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Dark goldenrod pigment production by *p. purpureogenum* using *Trigonella foenum-graecum* leaves and characterization and identification of phytochemicals by analytical techniques

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Abstract

In this study, the bioactive components of *Trigonella foenum-graecum* leaves have been evaluated using techniques like UV-Vis, FTIR, Proton-NMR spectroscopy and Gas Chromatography-Mass Spectrometry. The chemical compositions of the microbial extract of *Trigonella foenum-graecum* leaves were investigated from Gas Chromatography-Mass Spectrometry, the spectra of the compounds found in the extract were matched with the National Institute of Standards and Technology (NIST) library. GC/MS analysis of the microbial extract of *Trigonella foenum-graecum* leaves revealed the existence of The results of this study offers a platform for using *Trigonella foenum-graecum* leaves as the herbal alternative for the current synthetic antimicrobial agents.

Keywords: Ultraviolet spectroscopy, antimicrobial agents, NIST library.

1. Introduction

Nowadays, traditional herbal medicines are gaining popularity as the primary health care needs, as an alternative and complementary therapies [1]. These plants can be used as a wide range of biological and pharmacological activities such as anti-cancer, anti-inflammatory, diuretic, oxytocic, laxative, antispasmodic, antihypertensive, anti-diabetic, and anti-microbial functions. The secondary metabolites of plants provide humans with numerous biological active products which have been used extensively as drugs, foods, additives, flavors, insecticides, colorants, fragrances and chemicals [2]. The leaves of the plant have been used as a traditional remedy for numerous conditions including gastrointestinal disorders, gout, wound healing and inflammation, hyperlipidemia and diabetes [3]. The antihyperglycemic effects of fenugreek leaves and its subfractions are demonstrated in diabetic rats [4] dogs [5] and humans. The leaves also show beneficial effects in hypolipidemic subjects [6] and in cancer [7]. Supplementation of leaves in the diet enhances the antioxidant potential in control and diabetic rats [8]. It has been reported to have restorative and nutritive properties and to stimulate digestive processes, useful in healing of ulcers in digestive tract [9]. Bioactive compounds isolated from fenugreek leaves include saponins, amino acids, flavonoids, coumarins, mucilaginous fibers, and other vitamins and minerals [10, 3]. Flavonoids have remarkable biological activities, including inhibitory effects on enzymes, a modulatory effect on some cell types, protection against allergies, antibacterial, antifungal, antiviral, antimalarial, antioxidant, anti-inflammatory and anticarcinogenic properties [11]. Much of the hypoglycemia effect of fenugreek leaves in clinical studies is likely due to the inhibitory effects of mucilaginous fibers on glucose absorption [12, 13] Hence it was planned to study the phytochemicals present in the microbial of *Trigonella foenum-graecum* leaves by the analytical tools to validate its use in textiles.

2. Materials

The *Penicillium purpureogenum* - was purchased from NCI Pune, and the stock culture was maintained on a Potato Dextrose Agar (PDA) slants.

The leaves of *Trigonella foenum-graecum* were collected in fresh polythene bags from Kakinada, East Godavari district, A.P.

3. Methodology

3.1 Production of microbial pigments from *Trigonella foenum-graecum*

The leaves were initially washed in tap water, then with distilled water to remove soil and other contaminants. They were weighed and ground into a paste and as the carbon source.

3.1.1 Fermentation

A loopful of well sporulated culture of the *Penicillium Purpurogenum* was inoculated into a corresponding 250 mL Erlenmeyer flask containing 100 mL of production medium composed of PDB (2%), MgSO_4 (1%), MnSO_4 (1%), K_2HPO_4 (1%) and KH_2PO_4 (1%) and Urea (0.5%) with pH 5.5. The inoculated flask was incubated on a rotary shaker (200 rpm) at 25°C for 7-10 days.

3.1.2 Pigment extraction

After incubation, the broth obtained was taken and heated on a heating mantle at 70 degrees Celsius for 2 hours. After heating, the broth was filtered, separating the biomass and the filtrate. The pH of the filtrate was checked. The solution obtained was evaporated and concentrated at 70 degrees Celsius. The water molecules were slowly removed on evaporation leaving the solid concentrate. The concentrate was cooled immediately. The crude extract obtained was allowed for crystallization to form crystals of the pigment. The pigment obtained was purified and weighed.

3.2 UV-Visible Spectroscopy (UV-Vis)

The maximum absorbance of extracted and dried red pigment powder was determined by spectrophotometer (SPECORD 210-222K333 UV-Vis) at 500 nm wavelength⁵.

3.3. Fourier Transform Infra-Red (FT-IR) spectroscopy

The Fourier transform infrared (FT-IR) spectrum was recorded on a Bruker FT-IR spectrophotometer and the spectral range was 4000 – 500 cm^{-1} ⁶. The dried powder of the

red pigment was scanned by a Shimadzu FT-IR 8000 spectrophotometer in the 4000–400 cm^{-1} range using the KBr method at 27 °C.

3.4. Nuclear Magnetic Resonance Spectroscopy

The purified pigment was dissolved with dimethylsulphoxide ($\text{DMSO}-d_6$) and the sample was injected into a nuclear magnetic resonance (NMR) spectrometer (Bruker 400 MHz)⁷.

3.5. Gas Chromatography-Mass Spectrometry

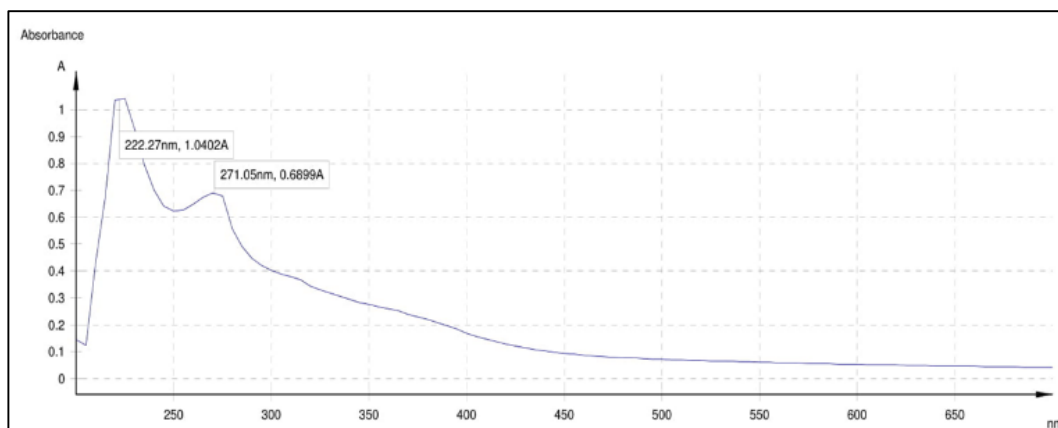
In the current study, the microbial extract of *Trigonella foenum-graecum* was screened for the presence of phytochemical compounds by qualitative test procedures followed by GC-MS analysis of novel compounds. This study was facilitated by using Gas Chromatography-Mass Spectrometry. The mass spectra of compounds in the microbial extract were matched with NIST (National Institute of Standards and Technology) and WILEY library.

