



E-ISSN: 2278-4136
P-ISSN: 2349-8234
JPP 2019; 8(3): 4740-4745
Received: 04-03-2019
Accepted: 06-04-2019

Punita Sharma
Maharaja Agrasen University,
Atal Shiksha Kunj, Kalujhanda,
Solan, Himachal Pradesh, India

Abhishek Awasthi
Maharaja Agrasen University,
Atal Shiksha Kunj, Kalujhanda,
Solan, Himachal Pradesh, India

Ishita Gupta
Maharaja Agrasen University,
Atal Shiksha Kunj, Kalujhanda,
Solan, Himachal Pradesh, India

Phytochemical investigation and evaluation of anti-oxidant potential of *Berberis lycium* roots from Himachal Pradesh

Punita Sharma, Abhishek Awasthi and Ishita Gupta

Abstract

Berberis lycium, a thorny evergreen shrub, commonly known as kashmal, found in subtropical and temperate regions of the world and mentioned in Indian traditional system of medicine. The present study deals with the preliminary phytochemical evaluation of different extracts of roots of *B. lycium*, isolation and identification of bioactive constituents from root extracts, its fractions and evaluation of their anti-oxidant potential by using hydrogen peroxide radical scavenging assay and reducing power assay. Extracts were fractionated and screened for their phytochemicals which resulted in isolation and identification of Stigmasterol (steroid), berberine, palmatine and jatrorrhizine (alkaloids), 4-methyl-7-hydroxycoumarin (coumarin) and 4,4-dimethylhexadeca-3-ol, responsible for the good anti-oxidant potential of the roots of *B. lycium*.

Keywords: *Berberis lycium*, phytochemical screening, isolation, phytoconstituents, anti-oxidation activity

Introduction

Since the time immemorial, human beings depend completely upon nature for their livelihood. Ancient civilizations valued plants the most as they supply humans with food, shelter and medical treatment. Studies show that maximum population of the world depends upon plant based medicine systems for their healthcare [1]. Different ethnic groups have their own traditional medical knowledge and experiences, on the basis of which they have developed different system of medicines [2, 3]. India is known for its diversity, where a number of traditional systems of medicine were invented (Ayurveda, Yoga, Naturopathy, Homeopathy, Siddha and Unani), flourished and practiced till date. India has varied climatic conditions and different composition of soil in different geographical regions that have caused a wide distribution of medicinal plant species which is why the people living over here have relied on plants for treatment of their medical implications. *Berberis lycium* Royle is a shrub that grows in the Himalayas from Kashmir to Kumaun at altitudes ranging between 900-2700m, belongs to family Berberidaceae [4]. Ancient manuscripts based on traditional medicinal system of India, i.e. Charka Samhitta, *B. lycium* is referred as “lekhaniya” for reducing obesity and scarifying, “arsaghna” for curing piles and haemorrhoids, and “kandughna”, curative intense itching sensation that can have various causes whereas according to Sushruta Samhitta, it is used for the treatment of uterine disorders, dysentery, indigestion and quick healing of wounds [5]. Dried decoction of roots is known as ‘Rasount’ which is reported to cure many disease such as boils, conjunctivitis, piles, leprosy, oral ulcers, liver disorders, kidney, chest and throat troubles, chronic diarrhoea, ophthalmia and skin diseases [6]. Previous studies shows that *B. lycium* plant has a wide range of biological activities including anti-hyperglycemic [7-8], anti-hyperlipidemic [9-10], anti-cancer and anti-tumor [11-12], wound and bone healing [13-14], anti-microbial [15-16], anti-coccidial [17], anti-oxidant [18-19], immunity enhancing [20-21], hepatoprotective [21], anti-urolithic and anthelmintic [22]. As mentioned in the earlier reported literature, *B. lycium* is found to be an important herb which shows numerous biological activities and used widely as folk medicine in Himachal Pradesh, India. In Himachal Pradesh, it is found in abundance in forest area and roadside. These has prompted us to carry out research to explore *B. lycium* found in Himachal Pradesh chemically and determine its anti-oxidant potential using *in-vitro* assays like hydrogen peroxide scavenging assay and reducing power assay.

