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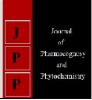
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Chelating agent and chemical fertilizer effect on copper uptake by *Helianthus annuus*

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Abstract

Phytoremediation has shown great potential as an alternative treatment for the remediation of heavy metal contaminated soils. The objective of this research was to investigate the ability of chelating agent and chemical fertilizer for enhancing the phytoremeditation of copper. Ethylenediamine tetraacetic acid (EDTA) and ammonium chloride (NH₄Cl) were applied to the soil at various dosages to elevate metal mobility. It was found that maximum copper extraction was observed by high dosage of chelating agent and chemical fertilizer alone with respect to control. The best treatment for maximum uptake of copper was noted in pots applied with combination of 9 mM EDTA/kg and 300 mg/kg of NH₄Cl with respect to chelating agent and chemical fertilizer alone and control. Chelating agent was much better for phytoextraction of copper than ammonium chloride but ammonium chloride in combination with chelating agent gave the better results.

Keywords: Copper, chelating agent (EDTA), chemical fertilizer (NH₄Cl), phytoextraction, sunflower

Introduction

Land application of biosolids, either as liquid sludge or composted material, often adds heavy metals into the soil. Soil pollution is one of the most vital environmental problem over the last decades (Raskin et al., 1997)^[15]. Heavy metal toxicity and its bioaccumulation in the food chain represent one of the major environmental and health problems of our society. Major sources of metals pollution are burning of fuels, mining and smelting of metallic ferrous ores, municipal sewage, fertilizers and pesticides. The most common pollutants of the environment are Copper, Chromium, Copper, Mercury, Lead, Nickel and Zinc. Cu in excess amounts caused chlorosis, reducing of root and shoot growth (Ait Ali *et al.*, 2002)^[1]. Copper is usually much more bioavailable in soils, and thus particularly toxic to many plant species (Pahlsson, 1989) ^[12]. However, few studies have been performed with biodegradable chelates as the ligand to enhance the phytoextraction of Cu from contaminated soils (Kulli et al., 1999; Kayser et al., 2000) ^[9, 8]. The removal of heavy metals from contaminated soil is a major problem and requires new techniques to enhance the process. Biological processes are being used to decontaminate soils. Some plants are hyper accumulators that actively take up heavy metals and accumulate at in aerial parts with high concentration from soil (Brooks, 1998)^[3], such plants can absorb and accumulate about 10,000 μ g/g of Zn and 1,000 μ g/g of Cu (Turgut et al., 2005)^[12]. Due to the advancement in technology, Plants are used as green technology or phytoremediation for soil amendments and agronomic practices to remove pollutants from the environment or to minimize its toxicity (Raskin and Ensley, 2000; Jing et al., 2007) [14, 7]. Ethylenediamine tetraacetic acid (EDTA) is the most effective chelating agent used for

phytoremediation because it has a strong chelating ability for different metals and it also increases the bioavailability and plant uptake of the metals in the soil (Salt *et al.*, 1998; Huang *et al.*, 1997; Norvell, 1991) ^[16, 6, 11]. About 80% of the total soil metal is solubilized and becomes available for phytoextraction when EDTA is applied (Haag- Kerwer *et al.*, 1999) ^[5]. Synthetic chelating agents such as EDTA allow plants which are not considered as hyper-accumulators to be usable for phytoremediation purposes, because chelates induce plants to take up more heavy metals than they normally accumulate (Liphadzi *et al.*, 2003) ^[10]. Nitrogen fertilizers play a vital role in phytoextraction along with plant growth and enhance heavy metal stress tolerance and absorption (Boroujerdnia and Ansari, 2007) ^[2]. The aim of this research work is to study the effects of chelating agent and Chemical fertilizer either alone or in combination for phytoextraction of copper through *Helianthus annuus* plant.