4. Results and Discussion

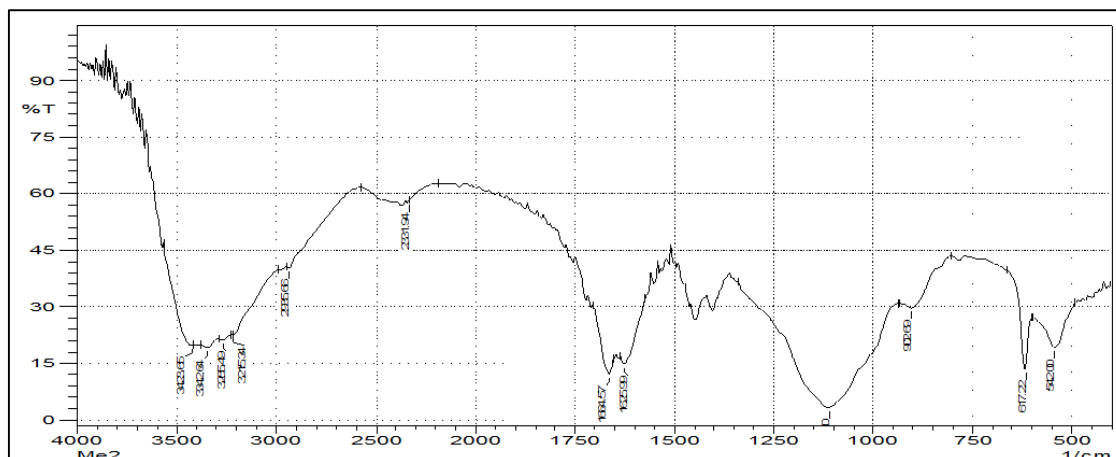
The pigment produced from fermentation of under the optimal conditions were pH 5.5, temperature 37°C and production time 8 days. The highest production of *Trigonella foenum-graecum* L. pigments is 1.9% i.e., 19.8g/L and the maximum absorbance at 222 nm (1.0402). The produced pigment was characterized using different techniques like UV Visible Spectroscopy, FTIR and ^1H NMR and GC-MS techniques.

Figure no 1:- Characterization of the main colored component of the microbial extract of *Trigonella foenum-graecum* leaves.