Correspondence

Punita Sharma
Maharaja Agrasen University,
Atal Shiksha Kunj, Kalujhanda,
Solan, Himachal Pradesh, India

Material and Methods

Collection, extraction and fractionation of plant material

The roots of *Berberis lycium* have been collected from Haripurdhar, Himachal Pradesh, India. Collected roots were dried under shade and kept free from foreign matter like soil, dust, insect, fungal and other extrinsic contamination. Dried plant material was grounded to coarse powder. Coarsely grounded plant material (1kg) was extracted with 95% ethanol in soxhlet for 48 hours. Extract after removal of ethanol under reduced pressure yielded hot ethanolic extract (190g) and subjected to fractionation by dissolving the extract in water and the solution was successively extracted with petroleum ether (three times), chloroform (three times) and n-Butanol (three times) in a separating funnel. The petroleum ether, chloroform and n-butanol fractions were distilled under reduced pressure to yield the residues petroleum ether (5g), chloroform (26g) and n-butanol (20g) fractions. The hot ethanolic extract and its fractions were taken for phytochemical screening.

Phytochemical Screening

The hot *ethanolic* extract and its fractions i.e. Petroleum ether, Chloroform and n-butanol fractions, were subjected to qualitative phytochemical analysis for the identification of phytoconstituents using standard tests for alkaloids (Dragendorff's reagent), tannins (Ferric Chloride test), saponins (Froth test), anthraquinones (Borntrager's reaction), flavanoids (NaOH test), carbohydrates (Benedict's reagent

and Molisch's reagent test), protein (Biuret test), steroids (Liebermann-Burchard's and Salkowski's test), terpenoids (Salkowski's test) and glycosides (Keller-killani test)^[23-27].

Isolation of Phytoconstituents

Isolation of phytoconstituents from the roots of *B. lycium* was done according to the method of Miana AG, (1973) with a little modification^[28]. The hot ethanolic extract (70g) was dissolved in 1L of water and acidified with dilute HCl upto pH 4-5. Acidified extract was kept for precipitation under low temperature overnight. Yellow colored precipitates were formed which were subjected to filtration. Thin layer chromatography of the precipitates was carried out using Dragendorff's reagent as visualizing agent showing three major spots indicating it to be a mixture of alkaloids which were separated by repeated column chromatography resulting in isolation of Compound 1 (eluted in Chloroform:Methanol:: 2:23), compound 2 (eluted in CHCl₃:MeOH:: 87:13) and compound 3 (eluted in CHCl₃:MeOH:: 4:1). The petroleum ether fraction of hot ethanolic extract yielded compound 4 when subjected to column chromatography using Petroleum ether: Ethyl acetate as eluting solvent while compound 5 and 6 were isolated from Chloroform fraction of hot ethanolic extract. The All the precipitated compounds were subjected to physical and spectroscopic analysis using melting point, UV-VIS spectrophotometer and ¹H and ¹³C- NMR for identification.

