



E-ISSN: 2278-4136
P-ISSN: 2349-8234
JPP 2019; SP1: 181-185

B Rakavi
Research Scholar, Tamil Nadu
Agricultural University,
Coimbatore, Tamil Nadu, India

N Sritharan
Faculty of Crop Physiology,
Tamil Nadu Agricultural
University, Coimbatore, Tamil
Nadu, India

Correspondence
B. Rakavi
Research Scholar, Tamil Nadu
Agricultural University,
Coimbatore, Tamil Nadu, India

(Special Issue- 1)
2nd International Conference
**“Food Security, Nutrition and Sustainable Agriculture -
Emerging Technologies”**
(February 14-16, 2019)

Physiological response of greengram under heat stress

B Rakavi and N Sritharan

Abstract

Greengram is a major grain legume used for consumption from ancient days for its higher protein content. Due to climate change, high temperature stress seriously affects the productivity of legumes and nutritional balance worldwide. Elevated temperature stress during sensitive crop growth stages affects biochemistry of the crops by production of reactive oxygen species which leads to membrane degradation and less physiological activity that affects the yield in greengram. With this background, the experiment was carried out in CO 8 variety to study the biochemical and yield traits of greengram in the temperature controlled growth chambers. Stress was given by raising the temperature up to 2°C and 4°C from the ambient at different growth stages. Results showed, that an increased production of malondialdehyde and H₂O₂ under elevated temperature than ambient and reduced the photosynthetic rate was also recorded. Antioxidant enzymes *viz.*, Superoxide dismutase and catalase activity were increased under elevated temperature so as to scavenge the free radicals. Yield traits like pod length and weight were drastically reduced in stressed plants. From the results, it was concluded that temperature stress on CO 8 greengram during flowering stage was more critical which caused poor fertility and ultimately resulted in low yield.

Keywords: H₂O₂, malondialdehyde content, photosynthetic rate, antioxidants and yield

Introduction

Pulses are popularly known as Poor man's meat and rich man's vegetable and contribute to nutritional security globally. Food and Agriculture Organization (FAO) declared 2016 as the International Year of Pulses recognizing protein deficiency as a global concern. Greengram is considered as extensively grown pulse crop of India. The percent share in total production of greengram is 9.72 during the year 2015-2016 (Directorate of Economics and Statistics, GOI). The productivity of greengram in India is very low and far below when compared to other greengram growing countries. This is mainly due to various abiotic stress factors. Among the various environmental stresses water and temperature play a major role. Pulses are very sensitive to drought, water logging and high temperature. Many countries could experience unprecedented heat stress due to global climate change (Gaur *et al.*, 2015) [10]. High temperature is implicated as a major limiting factor for yield decline in greengram (Zinn *et al.*, 2010) [25]. The elevated temperature stress causes oxidative damage to leaves due to production of Reactive Oxygen Species (ROS). ROS are produced continuously as byproducts of different metabolic pathways which are located in different cellular compartments such as chloroplast, mitochondria and peroxisomes (Navrot *et al.*, 2007) [18]. The increased ROS, decreased cell membrane permeability in chickpea (Deshmukh *et al.*, 2002) [7]. ROS by causing oxidative damage impairs photosynthetic efficiency, which also affects nitrogen fixing ability of mung bean rhizobia by restricting formation and spread of root hair. The detoxification of these ROS is very important and plants have evolved complex strategies to deal with them (Asthir *et al.*, 2012) [2]. The scavenging enzymes are called as antioxidant enzymes. They remove free radicals and prevent the membranes and DNA from oxidative damage and make the plant to survive under stress. There are various evidences indicating increased activity of antioxidant enzymes under heat stress which is crop specific (Babu *et al.*, 2007) [3]. The major enzymatic antioxidants are superoxide dismutase (SOD) and catalase (CAT) are reported to increase under various environmental stress (Munne-Bosch and Alegre, 2000) [17]. Heat stress during reproductive stage is becoming a serious constraint to grain legumes productivity as their

cultivation is expanding to warmer environment (Gaur *et al.*, 2015) [10]. Global climate change is making elevated temperature a critical factor for plant growth and productivity. In this light of view, the study aims to understand the ROS production, antioxidants, photosynthetic rate and yield of CO 8 variety for studying the physiological and biochemical changes in greengram under elevated temperature stress.