UV-Visible Spectra

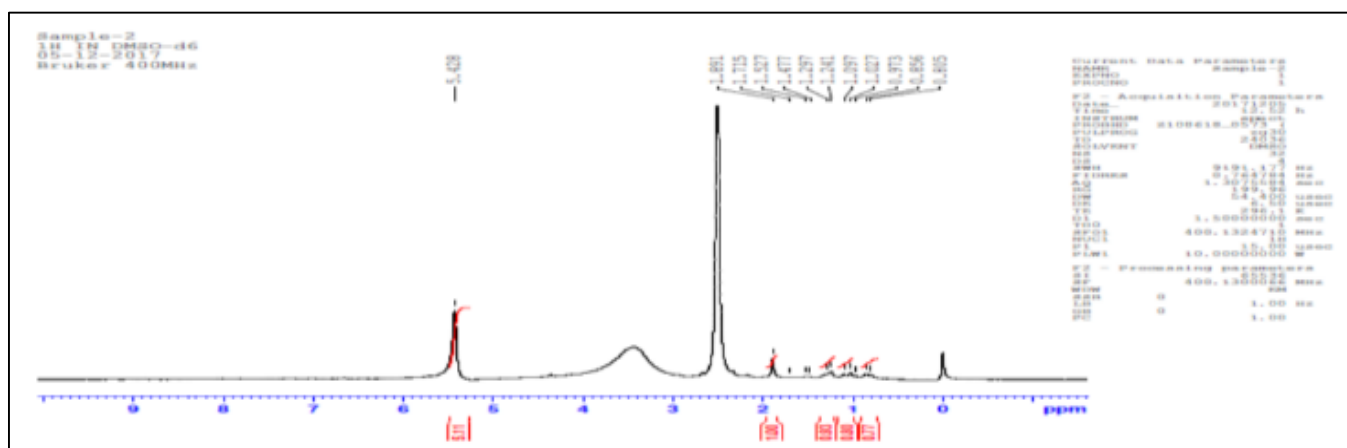


FTIR Spectra

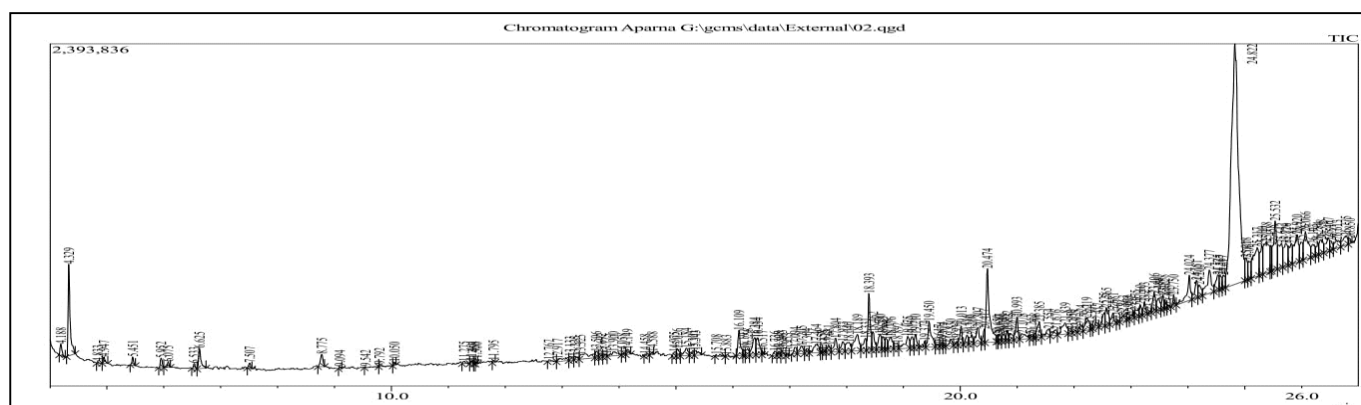


Absorption Peak value	Absorption range	Specific type of bond
3423.65	3500-3300	1° amines (doublet), 2° amines N-H stretch
3342.64	3500-3300	1° amines (doublet), 2° amines N-H stretch
2935.66	3000-2830	Alkanes C—H stretch
1664.57	1670-1640	Amides C=O stretch (Amide II band)
1625.99	1640-1550	Amides, 1° and 2° amines N-H stretch
1125.34	1300-1000	Alcohols, esters, ethers, -COOH, Anhydrides C-O stretch
902.69	1000-650	Alkenes C-H out of plane bend
617.22	800-600	Chloride C-Cl stretch