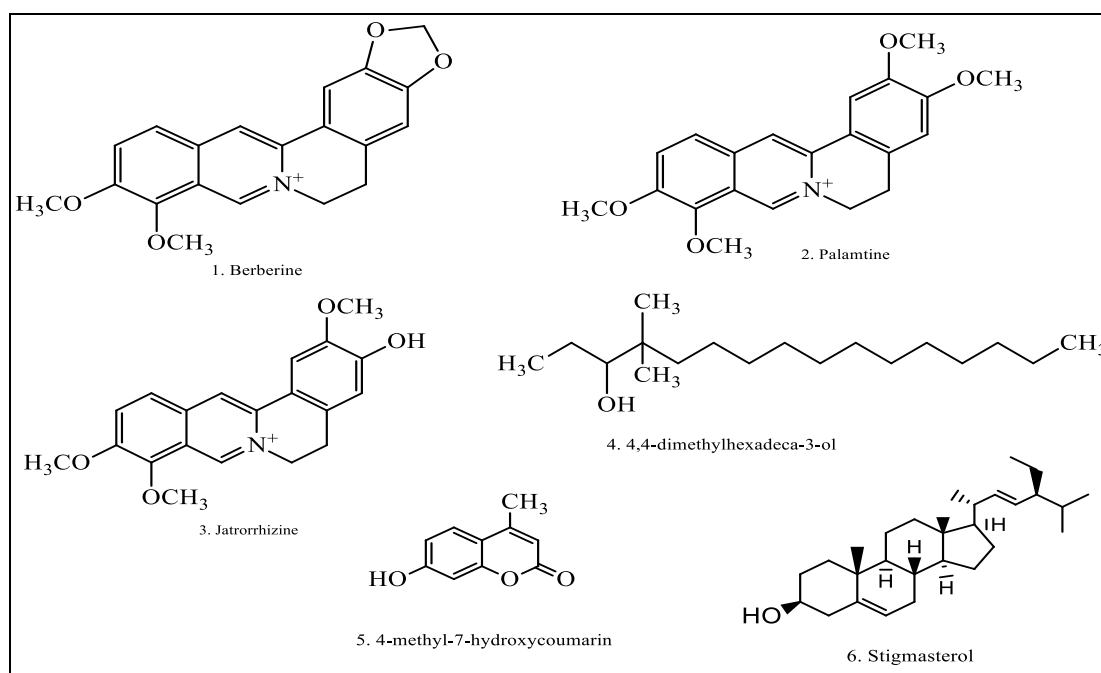


Fig 1: Structures of compounds isolated from different fractions of roots of *B. lycium*

Biological Activities

In-vitro anti-oxidant activity

Hydrogen peroxide scavenging (H₂O₂) assay

Earlier reports reveals that humans consume about 0.28mg/Kg/day hydrogen peroxide directly or indirectly from environment, that becomes toxic due to formation of hydroxyl radicals that can cause DNA damage in human body and initiates lipid peroxidation resulting into redox imbalance and oxidative stress. Thus, removing H₂O₂ or to terminate the long chain reactions of free radicals formed by it is an essential step^[29-30]. Antioxidants have capability to terminate the production of free radicals by donating electrons to free

radicals and stabilizing them that causes balance in the environment for proper cell functioning. The most important and common source of antioxidants is plants and fruits that are enriched which antioxidants and regulates proper functioning of body by resisting our body against various diseases^[30-31]. Plants can scavenge Hydrogen peroxide due to the presence of secondary metabolites that acts as antioxidants. The H₂O₂ scavenging potential of the different extracts of roots of *B. lycium* was estimated according to the method mentioned by Alam MN *et al.*, (2013)^[30-31].

Hydrogen Peroxide (40mM) solution was prepared in phosphate buffer (50mM, pH 7.4) whose concentration was

determined from Hydrogen peroxide calibration curve by measuring absorbance at 230nm using UV-VIS spectrophotometer. Plant extract of different concentrations (0.2-1.0 mg/ml) were prepared in distilled water and hydrogen peroxide containing phosphate buffer was added to these. Absorbance of the reaction mixtures at 230 nm was determined after 10 minutes against a blank solution containing phosphate buffer without hydrogen peroxide. The percentage of Hydrogen Peroxide Scavenging by the plant extract was calculated as follows:

$$\% \text{ scavenged (H}_2\text{O}_2) = [(A_c - A_t)/A_s] \times 100$$

Where A_c is the absorbance of control and A_s is the absorbance of the test.

Determination of the reducing power

Substances having reduction potential can react with potassium ferricyanide (Fe^{3+}) to form potassium ferrocyanide (Fe^{2+}) which reacts with FeCl_3 on addition and forms a ferric-ferrous complex (Perls's Prussian blue) which can be spectrophotometrically measured at 700nm. The free-radical scavenging activity of the reducing power of ethanolic extract and its fractions of roots of *B. lycium* was determined according to the method of Oyaizu (1986) described by Alam *et al.*, (2013) [30, 32]. Various concentrations (2.5 mg-12.5mg) of root extracts of *B. lycium* were prepared in methanol and

0.2 M Phosphate Buffer (5ml, pH 6.6) was added to it followed by addition of 1% potassium ferricyanide (5ml). The resulting mixture was incubated at 50°C for 20min and 10% trichloroacetic acid (5ml) was added to it. The incubated reaction mixture was centrifuged at 3000g for 10 min and the supernatant (5ml) was mixed with distilled water (5ml) and 1% ferric chloride (1ml). The absorbance of the final mixture was measured at 700nm using UV-VIS spectrophotometer. L-ascorbic acid at different concentrations was used as standard. All the tests were carried out in triplicates.