Materials and Methods

The pot culture experiment was conducted during 2017 at the open top chambers located in the Department of Crop Physiology, TNAU, Coimbatore. The latest variety in greengram CO 8 was taken for the experiment. The treatments were ambient temperature that exists under open field condition (T₁), Elevated temperature of 2°C from the ambient temperature (T₂), Elevated temperature of 4°C from the ambient temperature (T₃). The treatments were imposed during Vegetative (S₁), Flowering (S₂) and Pod development stages (S₃). The biochemical parameters and yield traits were recorded under elevated temperature stress. The experiment was laid out in Factorial Completely Randomized Design (FCRD) with four replications.

The lipid peroxidation level was determined by quantifying the malondialdehyde equivalents using 2-Thiobarbituric acid (TBA), as described by Hodges *et al.*, (1999) [12] and expressed in $\mu\text{mol g}^{-1}$ of fresh weight. H₂O₂ accumulation in leaves was visually detected by staining with 3,3-diaminobenzidine (DAB) using the method as described by Thordal Christensen *et al.* (1997) [20]. Leaves were immediately removed from plants, submerged in DAB solution (1 mg ml⁻¹ at pH 3.8) and incubated under light for six hours at 25° C for reaction and examined under light microscope by using LEICA 6SD. Photosynthetic gas exchange was measured from non-detached young and fully expanded leaves using a portable photosynthetic system (PPS) (ADC Bio-Scientific Ltd.) and expressed in $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$. Catalase activity was measured by the method described by Aebi (1974) [1] and expressed in $\mu\text{g H}_2\text{O}_2 \text{ min}^{-1} \text{ g}^{-1}$. Superoxide dismutase activity was determined by using nitro blue tetrazolium (NBT) salt as described by Champ and Fridovich (1971) [4] and expressed as enzyme units mg⁻¹ protein. The important components contributing to the yield potential of the crop were recorded at harvest. For pod length ten pods per plant were randomly taken from four selected plants at maturity and it was measured in cm. For pod weight number of pods produced in each plant was taken randomly from four plants in each replication from each treatment at the harvest stage. The total pod weight was weighed and expressed in gram per plant.

Results and Discussion

In this experiment, malondialdehyde (MDA) content was measured to determine the lipid peroxidation level because MDA is a byproduct of lipid peroxidation.

The content of MDA and hydrogen peroxide (H₂O₂) has been considered as an indicator of oxidative injury (Mandhania *et al.*, 2006; Moller *et al.*, 2007) [15, 16]. Results showed that T₃ have more MDA than T₁ and T₂. Among the treatments, T₁ (0.48 at S₁, 0.93 at S₂, 0.63 at S₃) was observed to have lowest value. The treatment T₃, (1.85 at S₁, 2.06 at S₂, 1.77 at S₃) recorded highest MDA content than other treatments (Fig.1.). We can visualize (Plate.1.) the production of H₂O₂ concentrations remained almost constant in leaves under ambient temperature condition in all the stages of plant growth. However, under elevated temperature, substantial

increase in H₂O₂ accumulation was observed. More accumulation of H₂O₂ observed in T₂ and T₃ at S₁, S₂ and S₃ than in T₁ plants where ROS accumulation strikingly lesser when compared with others. Among the treatments, more H₂O₂ accumulation was observed in T₃. Heat stress impairs mitochondrial functions thereby resulting in the induction of oxidative damage that manifests in lipid peroxidation, detected by malondialdehyde (MDA) content (Larkindale and Knight 2002; Vacca *et al.*, 2004) [14, 21].