H-NMR Spectra

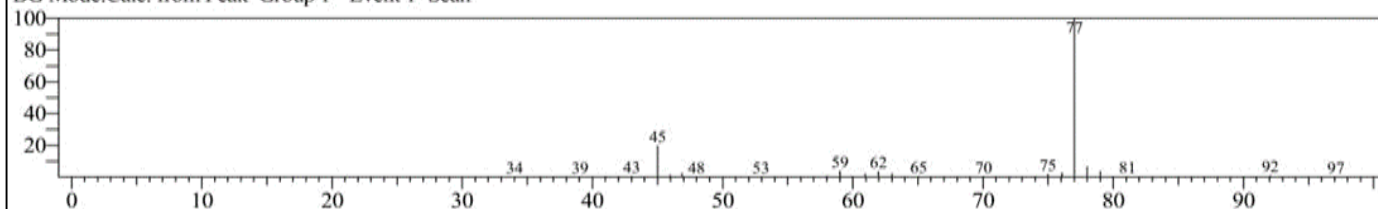


GC-MS Spectrum

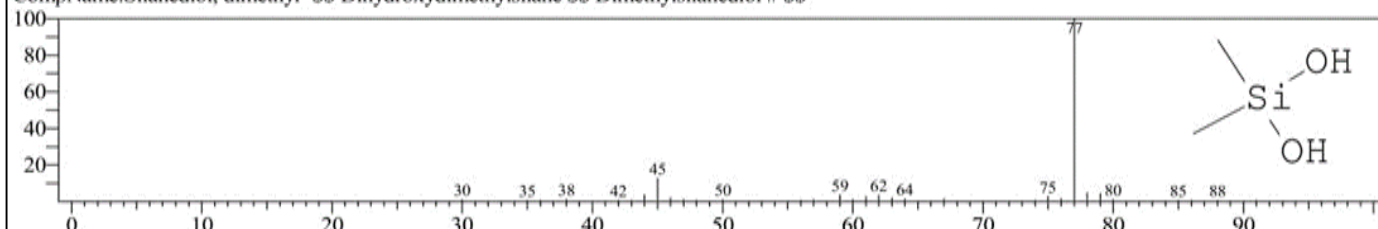


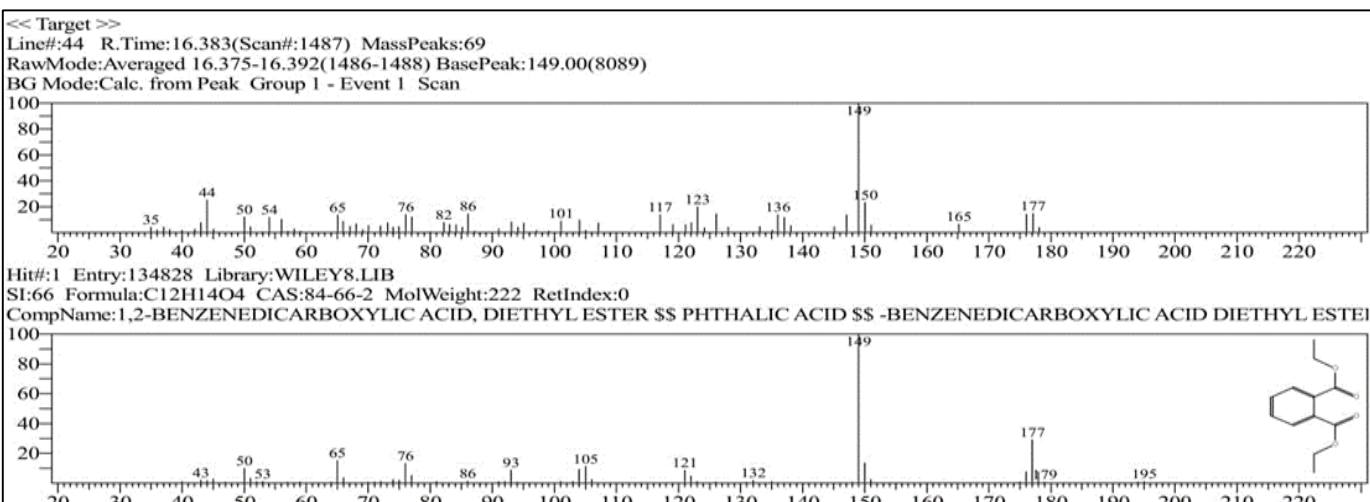
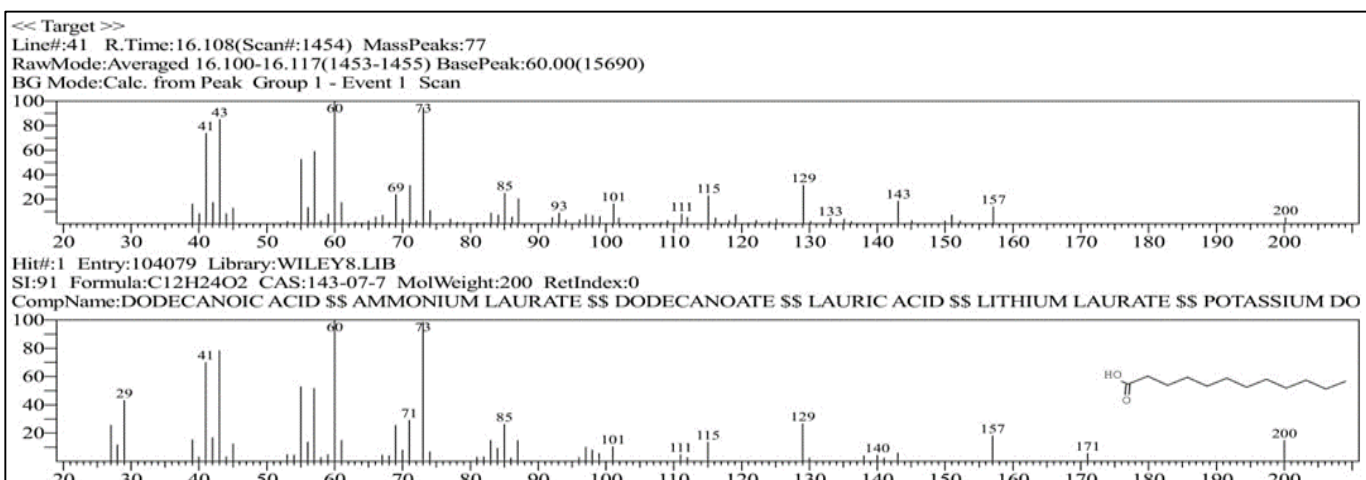
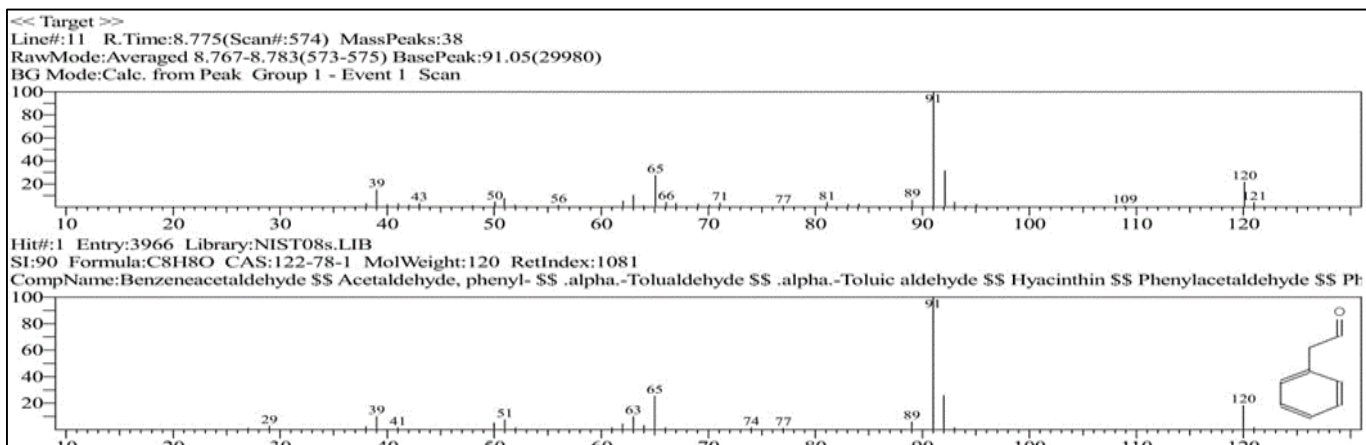
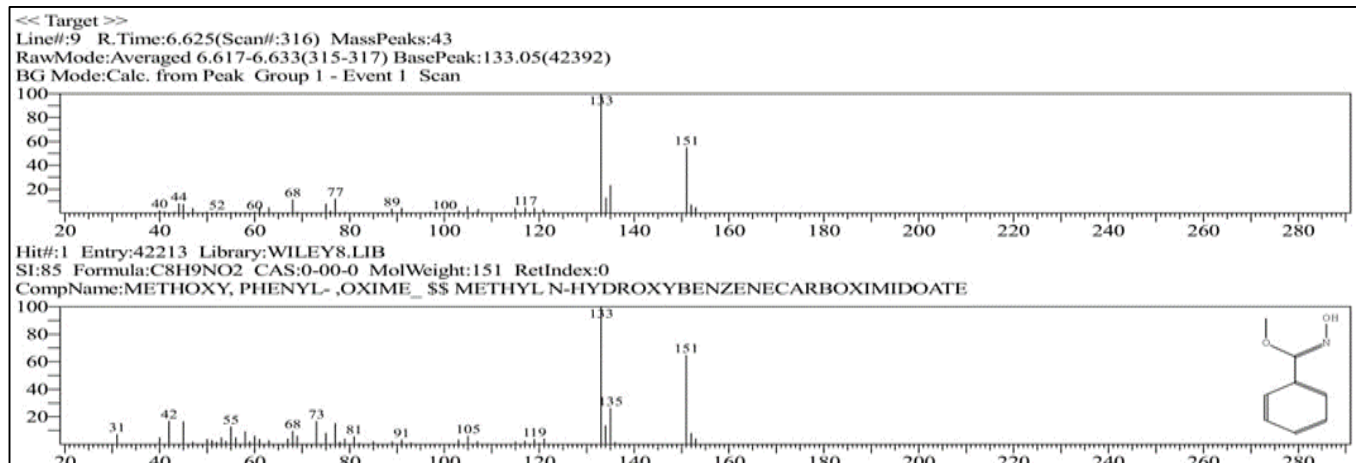
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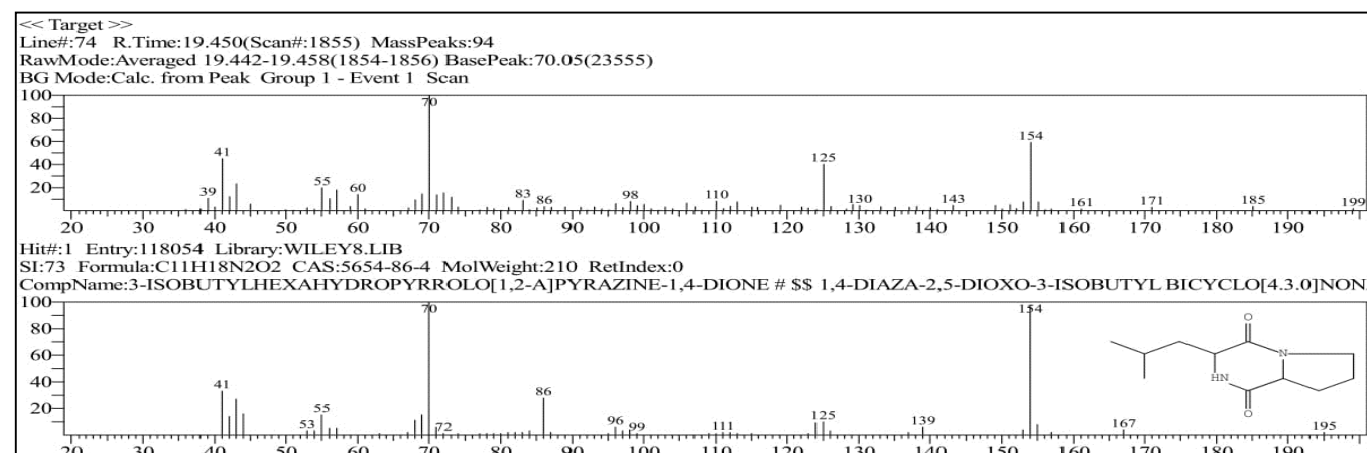