Results and Discussions

Qualitative analysis of phytoconstituent of *Berberis lycium* roots:

Hot ethanolic extract of roots of *Berberis lycium* collected from Himachal Pradesh were found to be rich in secondary metabolites like alkaloids, flavonoids, steroids, terpenoids and their glycosides whereas tannins, saponins, anthraquinones and anthocyanides are completely absent. Ethanolic extract was fractionated successively with Petrol, Chloroform and n-butanol to yield respective fractions. Phytochemical screening results of different fractions reveal that petrol fraction contains only alkaloids whereas chloroform fraction is rich in alkaloids, flavonoids, terpenoids and steroids while n-butanol fraction shows the presence of alkaloids and terpenoids. Detailed results of qualitative analysis of phytoconstituents of *B. lycium* roots are shown in Table 1.

Table 1: Preliminary phytochemical screening of ethanolic extracts and its fractions of roots of *B. lycium*

Phytochemicals	Test performed	Ethanol Extract	Fractions of ethanol extract of <i>B. lycium</i>		
			Petrol	CHCl ₃	n-BuOH
Alkaloids	Dragendorff's reagent	+	+	+	+
Tannins	Ferric Chloride test	-	-	-	-
Saponins	Froth test	-	-	-	-
Anthraquinones	Bortrager's reaction	-	-	-	-
Flavonoids	NaOH test	+	-	+	-
Carbohydrates	Benedict's reagent	+	+	+	+
Proteins	Biuret test	+	-	-	+
Steroids	Liebermann-Burchard test	+	-	+	-
Terpenoids	Salkowski's test	+	-	+	+
Glycosides	Keller-killani test	+	-	+	-

Isolation and characterization of Phytoconstituent

Berberine, palmatine and jatrorrhizine were isolated from the ethanolic extract of roots of *B. lycium* whereas 4,4-dimethylhexadeca-3-ol was isolated from petrol fraction and 4-methyl-7-hydroxycoumarin and stigmasterol isolated from chloroform fraction. The isolated compounds were identified and characterized by using different chromatographic, spectroscopic techniques and comparing the results with existing literature [19, 33]. The experimental data of the isolated compounds is as under.

- Berberine:** Yellow crystals, soluble in methanol. Molecular formula [$\text{C}_{20}\text{H}_{18}\text{NO}_4^+$], Melting Point- 198°C. UV (MeOH), λ_{max} (nm) = 236, 357, 469. ¹H-NMR (DMSO, 400MHz), δ (ppm):- 3.20 (t, 2H, J = 6.1 Hz, H-5), 4.05 (s, 3H, -OCH₃), 4.09 (s, 3H, -OCH₃), 4.94 (t, 2H, J = 6.2 Hz, H-6), 6.15 (s, 2H, -OCH₂O-), 7.06 (s, 1H, H-4), 7.75 (s, 1H, H-1), 7.99 (d, 1H, J = 9.12 Hz, H-11), 8.16 (d, 1H, J = 9.16 Hz, H-12), 8.93 (s, 1H, H-13), 9.89 (s, 1H, H-8).
- Palmatine:** Yellow crystals, soluble in methanol. Molecular formula [$\text{C}_{21}\text{H}_{22}\text{NO}_4^+$], Melting Point- 217°C. UV (MeOH), λ_{max} (nm) = 231, 351, 441. ¹H-NMR (DMSO, 400MHz), δ (ppm):- 3.20 (t, 2H, J = 6.40 Hz, H-

5), 4.04 (m, 6H, 2-OCH₃), 4.08 (m, 6H, 2-OCH₃), 4.95 (t, 2H, J = 6.0 Hz, H-6), 7.05 (s, 1H, H-4), 7.74 (s, 1H, H-1), 7.99 (d, 1H, J = 9.08 Hz, H-11), 8.15 (d, 1H, J = 9.12 Hz, H-12), 8.93 (s, 1H, H-13), 9.89 (s, 1H, H-8).