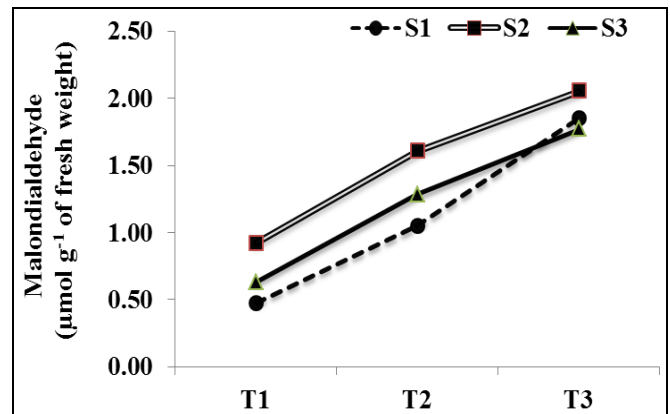


Fig 1: Effect of elevated temperature on malondialdehyde content

T₁: Ambient temperature that exists under open field condition

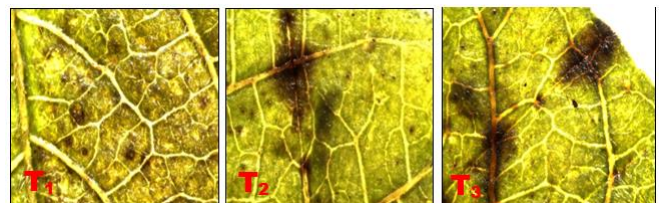
T₂: Elevated temperature of 2°C from the ambient temperature

T₃: Elevated temperature of 4°C from the ambient temperature

S₁: Vegetative stage

S₂: Flowering stage

S₃: Pod development stage



The *in situ* detection of H₂O₂ in greengram leaves at vegetative stage



The *in situ* detection of H₂O₂ in greengram leaves at flowering stage



The *in situ* detection of H₂O₂ in greengram leaves at pod development stage

Plate 1: Effect of elevated temperature in H₂O₂ production

Photosynthetic rate was recorded during vegetative (S₁) and flowering stages (S₂) alone. In control (T₁) the photosynthetic rate found to be 16.8 at S₁ and 23.5 at S₂.

The photosynthetic rate was reduced under elevated

temperature in T₂ (13.5 at S₁, 21.8 at S₂) and in T₃ (11.3 at S₁, 17.0 at S₂). Among the treatments, plants grown in ambient temperature condition (T₁) performed better in all the stages when compared to elevated temperature condition (T₂ and T₃) (Fig.2.). The reduction of whole leaf photosynthesis by high temperature might be caused by disruption of the functional integrity of the photosynthetic apparatus associated with the production of Reactive Oxygen Species (Camejo *et al.*, 2005; Guo *et al.*, 2006) [5, 11] that cause damage to the thylakoid membrane. The results of the present study is similar to the findings of Djanaguiraman *et al.* (2011) [9] who found that high temperature stress during flowering in soybean plants decreased the photosynthetic rate by 20.5 per cent compared with those grown at ambient temperature. The results of the present study lead to the conclusion that decrease in photosynthetic rates are more pronounced when temperature stress was applied during flowering stage (S₂).

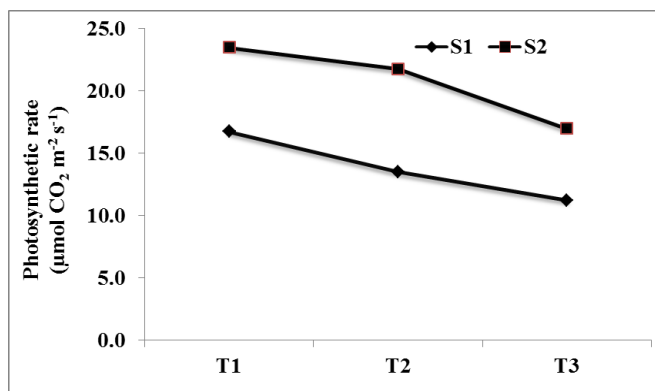
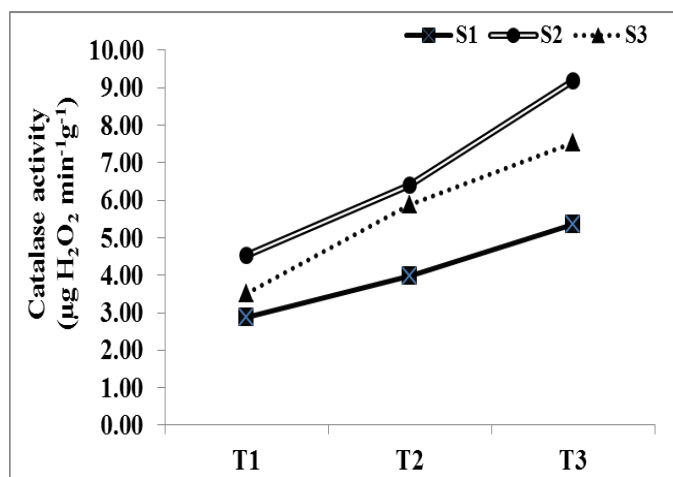
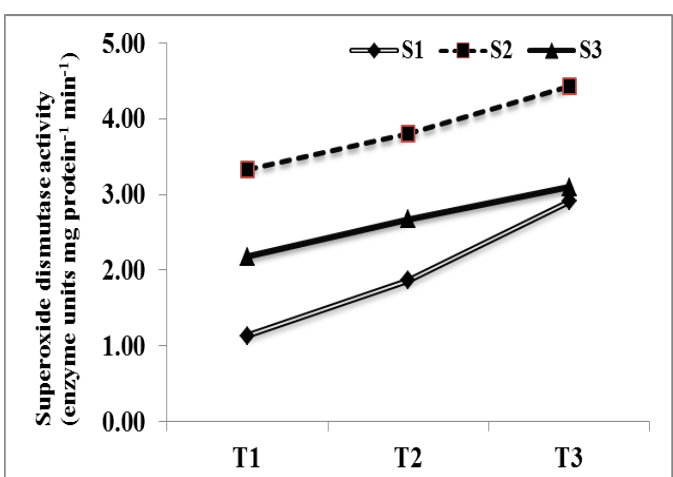