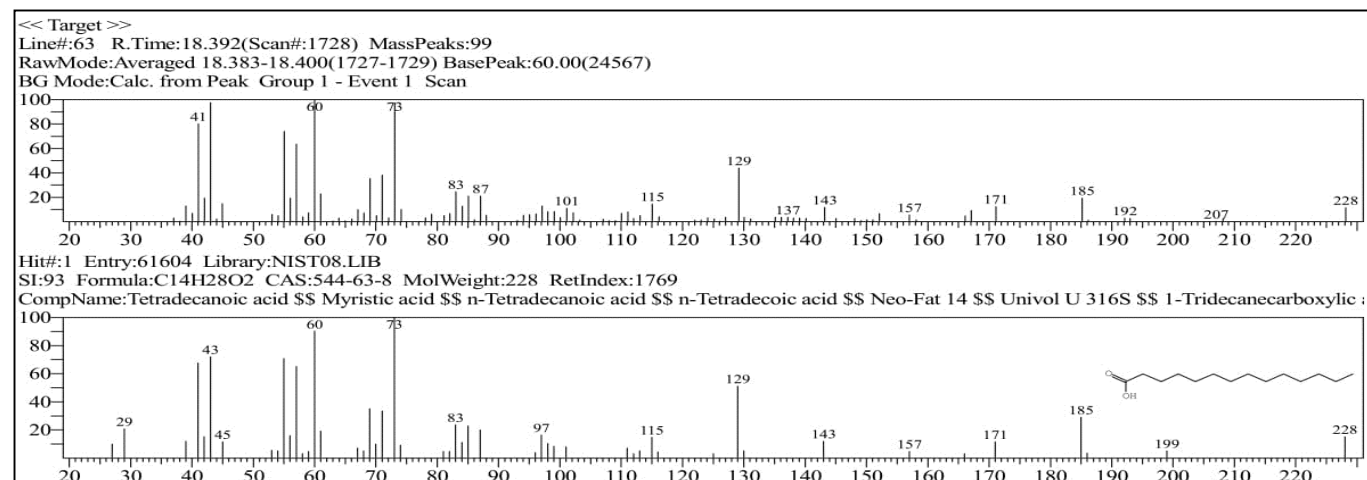
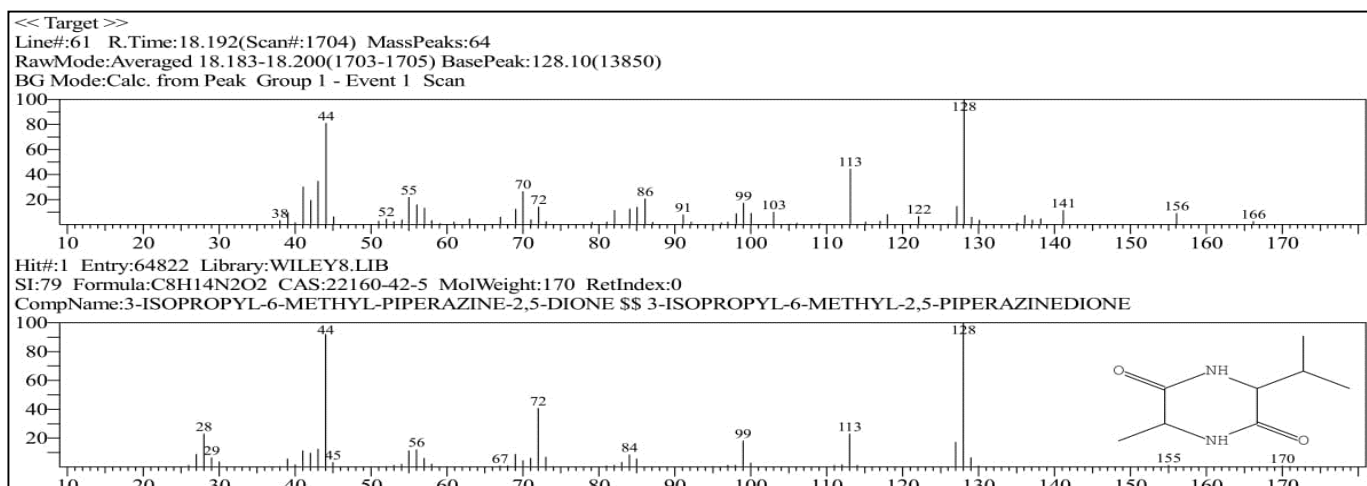
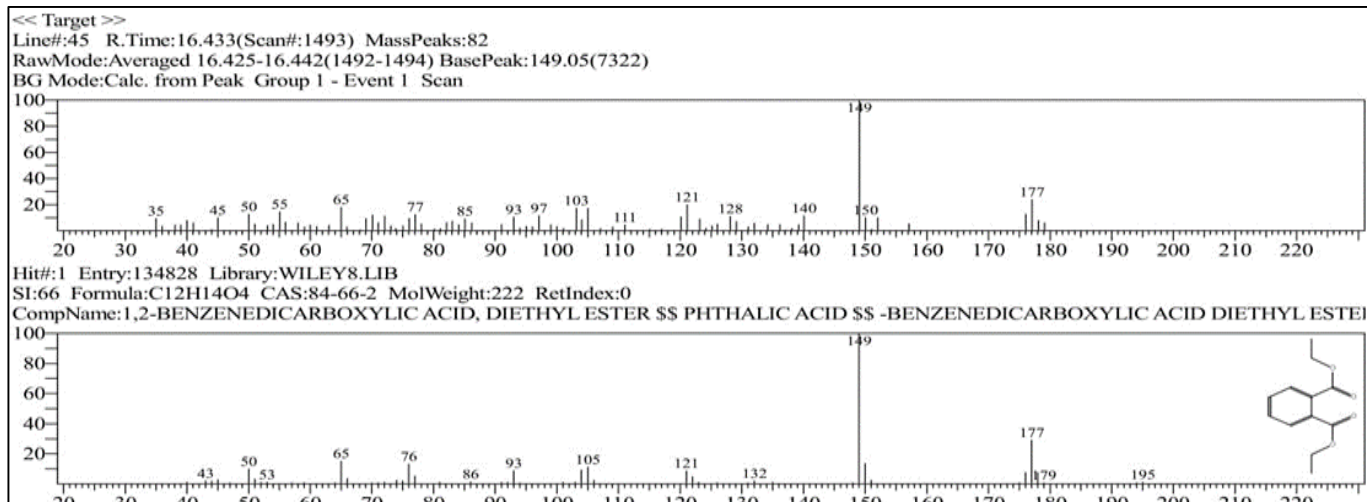
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 RawMode:Averaged 4.317-4.333(39-41) BasePeak:77.00(366923)
 BG Mode:Calc. from Peak Group 1 - Event 1 Scan