- Jatrorrhizine:** Yellow crystals, soluble in methanol. Molecular formula [$\text{C}_{20}\text{H}_{20}\text{NO}_4^+$], Melting point- 236°C. UV (MeOH), λ_{max} (nm) = 229, 307, 436. ¹H-NMR (DMSO, 400MHz), δ (ppm):- 3.16 (s, 3H, -OCH₃), 3.20 (t, 2H, J = 6.1 Hz, H-5), 4.06 (s, 3H, -OCH₃), 4.09 (s, 3H, -OCH₃), 4.94 (t, 2H, J = 6.04 Hz, H-6), 6.16 (s, 1H, C3-OH), 7.07 (s, 1H, H-4), 7.76 (s, 1H, H-1), 7.96 (d, 1H, J = 9.12 Hz, H-11), 8.18 (d, 1H, J = 9.16 Hz, H-12), 8.92 (s, 1H, H-13), 9.88 (s, 1H, H-8).
- 4,4-dimethylhexadeca-3-ol:** White sticky mass, soluble in ethyl acetate. Molecular formula [$\text{C}_{18}\text{H}_{38}\text{O}$], Melting point- 162°C. ¹H-NMR (CDCl₃, 400MHz), δ (ppm):- 0.46 (s, 3H, -CH₃), 1.02 (t, 3H, J = 7.16 Hz, -CH₃), 1.96 (m, 3H, -CH₃), 2.36 (t, 2H, J = 5.18 Hz, -CH₂), 2.56 (m, 2H, -CH₂), 4.19 (t, 3H, J = 5.14 Hz, -CH₃). ¹³C-NMR (CDCl₃, 300MHz), δ (ppm): 15.36 (CH₃, C-16), 19.16 (CH₃, C-1), 22.91 (CH₃, C-17 and C-18), 31.15 (CH₂, C-2), 33.61 and 33.63 (CH₂, C-5 and C-15), 37.14 (C, C-4), 75.23 (CH, C-3).

5. **4-methyl-7-hydroxycoumarin:** White crystals, soluble in chloroform. Molecular formula [C₁₀H₈O₃], Melting Point- 189°C. ¹H-NMR (CDCl₃, 400MHz), δ (ppm):- 3.28 (s, 3H, -CH₃), 3.59 (s, 1H, H-3), 6.19 (s, 1H, H-8), 6.61 (d, J = 6.63 Hz, 1H, H-5), 7.51 (d, J = 6.69 Hz, 1H, H-6). ¹³C-NMR (CDCl₃, 400MHz), δ (ppm): 19.07 (CH₃, C-8), 102.35 (CH, C-8), 114.09 (C, C-4a), 115.61 (CH, C-3), 117.31 (CH, C-6), 129.06 (CH, C-5), 153.95 (C, C-8a), 156.87 (C, C-4), 166.78 (C, C-7), 167 (C, C-2).
6. **Stigmasterol:** White solid, soluble in chloroform. Molecular formula [C₂₉H₄₈O], Melting point- 186°C. ¹H-NMR (CDCl₃, 400MHz): 0.66 (s, 3H, H-18), 0.72 (d, 3H, J = 6.32 Hz, H-27), 0.80 (d, 3H, J = 6.33 Hz, H-26), 0.83 (t, 3H, J = 7.26 Hz, H-29), 1.06 (s, 3H, H-19), 1.13 (d, 3H, J = 6.3 Hz, H-21), 3.52 (m, 1H, H-3), 5.36 (s, 1H, H-6), 5.09 (dd, 1H, J = 15.19 and 8.36 Hz, H-23), 5.11 (dd, 1H, J = 15.17 and 8.38 Hz, H-22).

Biological Activities

In-vitro anti-oxidant activity

Hydrogen peroxide scavenging activity

Hydrogen peroxide scavenging potential of plant extracts is due to the secondary metabolites present in them which can donate electrons to the free radicals generated by hydrogen peroxide, thus neutralizing free radicals to water. Current study reveals that hot ethanolic extract of roots of *B. lycium* and its fractions were capable of scavenging hydrogen peroxide in a concentration-dependent manner. The scavenging activity of ethanolic extract and its fractions at 0.8mg/ml concentration is above 40%. At 1.0 mg/ml

concentration all the fractions except petroleum ether fraction exhibit more than 50% scavenging activity indicating the roots of *B. lycium* to be a good antioxidant. The inhibiting percentage of various extracts is in the following order: ethanol extract > Chloroform fraction > n-butanol fraction > petrol fraction (Fig 2).