Fig 2: Effect of elevated temperature on Photosynthetic rate

Plants use antioxidant enzymes to detoxify reactive oxygen species and mitigate oxidative stress-induced damage under elevated temperature (Shah *et al.*, 2001) [19]. Many studies demonstrated that high temperature injury was caused by the excessive production of reactive oxygen radicals, the low activities of antioxidant enzymes, and the membrane damage in plants (Zhang *et al.*, 2006; Zhu *et al.*, 2005) [11, 23, 24]. Antioxidants activity shows the plants condition and stress level. Activities of different antioxidant enzymes are temperature sensitive and activation occurs at different temperature ranges but the activities of these enzymes increase with increasing temperature (Chakraborty and Pradhan, 2011) [6]. Similar finding was observed in the present investigation, Catalase and Superoxide dismutase activity (SOD) was increased under temperature stress condition (Fig.3a and 3b). The CAT activity of T₃ (5.36 at S₁, 9.18 at S₁, 7.53 at S₁) had lower activity than T₂ and T₁. Whereas T₁ (2.89 at S₁, 4.54 at S₂, 3.51 at S₃) showed its better performance than other treatments. The data on CAT activity in T₂ (3.99 at S₁, 6.41 at S₂, 5.89 at S₃) was higher than T₃. SOD also showed similar trend that under elevated temperature, the enzyme SOD activity was high in T₃ (2.91 at S₁, 4.43 at S₂, 3.10 at S₃) whereas T₁ recorded (1.13 at S₁, 3.33 at S₂, 2.18 at S₃) the lowest SOD activity. The per cent increase in SOD activity was 39.2 at vegetative stage (S₁), 12.4 at flowering stage (S₂) and 18.4 at pod development stage (S₃). When the temperature was increased to 4°C, the SOD activity still enhanced to 61.2% increase at vegetative stage, 24.8% increase at flowering stage and 29.7% increase at pod development stage. This explains that the plants are trying to survive under elevated temperature to maintain its growth and development process.



a. Catalase activity



b. Superoxide dismutase activity

Fig 3: Effect of elevated temperature on

The brief exposure of high temperature stress (32-35°C) in chickpea reduced pod set and hence grain yield in the controlled environments (Devasirvatham *et al.* 2010) [8]. In results, T₁ (6.46) showed higher pod length than other treatments (Table.1.). In both the treatments, T₂ (6.12 at S₁, 5.53 at S₂, 5.39 at S₃) and T₃ (5.72 at S₁, 4.14 at S₂, 4.59 at S₃) the pod length were reduced than T₁. Especially the plants where elevated temperature (T₃) imposed during pod development stage (S₃) have recorded the lowest pod length. Total Pod weight showed similar trends that T₁ (4.24) recorded maximum pod weight showing its better

performance under ambient temperature condition. Plants imposed with elevated temperature have recorded the lowest number of pod weight with a mean value of 2.83 (T₂) and 2.11 (T₃). Elevated temperature imposed at vegetative stage did not show much reduction in the yield components. But the impact was greater when plants exposed to temperature stress at flowering stage and pod development stage (Plare.2). Similar results observed by Kumar *et al.* (2013) [13] in chickpea. Our results confirmed by the earlier findings of the Wang *et al.* (2006) [22] and Kumar *et al.* (2013) [13].