Materials and methods

Soil preparation and Experimental procedure

The soil was collected from the agricultural field of College of Forestry, SHUATS, Allahabad, thoroughly mixed, dried and then sieved through 4 mm mesh sieve to remove the fiber and other non-soil particulates from the soil.

Plastic pots (cm height and cm diameter) were filled with 5 kg soil that had been previously sieved using 4 mm mesh sieve. The soil was artificially spiked with heavy metal like copper in the form of CuSo₄.5H₂o at the rate of 150 mg/kg five days before sowing. After 5 days of heavy metal application, the seeds of sunflower were sown in each pot. After growing for 3-4 weeks, a dose of 0, 5, 7 and 9 mM / kg of chelating agent (EDTA) and 0, 100, 200 and 300 mg / kg of chemical fertilizer (NH₄Cl) respectively were applied to soil. Data on heavy metal concentration was collected after harvesting of crop. Different agronomic procedures were done during the entire experiment.

Plant analysis

The plants after harvesting were first washed with tap water, distilled water and finally by double distilled water to remove attached soil, separated into roots, stem and leaves, shredded and dried. The vegetation was dried first at room air temperature and then in an oven at 65°C. Dried plant parts were then crushed and powdered separately. Powdered plant samples were then put separately in well washed dried and suitably labeled flasks. A di-acid mixture comprised of concentrated HNO3 and HClO4 was used. To one gram of plant material, 5 ml of concentrated HNO3 was added and kept overnight. Then next day, 12 ml of di-acid mixture (conc. $HNO_3 + HClO_4$ in the ratio of 3:1) was added and digested on hot plate till white reddish brown fumes of perchloric acid comes out. Plant samples slowly begin to dissolve and digest in di-acid mixture. After few hours, plant samples dissolved completely in the digestion mixture and the solution was then evaporated until only about 2 ml was left in the flask. After cooling double distilled water was added and the samples were filtered through Whatman No. 42 filter paper. Digested samples were then transferred for heavy metal analysis using Atomic Absorption spectrophotometer.

Results and Discussion

Phyto-accumulation of Copper in different parts

The effect of different treatments on the accumulation of Cu in the roots and its translocation into the stem and leaf tissues were evaluated. The level of Cu content in the roots stems, and leaf tissues are presented in Table 1. Our results indicated that Chelating agent and Chemical fertilizer alone and in combination played different roles in the accumulation and translocation of Cu into different parts of the plant. The accumulation of Cu in roots, stem and leaves significantly increased with all treatments as compared to control.

The data revealed that maximum copper uptake 2.99, 2.51 and 1.25 ppm in case of chemical fertilizer alone was found in roots, stem and leaves respectively in F₃ (300 mg / kg NH₄Cl) and minimum 2.33, 1.87 and 0.96 ppm respectively were recorded in F_0 (0 mg / kg NH₄Cl). The data also suggested that in case of chelating agent alone, the maximum copper uptake 4.03, 3.43 and 1.92 ppm was recorded in C₃ (9 mM EDTA/kg) and minimum 1.25, 0.91, 0.34 ppm was recorded in C_0 (0 mM/kg of EDTA), but in case of interaction effect of both EDTA and NH₄Cl, it was observed that the maximum uptake of copper 4.31, 3.61 and 2.23 ppm was found in roots, stem and leaves respectively in $T_{\rm 15}$ (9 mM / kg EDTA and $300\ mg$ / kg NH4Cl) and minimum 0.56, 0.30 and 0.19 ppm respectively were recorded in T₀ (no chelant and chemical fertilizer). The data clearly revealed that both increasing concentration of ammonium chloride and EDTA increased copper uptake and chelating agent was much better than ammonium chloride. From the study it is clear that the EDTA

in combination with ammonium chloride boosted the copper uptake more than the EDTA and ammonium chloride did alone. The results indicated that the sunflower can tolerate heavy metal concentration and EDTA and NH₄Cl had potential to promote the uptake of copper. Combined effect of 9 mM EDTA / kg and 300 mg / kg NH₄Cl was found best but low dose of EDTA should be used because of their environmental risks. Our results were also in accordance with (Ebrahimi, 2013)^[4] who found that Cu content in the roots of the plant increased when the dose of EDTA was 10 mmolkg⁻¹. Soil amended with N fertilizer resulted in the highest accumulation of Pb and Cu (Rahman *et al.*, 2013)^[13]. Nitrogen fertilizer boosted growth parameters increasing root and shoot dry matter significantly, allowing a higher uptake of toxic metals by plants (Ebrahimi 2013)^[4].