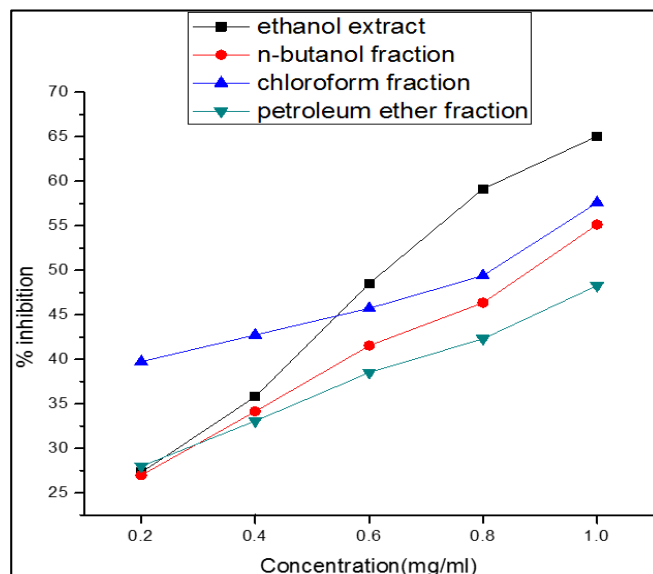


Fig 2: Percentage of H₂O₂ scavenging by alcoholic extract and its fractions of roots of *B. lycium* at different concentrations

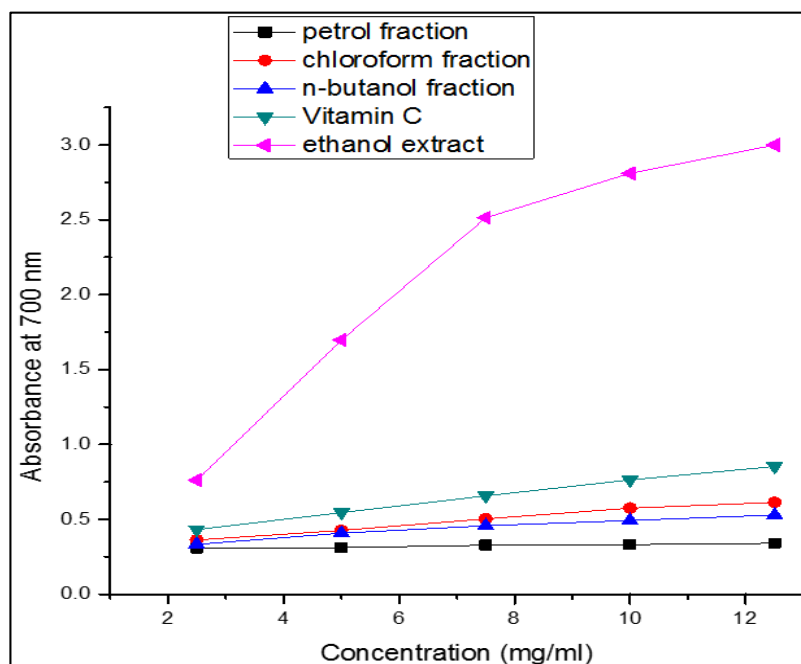


Fig 3: Reducing powers of alcoholic extract and its fractions of roots of *B. lycium* at different concentrations

Reducing power activity

The reducing capacity of the ethanolic extract and its fractions of roots of *B. lycium* were compared with Vitamin C (L-ascorbic acid). A higher value of absorbance indicates a stronger reducing power of the extracts/fractions. The reduction potential of ethanolic extract and its fractions, i.e. petroleum ether, chloroform and n-butanol fractions, of the roots of *B. lycium* at various concentrations is illustrated in Fig 3. All extracts have shown good reducing powers that are comparable with L. The reducing powers of the samples are

in the following order: Ethanol extract > CHCl₃ fraction > n-butanol fraction > petroleum ether fraction. The ethanolic extract of the roots of *B. Lycium* had shown better reducing power than Vitamin C. On the basis of present investigation it was found that the reducing powers of ethanolic extract and its fractions also increased with the increase of their concentrations. So, it can be concluded that extract and its fractions exhibited increase in their reducing potential in concentration dependent manner.

Conclusion

Berberis lycium is a significant herb mentioned in Indian Pharmacopeia and has great ethno medicinal values. In Himachal Pradesh, *Berberis lycium* is a main source of primary healthcare in the rural area, its ripened fruits are edible and its tender shoots are chewed for curing skin diseases and a blood purifier³⁴. In this study, phytochemical investigation of roots of *B. lycium* was carried out showing that the roots are rich in secondary metabolites such as alkaloids, flavonoids, steroids and Terpenoids which resulted in the isolation of stigmaterol (steroid), berberine (alkaloid), palmatine (alkaloid), jatrorrhizine (alkaloid), 4-methyl-7-hydroxycoumarin (coumarin) and 4,4-dimethylhexadeca-3-ol. Hot ethanolic extract was also used for determination of free radical scavenging or its antioxidant profile (Hydrogen peroxide scavenging assay) and reducing power that resulted in good anti-oxidant potential of this plant indicating ethanolic extract to display maximum potential among ethanolic extract and its fractions. Our study on its active constituents and anti-oxidant potential made us to conclude that *B. lycium* from Himachal Pradesh can be explored to great extent to be developed and used commercially for its medicinal values.