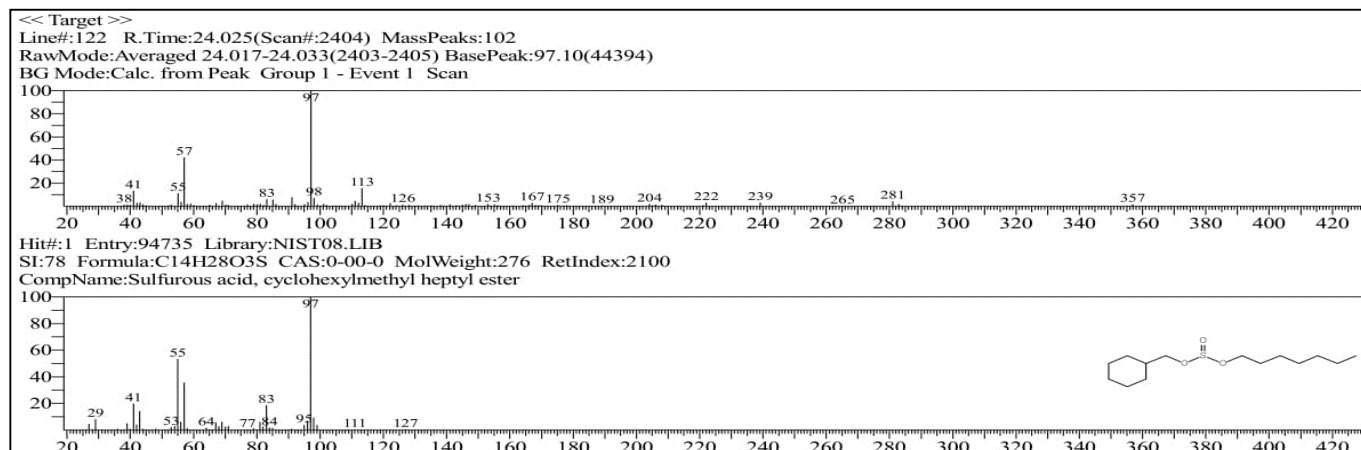
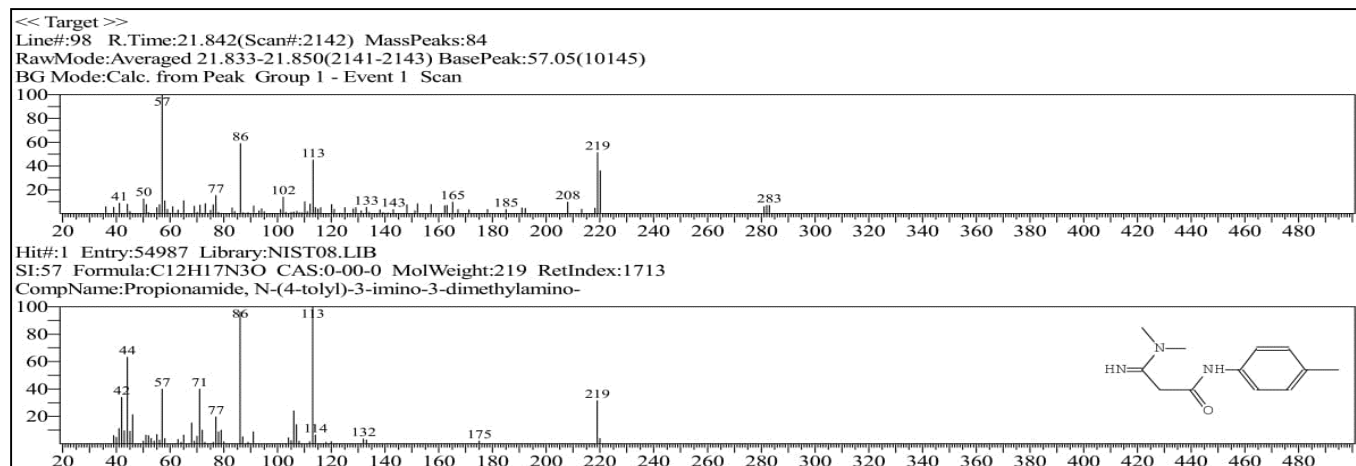
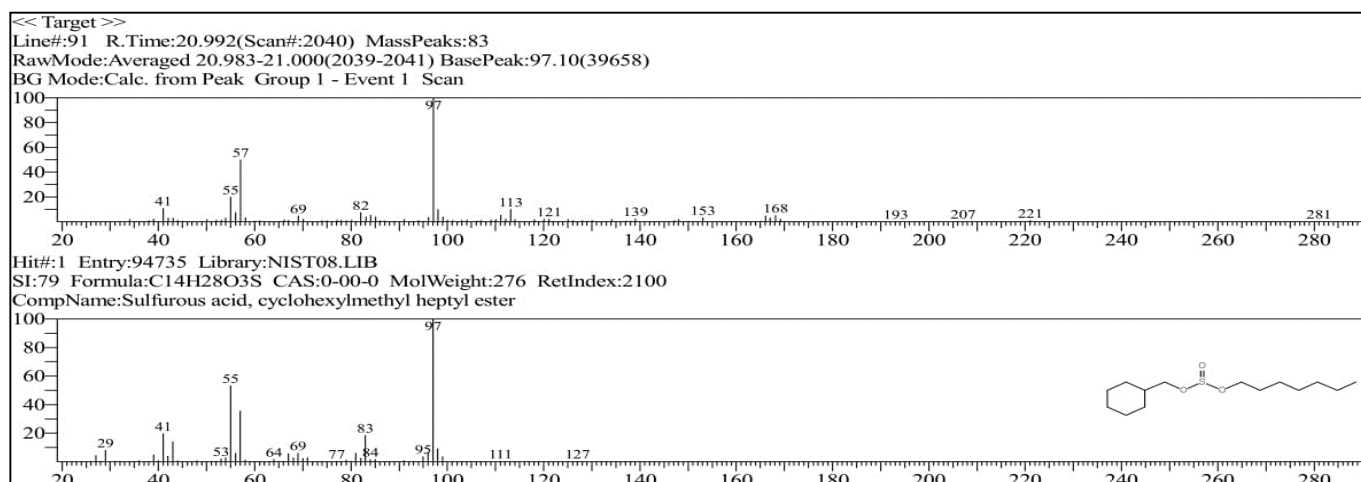
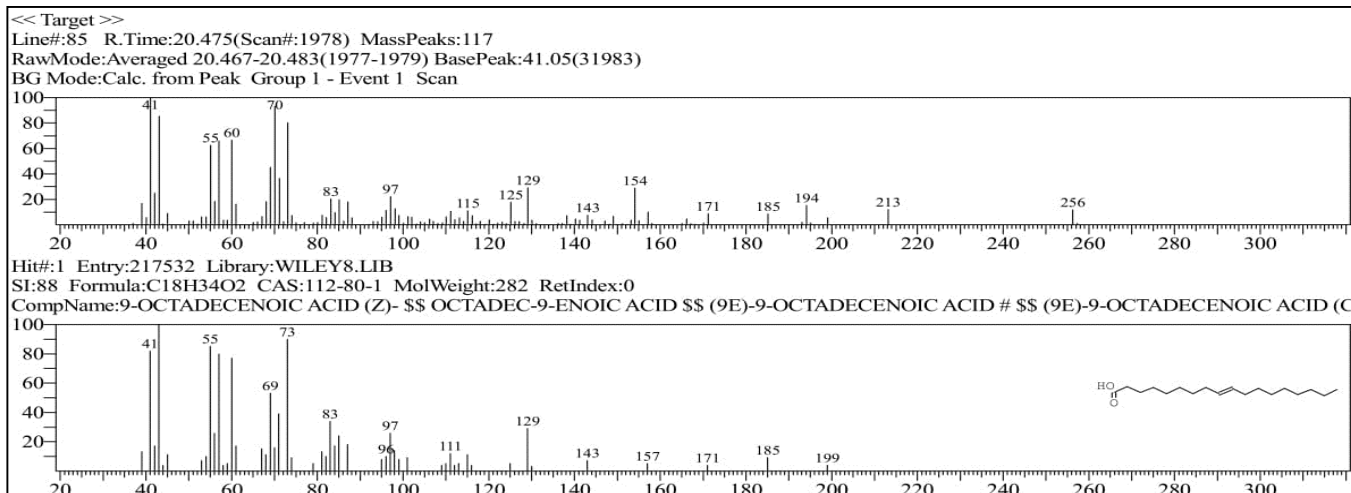