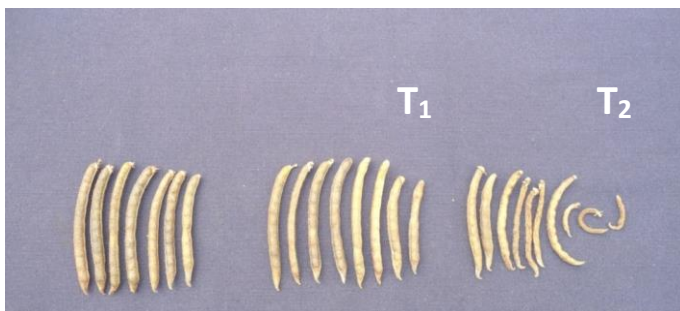
Table 1: Effect of elevated temperature on Yield traits

Treatments	Total Pod weight (g plant ⁻¹) Crop growth stages				Hundred seed weight (g) Crop growth stages			
	S ₁	S ₂	S ₃	Mean	S ₁	S ₂	S ₃	Mean
T ₁	4.24				3.75			
T ₂	3.82	2.83	3.24	3.29	3.59	3.45	3.43	3.48
T ₃	3.27	2.11	2.31	2.56	3.26	2.93	3.20	3.12
Mean	3.77	3.05	3.26	3.36	3.53	3.37	3.45	3.45
		T	S	T×S	T	S	T×S	T
SED		0.18	0.18	0.32	0.06	0.06	0.11	0.06
CD (P<0.05)		0.38**	0.38*	0.66 ^(NS)	0.13**	0.13 ^(NS)	0.22 ^(NS)	0.13**

*, ** and NS denote significance level at $P<0.05$, $P<0.01$ and non-significant respectively



Stress imposed at vegetative stage



Stress imposed at flowering stage



Stress imposed at pod development stage

Plate 2: Effect of elevated temperature on yield (pods) during different crop growth stages

Conclusion

The results of this study showed that high temperature caused negative effect on growth which could be due to the generation of high levels of ROS. Antioxidants produced scavenged the ROS and made the plants to survive under elevated temperature stress at an extent. From the present study, it was concluded that, when greengram crop undergoes to an elevated temperature of than 2°C and 4°C from the ambient, showed significant changes in its physiology, biochemical and yield attributes.

References

1. Aebi H. Catalases. *In*: Bergmeyer, H.U. (ed) Methods of enzymatic analysis. Academic Press. New York. 1974; 2:673-684.