Table 1: Effect of Chelating agent and Chemical fertilizer on uptake of Copper by root, stem and leaves of Sunflower.

Treatment combination	Root	Stem	Leaves
$T_0(C_0F_0)$	0.56	0.30	0.19
$T_1(C_0F_1)$	1.29	0.92	0.31
$T_2(C_0F_2)$	1.47	1.13	0.39
$T_3(C_0F_3)$	1.69	1.29	0.48
$T_4(C_1F_0)$	2.07	1.52	0.70
$T_5(C_1F_1)$	2.23	1.75	0.73
$T_6(C_1F_2)$	2.36	1.95	0.80
$T_7(C_1F_3)$	2.52	2.14	0.83
$T_8(C_2F_0)$	2.92	2.44	1.22
$T_9(C_2F_1)$	3.11	2.63	1.28
$T_{10}(C_2F_2)$	3.25	2.81	1.34
T ₁₁ (C ₂ F ₃)	3.44	3.00	1.46
$T_{12}(C_3F_0)$	3.76	3.21	1.72
T ₁₃ (C ₃ F ₁)	3.98	3.39	1.81
T ₁₄ (C ₃ F ₂)	4.08	3.49	1.92
$T_{15}(C_3F_3)$	4.31	3.61	2.23
CD (P=0.05)	0.030	0.034	0.036

C= chelating agent, F= chemical fertilizer

 $\begin{array}{l} T_0 = 0 \ \text{mM EDTA/kg} + 0 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_1 = 100 \ \text{mg/kg} \\ \text{NH}_4\text{Cl}, \ T_2 = 200 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_3 = 300 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_4 = 5 \ \text{mM EDTA/kg}, \ T_5 = 5 \ \text{mM EDTA/kg} + 100 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_7 = 5 \ \text{mM} \\ \text{EDTA/kg} + 300 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_8 = 7 \ \text{mM EDTA/kg}, \ T_9 = 7 \\ \text{mM EDTA/kg} + 100 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_{10} = 7 \ \text{mM EDTA/kg} + 200 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_{10} = 7 \ \text{mM EDTA/kg} + 200 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_{11} = 7 \ \text{mM EDTA/kg} + 300 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_{12} = 9 \ \text{mM EDTA/kg} + 100 \\ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_{13} = 9 \ \text{mM EDTA/kg} + 100 \\ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_{14} = 9 \ \text{mM EDTA/kg} + 200 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_{15} = 9 \ \text{mM EDTA/kg} + 300 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \ T_{15} = 9 \ \text{mM EDTA/kg} + 300 \ \text{mg/kg} \ \text{NH}_4\text{Cl}, \end{array}$

Conclusion

The study demonstrated that chelating agent and chemical fertilizer are efficient in increasing the concentrations of Cu in different parts of Sunflower grown in metal contaminated soil. Chelating agent was much more efficient than chemical fertilizer in enhancing concentration of Copper. Application of 9 mM / kg EDTA and 300 mg / kg NH₄Cl appeared to be the best treatment for phytoextraction of Cu from contaminated soil. From the study it is clear that chemical fertilizer should be used with chelating agent in order to have a good biomass and maximum uptake of heavy metal. Sunflower was the best plant to carry out the phytoextraction of Copper.

References

1. Ait Ali N, Bernal MP, Ater M. Tolerance and bioaccumulation of copper in Phragmites australis and

Zea mays. International Journal of Plant & Soil Science. 2002; 39:103-111.