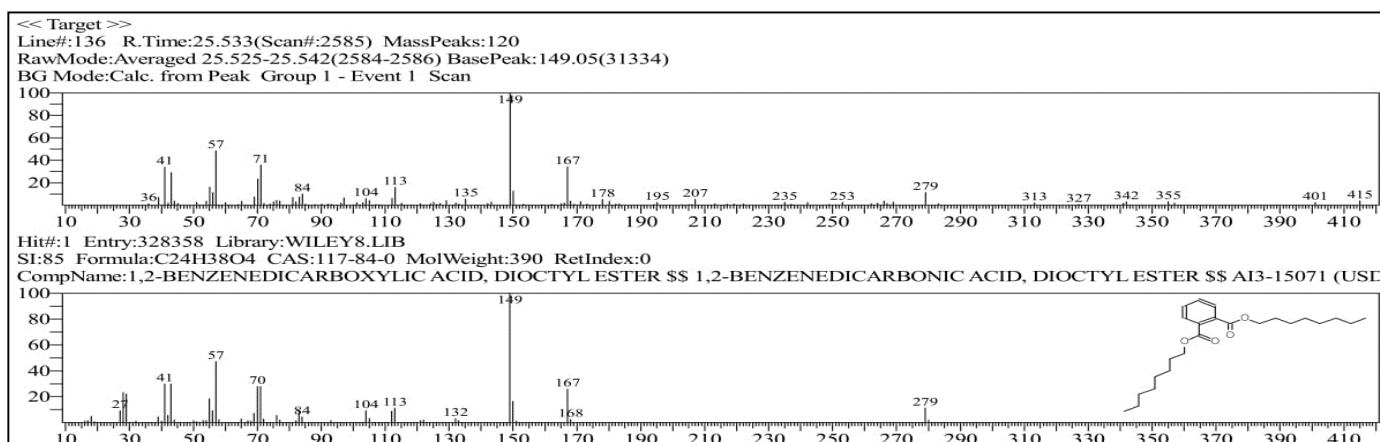
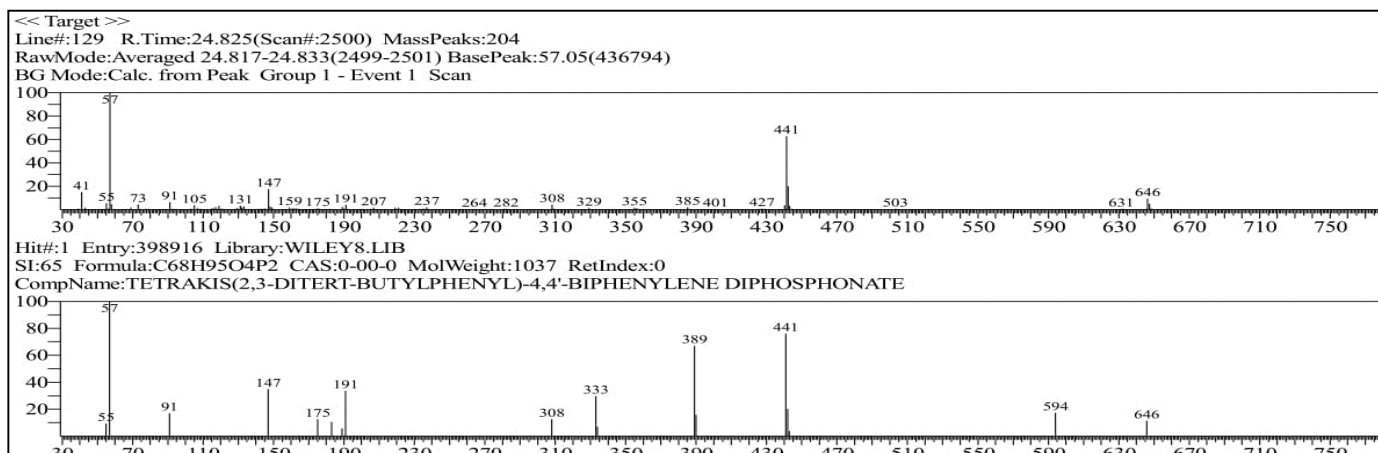
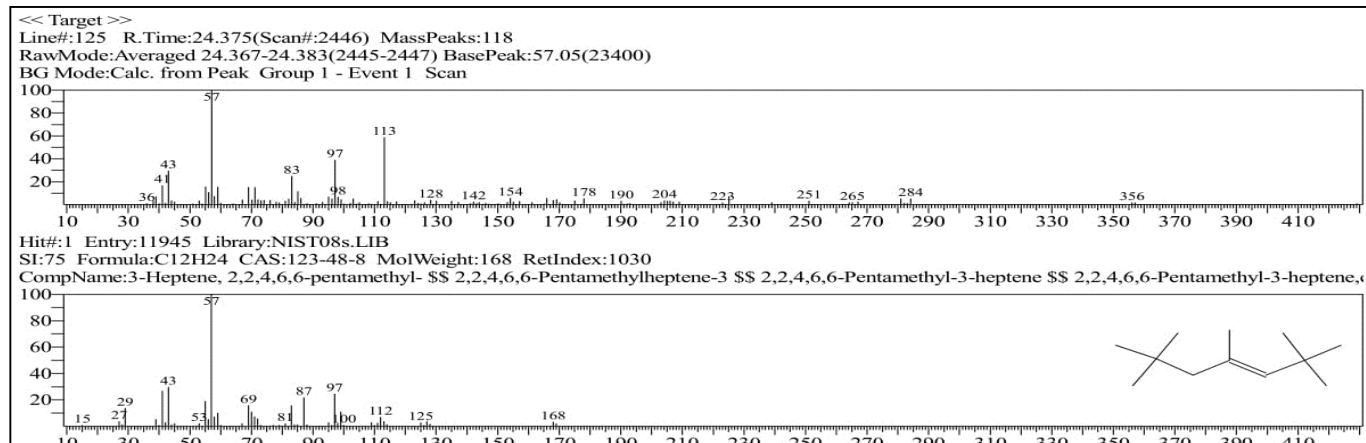
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 CompName:Silanediol, dimethyl- \$\$ Dihydroxydimethylsilane \$\$ Dimethylsilanediol # \$\$