- Asthir B, Koundal A, Bains NS. Putrescine modulates antioxidant defense response in wheat under high temperature stress. *Biol. Plant.* 2012; 56:757-761.
- Babu S, Sheeba A, Yogameenakshi P, Anbumalarnathi J, Rangasamy P. Effect of salts stress in the selection of salt tolerant hybrids in (*Oryza sativa* L.) under *in vitro* and *in vivo* condition. *Asian J Plant Sci.* 2007; 6(1):137-142.
- Beau-Champ C, Fridovich I. Superoxide dismutase: Improved assays and assay applicable to acrylamide gels. *Annual Biochemistry.* 1971; 44:276-87.
- Camejo D, Rodriguez P, Angeles MM, Dell, Amico JM, Torrecillas A, Alarcon JJ. High temperature effects on photosynthetic activity of two tomato cultivars with different heat susceptibility. *J Plant Physiol.* 2005; 167:281-289.
- Chak:aborty U, Pradhan D. High temperature-induced oxidative stress in *Lens culinaris*, role of antioxidants and amelioration of stress by chemical pre-treatments. *Journal of Plant Interactions.* 2011; 6(1):43-52.
- Deshmukh PS, Kushwaha SR. Variability in membrane injury index in chickpea genotypes. *Indian J Plant Physiol.* 2002; 7(3):285-287.
- Devasirvatham V, Tan DKY, Trethowan RM, Gaur PM, Mallikarjuna N. Impact of high temperature on the reproductive stage of chickpea. *In*: 'Food security from sustainable agriculture. Proceedings of the 15th Australian Society of Agronomy Conference, Lincoln, New Zealand, 2010, 15.
- Djanaguiraman M, Prasad PVV, Boyle DL, Schapaugh WT. High-temperature stress and soybean leaves: Leaf Anatomy and Photosynthesis. *Crop Sci.* 2011; 51:2125-2131.
- Gaur PM, Samineni S, Krishnamurthy L, Kumar S, Michel Ghanem E, Stephen Beebe E *et al.* High temperature tolerance in grain legumes. *Legume Perspectives.* 2015; 7:23-24.
- Guo YP, Zhou HF, Zhang LC. Photosynthetic characteristics and protective mechanisms against photo oxidation during high temperature stress in two Citrus species. *Sci. Hort.* 2006; 108:260-267.
- Hodges D, DeLong J, Forney C, Robert Prange K. Improving the thiobarbituric acid reactive-substances assay for estimating lipid peroxidation in plant tissues containing anthocyanin and other interfering compounds. *Planta.* 1999; 207(4):604-611
- Kumar N, Nandwal AS, Waldia RS, Kumar S, Devi S, Singh S *et al.* High Temperature tolerance in chickpea genotypes as evaluated by membrane integrity, heat susceptibility index and chlorophyll fluorescence techniques. *Ind. J Agrl. Sci.* 2013; 83(4):467-471.
- Larkindale J, Knight MR. Protection against heat stress induced oxidative damage in *Arabidopsis* involves calcium, abscisic acid, ethylene and salicylic acid, *Plant*

- Physiol. 2002; 128:682-695.
15. Mandhania S, Madan S, Sawhney V. Antioxidant defense mechanism under salt stress in wheat seedlings. *Biol. Plant.* 2006; 50:227-231.
 16. Moller IM, Jensen PE, Hansson A. Oxidative modifications to cellular components in plants. *Ann. Rev. Plant Biol.* 2007; 58:459-481.
 17. Munné-Bosch S, Alegre L. Changes in carotenoids, tocopherols and diterpenes during drought and recovery, and the biological significance of chlorophyll loss in *Rosmarinus officinalis* plants. *Planta.* 2000; 210:925-931.
 18. Navrot N, Rouhier N, Gelhaye E, Jacquot J. Reactive oxygen species generation and antioxidant systems in plant mitochondria. *Physiol. Plant.* 2007; 129:185-195.
 19. Shah K, Kumar RG, Verma S, Dubey RS. Effect of cadmium on lipid peroxidation, superoxide anion generation and activities of antioxidant enzymes in growing rice seedlings. *Plant Science.* 2001; 161(6):1135-1144.
 20. Thordal Christensen H, Zhang ZG, Wei YD, Collinge DB. Subcellular localization of H₂O₂ in plants. *Plant Journal.* 1997; 11:1187-1194.
 21. Vacca RA, De Pinto MC, Valenti D, Passarella S, Maria E, De Gara L. Production of reactive oxygen species, alteration of cytosolic Ascorbate peroxidase, and impairment of mitochondrial metabolism are early events in heat shock induced programmed cell death in tobacco Bright Yellow 2 cells. *Plant Physiol.* 2004; 134:1100-1112.
 22. Wang J, Gan YT, Clarke F, McDonald CL. Response of chickpea yield to high temperature stress during reproductive development. *Crop Sci.* 2006; 46:2171-2178.
 23. Zhang Y, Mian MAR, Bouton JH. Recent molecular and genomic studies on stress tolerance of forage and turf grasses. *Crop Sci.* 2006; 46:497-511.
 24. Zhu X, Gong H, Chen G, Wang S, Zhang C. Different solute levels in two spring wheat cultivars induced by progressive field water stress at different developmental stages. *J Arid Environ.* 2005; 62:1-14.
 25. Zinn KE, Tunc-Odemir M, Harper JF. Temperature stress and plant sexual reproduction: uncovering the weakest links. *J Expt. Bot.* 2010; 61:1959-1968.