References

- Hunt DI. Ecological ethnobotany: stumbling toward new practices and paradigms. *MASA Journal*. 2000; 16(1):1-13.
- Liu Y, Dao Z, Liu Y, Long C. Medicinal plants used by the Tibetan in Shangri-la, Yunnan, China. *Journal of ethnobiology and ethnomedicine*. 2009; 5(15):doi: 10.1186/1746-4269-5-15.
- Zadeh KA. Overview of national drug policy of Iran. *Iranian Journal of Pharmaceutical Research*. 2003; 2:1-2.
- Anonymous. The wealth of India (Raw materials). Vol 2, Publication and Informatics Directorate, CSIR, New Delhi, 1988, 115-118.
- Dev S. A selection of Prime ayurvedic plant drugs Ancient and Modern Concordance. Anamaya Publishers, New Delhi, 2006, 103-105.
- Sharma R. Medicinal Plants of India An Encyclopedia, Daya Publishing House, Delhi, 2003, 33-34.
- Gulfraz M, Qadir G, Nosheen F, Parveen Z. Antihyperglycemic effects of *Berberis lycium* Royle in alloxan induced diabetic rats. *Diabetologia Croatica*. 2007; 36(3):49-54.
- Mustafaa KG, Ganai BA, Akbar S, Dar MY, Tantry MA, Masood A. The extracts of *Berberis lycium* and diabetes mellitus in alloxan monohydrate induced diabetic rats. *Journal of Pharmacy Research*. 2011; 4(8):2570-2573.
- Ahmed M, Alamgeer, Sharif T, Zabta M, Akbar A. Effect of *Berberis lycium* Royle on lipid profile in alloxan induced diabetic rabbits. *Ethnobotanical Leaflets*. 2009; 13:702-708.
- Rahimi-Madiseh M, Heidarian E, Rafieian-kopaei M. Biochemical components of *Berberis lycium* fruit and its effects on lipid profile in diabetic rats. *Journal of HerbMed Pharmacology*. 2014; 3(1):15-19.
- Khan M, Giessrigl B, Vonach C, Madlener S, Prinz S, Herbaceck I *et al*. Berberine and a *Berberis lycium* extract inactivate Cdc25A and induce α -tubulin acetylation that correlate with HL-60 cell cycle inhibition and apoptosis. *Mutation Research*. 2010; 683(1):123-130.
- Issat T, Jakobisiak M, Golab J. Berberine, a natural cholesterol reducing product, exerts antitumor cytostatic/cytotoxic effects independently from the mevalonate pathway. *Oncology Reports*. 2006; 16:1273-1276.
- Asif A, Kakub G, Mehmood S, Khunum R, Gulfraz M. Wound healing activity of root extracts of *Berberis lycium* Royle in rats. *Phytotherapy Research*. 2007; 21(6):589-591.
- Ahmad A, Mehmood S, Gulfraz M. Bone healing properties of *Berberis lycium* (Royal): A case study. *Case Study and Case Report*. 2011; 1(1):1-5.
- Hussain MA, Khan MQ, Habib T, Hussain N. Antimicrobial activity of the crude root extract of *Berberis lycium* Royle. *Advances in Environmental Biology*. 2011; 5(4):585-588.
- Malik AT, Kamili NA, Chishti ZM, Ahad S, Tantry AM, Hussain RP *et al*. Breaking the resistance of *Escherichia coli*: Antimicrobial activity of *Berberis lycium* Royle. *Microbial Pathogenesis*. 2017; 102:12-20.
- Malik TA, Kamili AN, Chishti MZ, Tanveer S, Ahad S, Johri RK. *In vivo* anticoccidial activity of berberine [18, 5,6-dihydro-9,10-dimethoxybenzo(g)-1,3-benzodioxolo-(5,6-a) quinolizinium]-An isoquinoline alkaloid present in the root bark of *Berberis lycium*. *Phytomedicine*. 2014; 21(5):663-669.
- Mashwani ZUR, Khan MA, Irum S, Ahmad M. Antioxidant potential of root bark of *Berberis lycium* Royle from Galliyat, Western Himalaya, Pakistan. *Pakistan Journal of Botany*. 2013; 45:231-234.
- Sabir S, Tahir K, Rashid N, Naz S, Masood B, Shah AM *et al*. Phytochemical and antioxidant studies of *Berberis lycium*. *Pakistan Journal of Pharmaceutical Sciences*. 2013; 26(6):1165-1172.
- Mushtaq M, Naz S, Khan S, Rehman S, Khan RU. *In vivo* effect of *Berberis lyceum* and *Silybum marianum* on production performance and immune status of broiler chickens. *Archiv Tierzucht*. 2013; 56(91):911-916.
- Chand N, Durrani FR, Ahmad S, Khan A. Immunomodulatory and hepatoprotective role of feed added *Berberis lycium* in broiler chicks. *Journal of the Science of Food and Agriculture*. 2011; 91(10):1737-1745.
- Shah MA, Sherwani SK, Sualeh M, Kanwal S, Khan HN, Kazmi SU. *In vitro* anthelmintic and antiuro lithic assessment of *Berberis lycium* root bark. *Journal of Pharmacognosy and Phytochemistry*. 2014; 3(2):205-210.
- Srivastava P, Srivastava G. Pharmacological and phytochemical screening of *Desmodium gangeticum* and *Moringa oleifera*. *Research Journal of Chemistry and Environment*. 2018; 22(5):6-10.
- Gupta I, Sharma P. Investigation of Phytoconstituents from Leaves, Seeds, Bark and Pods of *Cassia fistula* Plant. *Journal of Chemistry and Chemical Sciences*. 2018; 8(5):886-890.
- Auwal SM, Saka S, Mairiga AI, Sanda AK, Shuaibu A, Ibrahim A. Preliminary Phytochemical and Elemental Analysis of Aqueous and Fractionated Pod Extracts of *Acacia nilotica* (Thorn mimosa). *Veterinary Research Forum*. 2014; 5(2):95-100.
- Kumar A, Patra S. Qualitative and Quantitative Analysis of Secondary Phytochemical in *Gymnema Sylvestre*, *Indian journal of scientific research*. 2017; 12(2):150-156.
- Ismail MA, Mohamed AE, Marghany RM, Abdel-Motaal FF, Abdel-Farid BI, El-Sayed AM. Preliminary Phytochemical Screening, Plant Growth Inhibition and