Major phytochemical compounds identified in the microbial extract of *Trigonella foenum-graecum* leaves

S. No	RT(min)	Name of the compound	Molecular Formula	Molecular Weight	Peak Area %	Pharmacological purpose
1	4.329	Silanediol, dimethyl-	C ₂ H ₈ O ₂ Si	92	2.65	No activity reported
2	6.625	Methoxy, Phenyl-Oxime-	C ₈ H ₉ NO ₂	151	0.69	Used as Antidote for
3	8.775	Benzene acetaldehyde	C ₈ H ₈ O	120	0.45	Flavoring agent
4	16.109	Dodecanoic Acid	C ₁₂ H ₂₄ O ₂	200	0.94	Flavoring agent, Herbicide, Insecticide, Surfactants
5	16.384 16.434	1,2-Benzenedicarboxylic acid diethyl ester	C ₁₂ H ₁₄ O ₄	222	0.94 0.78	Flavoring agent, Phthalates, Adhesives, and sealant chemicals, Odor agents, and Plasticizers
6	18.189	3-Isopropyl-6-methyl-piperazine-2,5-dione	C ₈ H ₁₄ N ₂ O ₂	170	1.16	Used as an anticonvulsant
7	18.393	Tetradecanoic acid	C ₁₄ H ₂₈ O ₂	228	1.89	Flavoring agent, Surfactants
8	19.450	3-Isobutyl hexahydro pyrrolo (1,2a) pyrazine	C ₁₁ H ₁₈ N ₂ O ₂	210	1.28	Bitter component of sake and contributes to the flavour of bee
9	20.474	9-Octadecenoic acid (Z)	C ₁₈ H ₃₄ O ₂	282	3.51	Flavoring agent, Food Additive
10	20.993 24.024	Sulfurous acid	C ₁₄ H ₂₈ O ₃ S	276	0.70 1.26	Bleaching Activity
11	21.839	Propionamide	C ₁₂ H ₁₇ N ₃ O	219	0.83	No activity reported
12	24.377	3-Heptene	C ₁₂ H ₂₄	168	1.50	Branched unsaturated hydrocarbons
14	24.822	Tetrakis (2,3-Ditert-butylphenyl)-4,4'-Biphenylene diphosphonate	C ₆₈ H ₉₅ O ₄ P ₂	1037	24.65	stabilizer in polymers where it functions as an antioxidant
15	25.532	1,2-Benzenedicarboxylic acid	C ₁₄ H ₁₈ O ₄	254	2.34	Plasticizers

GC-MS analysis of a microbial extract of *Trigonella* leaves detected 15 phyto compounds having antioxidant and flavoring properties. Especially Tetrakis (2, 3-Ditert-butylphenyl)-4, 4-Biphenylene diphosphonate is used as a stabilizer for polymers.

5. Conclusion

The most significant outcome of this study was the production of dark goldenrod pigment from *P. Purpureogenum* responded by producing high concentrations of the pigment of *T. Foenum* leaves. The results of the optimization, spectroscopic characterization indicates that the isolated pigment having different phytochemicals were produced having flavoring agents, antioxidant activity. The structural elucidation from GCMS shows that the structure of the main pigment constituent in the pigment. To the best of our knowledge, this is the first study to report red pigment production by *P. Purpureogenum* from *Trigonella* leaves.

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