J Plant Biol. Res.* 2018; 6:1092
32. Malusa E, Sas-paszt L, Ciesielska J. Technologies for beneficial micro-organisms inoculation used as biofertilizers. *The Scientific World*, 2012. doi:10.1100/2012/491206
33. Manashi B, Srijita P, Adit GC, Pinaki R, Jahnvi S. Biofertilizer as Prospective Input for Sustainable Agriculture in India. *Int. J Current Micr. Applied Sci.* 2017; 11:1177-1186.
34. Martyniuk S, Martyniuk M. Occurrence of *Azotobacter* Spp. In Some Polish Soils. *J Envi. Stu.* 2003; 12:371-374.
35. Mazid M, Khan TA, Mohammad F. Potential of NO and H<sub>2</sub>O<sub>2</sub> as signaling molecules in tolerance to abiotic stress in plants. *Journal of Industrial Research & Technology.* 2011; 1:56-68.
36. Meena VS, Maurya BR, Verma JP, Aeron A, Kumar A, Kim K, *et al.* Potassium solubilizing rhizobacteria (KSR): Isolation, identification, and K-release dynamics from waste mica. *Ecol. Eng.* 2015; 81:340-347.
37. Mishra DJ, Rajivir S, Mishra UK, Kumar SS. Role of biofertilizers in organic agriculture: a review. *Research journal of recent sciences.* 2013; 2:39-41.
38. Mishra P, Dash D. Rejuvenation of biofertilizer for sustainable agriculture and economic development. *Consilience: The Journal of Sustainable Development.* 2014; 11:41-61.
39. Oliveira CA, Alvesb VMC, Marreib IE, Gomesb EA, Scottia MR, Carneiro NP. Phosphate solubilizing microorganisms isolated from rhizosphere of maize cultivated in an oxisol of the Brazilian Cerrado Biome. *Soil. Biol. Biochem.* 2009; 41:1782-1787.
40. Pandey A, Kumar SJ. Soil beneficial bacterial and their role in plant growth promotion. *Science Indian Research.* 1989; 48:134-14.
41. Peoples MB, Craswell ET. Biological nitrogen fixation: Investments, expectations and actual contributions to agriculture. *Plan. Soil.* 1992; 141:13-3.
42. Ponnurugan P, Gopi C. Distribution pattern and screening of phosphate solubilizing bacteria isolated from different food and forage crops. *Journal of Agronomy.* 2006; 5:600-604.
43. Prajapati K, Modi H. Growth promoting effect of potassium solubilizing *Enterobacter hormaechei* (KSB-8)

- on cucumber (*Cucumis sativus*) under hydroponic conditions. *Int. J Adv. Res. Biol. Sci.* 2016; 3:168-173.
44. Puente M, Li C, Bashan Y. Microbial populations and activities in the rhizosphere of rock-weathering desert plants. II. Growth promotion of cactus seedlings. *Plant Biology.* 2004; 6:643-650.
  45. Raja N. Biopesticides and Biofertilizers: Ecofriendly Sources for Sustainable Agriculture. *J Biofertil. Biopistici.* 2013; 4:112.
  46. Ram H, Malik SS, Dhaliwal SS, Kumar B, Singh Y. Growth and productivity of wheat affected by phosphorus-solubilizing fungi and phosphorus levels. *Plant Soil Environ.* 2015; 61:122-126.
  47. Rawia EA, Nemat MA, Hamouda HA. Evaluate effectiveness of bio and mineral fertilization on the growth parameters and marketable cut flowers of *Matthiola incana* L. *Am. European J Env. Sci.* 2009; 5:509-518.
  48. Saha M, Maurya BR, Meena VS, Bahadur I, Kumar A. Identification and characterization of potassium solubilizing bacteria (KSB) from Indo-Gangetic Plains of India. *Biocatal. Agric. Biotechnol.* 2016; 7:202-209.
  49. Santos VB, Araujo SF, Leite LF. Soil microbial biomass and organic matter fractions during transition from conventional to organic farming systems. *Geoderma.* 2012; 170:227-31.
  50. Saravanan VS, Madhaiyan M, Osborne J, Thangaraju M. Ecological occurrence of *Gluconacetobacter diazotrophicus* and nitrogen-fixing Acetobacteraceae members: their possible role in plant growth promotion. *Microb. Ecol.* 2008; 55:130.
  51. Sharma SB, Sayyed RZ, Trivedi MH. Phosphate solubilizing microbes: sustainable approach for managing phosphorus deficiency in agricultural soils. *Springer Plus.* 2013; 2:587.
  52. Sparks DL, Huang PM. Physical chemistry of soil potassium. *Potassium in agriculture.* 1985, 201-276
  53. Steenhoudt O, Vanderleyden J. *Azospirillum*, a free living nitrogen-fixing bacterium closely associated with grasses: genetic, biochemical and ecological aspects. *FEMS Microbiol. Rev.* 2000; 24:487-506.
  54. Stevenson FJ. Cycles of soil (carbon, nitrogen, phosphorus, sulfur, micronutrients). John Wiley and Sons New York, 1986, 231-284.
  55. Subhashini DV. Growth promotion and increased potassium uptake of tobacco by Potassium-Mobilizing Bacterium *Frateuria aurantia* grown at different potassium levels in vertisol. *Communications in Soil Science and Plant Analysis.* 2015; 46(2):210-220.
  56. Sudhir U, Meshram A, Jager G. Antagonism of *Azotobacter chroococcum* isolates to *Rhizoctonia solani*. *European Journal of Plant Pathology.* 1983; 89:91-197.
  57. Sundaravarathan S, Kannaiyan S. Influence of *Azolla* and *Sesbania rostrata* application on changes in microbial population and enzymes in rice soils. *Biotechnology of Biofertilizers* (ed. S. Kannaiyan), 2002, 251-225.
  58. Thajuddin N, Subramanian G. Cyanobacterial biodiversity and potential applications in biotechnology. *Curr. Sci.* 2005; 89:47-5.
  59. Trabelsi D, Mhamdi R. Microbial inoculants and their impact in microbial soil microbial communities: a review. *Biomed. Res. Int.* 2013, 11.
  60. Vassilev N, Vassileva M, Bravo V, Fernandez M, Nikolaev I. Simultaneous phytase production and rock phosphate solubilization by *Aspergillus niger* grown on dry olive wastes. *Ind. Crop. Prod.* 2007, 332-336.
  61. Vessey JK. Plant growth promoting rhizobacteria as biofertilizers. *Plant and Soil.* 2003; 255:571-586.
  62. Yao Q, Xiaolin L, Gu F, Peter C. Mobilization of sparingly soluble inorganic phosphates by the external mycelium of an arbuscular mycorrhizal fungus. *Plant and Soil.* 2001; 230:279-285.
  63. Yao YB, Zhanga B, Yuhua T, Miao Z, Ke ZB, Bowen Z, *et al.* Azolla biofertilizer for improving low nitrogen use efficiency in an intensive rice cropping system *Field Crops Research.* 2018; 216:158-164.
  64. Youssef, MMA, Eissa MFM. Biofertilizers and their role in management of plant parasitic nematodes: A review. *Biotechnology Pharmaceutical Resources.* 2014; 5:1-6.