- Antimicrobial Activity Studies of *Faidherbia albida* legume extract. Journal of the Saudi Society of Agricultural Sciences. 2016; 15:112-117.
28. Miana AG. Tertiary Dihydroprotoberberine Alkaloids of *Berberis Lycium*. Phytochemistry. 1973; 12:1822-1823.
 29. Mukhopadhyay D, Dasgupta P, Roy SD, Palchoudhuri S, Chatterjee I, Ali S *et al.* A Sensitive In vitro Spectrophotometric Hydrogen Peroxide Scavenging Assay using 1,10-Phenanthroline. Free Radicals and Antioxidants. 2016, 6(1).
 30. Alam MN, Bristi NJ, Rafiquzzaman M. Review on in vivo and in vitro methods evaluation of antioxidant activity. Saudi Pharmaceutical Journal. 2013; 21(2):143-152.
 31. Ruch RJ, Cheng SJ, Klaunig JE. Prevention of cytotoxicity and inhibition of intercellular communication by antioxidant catechins isolated from Chinese green tea. Carcinogenesis. 1989; 10(6):1003-1008.
 32. Oyaizu M. Studies on products of browning reactions: antioxidant activities of products of browning reaction prepared from glucosamine. Japan journal of Nutrition. 1986; 44(6):307-315.
 33. Hsieh JT, Chia CY, Wu CY, Chen YC. Chemical Constituents from the Stems of *Mahonia japonica*. Journal of the Chinese Chemical Society. 2004; 51:443-446.
 34. Thakur P, Sarika. Ethno-medical uses of some plants of Potter's hill in Shimla (Himachal Pradesh, India). Biological forum- An International Journal. 2016; 8(2):417-422.