- Boroujerdnia M, Ansari NA. Effect of different levels of nitrogen fertilizer and cultivars on growth, yield and yield components of romaine lettuce (*Lactuca sativa* L.). Middle Eastern and Russian Journal of Plant Science and Biotechnology. 2007; 1:47-53.
- 3. Brooks RR. Photochemistry of hyper accumulators. In: R.R. Brooks (Ed.), Plants those Hyper accumulate Heavy Metals, New York: CAB International, 1998, 15-53.
- Ebrahimi M. Effect of EDTA application on heavy metals uptake and germination of *Echinochloa crus galii* (L.) Beave in contaminated soil. Intl J Agri Crop Sci. 2013; 6(4):197-202.
- Haag-Kerwer A, Schafer HJ, Heiss S, Walter C, Rausch T. Copper exposure in *Brassica juncea* causes a decline in transpiration rate and leaf expansion without effect on photosynthesis. Journal of Experimental Botany. 1999; 50:1827-1835.
- Huang JW, Chen J, Berti WR, Cunningham DS. Phytoremediation of lead-contaminated soils: Role of synthetic chelates in lead phytoextraction. Environmental Science and Technology. 1997; 31:800-805.
- Jing YD, He ZL, Yang XE. Role of soil rhizobacteria in phytoremediation of heavy metal contaminated soils. Journal of Zheijang University SCIENCE B. 2007; 8:192-207.
- Kayser A, Wenger K, Keller A, Attinger W, Felix HR, Gupta SK *et al.* Enhancement of phytoextraction of Zn, Cd, and Cu from calcareous soil: the use of NTA and sulfur amendments. Environ. Sci. Technol. 2000; 34:1778-1783.
- Kulli B, Balmer M, Krebs R, Lothenbach B, Geiger G, Schulin R. The influence of nitrilotriacetate on heavy metal uptake of lettuce and ryegrass. J Environ. Qual. 1999; 28:1699-1705.
- 10. Liphadzi MS, Kirkham MB, Mankin KR, Paulsen GM. EDTA assisted heavy metals uptake by poplar and sunflower grown at a long-term sewage-sludge farm. Plant and Soil. 2003; 257:171-182.
- Norvell WA. Reactions of metal chelates in soils and nutrient solutions, In: Mortvedt, J.J., Cox, F.R., Shuman, L.M., Welch, R.M. (Eds.), Micronutrients in Agriculture, 2nd edn. Book Series. Soil Science Society of America, Madison, Wisconsin. 1991; 4:187-227.
- Pahlsson AMB. Toxicity of heavy-metals (Zn, Cu, Cd, Pb) to vascular plants-a literature review. Water Air Soil Pollut. 1989; 47:287-319.
- 13. Rahman MM, Azirun MS, Boyce NA. Enhanced Accumulation of Copper and Lead in Amaranth (*Amaranthus paniculatus*), Indian Mustard (*Brassica juncea*) and Sunflower (*Helianthus annuus*). Plos One. 2013; 8(5):1-9.
- 14. Raskin I, Ensley BD. Phytoremediation of toxic metals: using plants to clean up the environment. New York: John Wiley, Cosio, C, 2000, 247-271.
- 15. Raskin I, Smith RD, Salt DE. Phytoremediation of metals; Using Plants to remove pollutants from the environment. Journal of Current Opinion in Biotechnology. 1997; 8:221-126.
- Salt DE, Smith RD, Raskin I. Phytoremediation. Annual Review of Plant Physiology and Plant Molecular Biology. 1998; 49:643-668.
- 17. Turgut C, Pepe MK, Cutright T. The Effect of EDTA on *Helianthis annuus* Uptake, Selectivity, and Translocation

of Heavy Metals When Grown in Ohio, New Mexico and Columbia Soils. Chemosphere. 2005; 58:1087-1095.