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Exopolysaccharide production and lipolytic activity of promising Lactic acid bacteria isolated from traditional artisan curds of Karnataka for quality yoghurt preparation

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Abstract

As many as 77 lactic acid bacterial strains were isolated from various natural sources and artisan curd samples of Karnataka. They were further subjected for exopolysaccharide (EPS) production and lipolytic activity isolate numbers BC19 and RSC64 produced the highest EPS, 1.65 mm each. Isolate SCR59 produced the highest lipolytic activity of 16.5mm zone of solubilization, diameter. The promising ten isolates were selected and three starter culture consortia were prepared. Based on organoleptic evaluation, starter culture consortia no. I was found to be the best treatment with the overall acceptance of 8.31.

Keywords: Exopolysaccharide, lipolytic activity, lactic acid bacteria, isolates, artisan curds

Introduction

Lactic acid bacteria (LAB) are Gram positive, non-spore forming, Catalase negative, acid tolerant, fastidious, non-motile and facultative anaerobic friendly gut bacteria. They are constitute heterogeneous group of industrially important bacteria. In food industries, they used as preservatives, acidulant and flavouring agents by the virtue of their probiotic traits. They are also used as starter cultures in food fermentations such as beverages, yoghurt, vegetables, cereals, meat, cocoa beans etc. They are also utilized in the development of functional foods and more specifically, their application as vaccines, pro and prebiotics, neutraceuticals has attracted new research arena for food scientists and health professionals (Mozzi *et al.*, 2010 ^[6]; De Vuyst and Leroy, 2004) ^[3].

LAB producing exopolysaccharides (EPS) have received much attention of scientific community in the recent years, due to their useful role in the improvement of physical, rheological and sensory properties of fermented milk (Behare *et al.*, 2013) ^[1]. EPS are long-chain polysaccharides produced extracellularly mainly by bacteria and microalgae. EPS consist of branched, repeating units of sugars or sugar derivatives. These sugar units are mainly glucose, galactose, mannose, N-acetyglucosamine, N-acetyl galactosamine and rhamnose, in variable ratios.

To boost the taste of yoghurt, different fruit pulps can be added (either individually or in combination) which also confer flavor and color to yoghurt to attract the consumers. Such attempts can further augment rheological and sensory attributes of the yoghurt. They are supplemented as probiotics that are live microbial food supplements which beneficially affect the host by improving the intestinal microbial balance. Hence, as many as 77 isolates of lactic acid bacteria were obtained from various sources such as traditional artisan curds, fermented vegetables, Dosa and Idli batter, etc. out of which 10 promising were selected based on EPS production and lipolytic activity.

Thus, selection of efficient LAB strains isolated from diverse natural environments and traditional artisan curds serve as efficient starter cultures for producing yoghurt with improved nutritional, rheological and sensory properties. Hence, in order to obtain productive LAB strains which impart health benefits for exploring their application in developing yoghurt is the objective of the present study.

Materials and methods

Isolation of LAB strains, media and culture conditions

As many as 77 LAB strains were isolated from various sources such as traditional fermented arisan curds of Karnataka such as Kohlar, Banashankari, Almatti areas, fermented vegetables

etc. by the method of serial dilution and plate count (Table.1). The 48 h grown cultures were further purified and maintained on de Mann Rogosa Sharpe broth (MRS) (De Man *et al.*, 1960^[2]) at -80^o C by DMSO (cryoprotectant) method.

EPS production by the isolates was estimated according to the method given by Rimida and Abraham (2003). The isolates were streaked on MRS agar and incubated at 30° C for 48 h. The sticky aspect of the colonies was determined by testing them for slime formation using the inoculation loop method. The mucoidness and ropyness was observed and measured in mm using inoculation loop method. The isolates were considered positively slimy producer if the length of slime was above 1.5 mm.

| Table 1: Sources of the samples used for isolation of lactic acid |
|---|
| bacteria |

| Isolate No | Isolate Code No. | Source of the isolate | | |
|------------|------------------|-----------------------|--|--|
| 1 | KC1 | Kolhar Curd | | |
| 2 | KC2 | Kolhar Curd | | |
| 3 | KC3 | Kolhar Curd | | |
| 4 | KC4 | Kolhar Curd | | |
| 5 | KC5 | Kolhar Curd | | |
| 6 | KC6 | Kolhar Curd | | |
| 7 | KC7 | Kolhar Curd | | |
| 8 | KC8 | Kolhar Curd | | |
| 9 | KC9 | Kolhar Curd | | |
| 10 | KC10 | Kolhar Curd | | |
| 11 | BC11 | Badami Curd | | |
| 12 | BC12 | Badami Curd | | |
| 13 | BC13 | Badami Curd | | |
| 14 | BC14 | Badami Curd | | |
| 15 | BC15 | Badami Curd | | |
| 16 | BC16 | Badami Curd | | |
| 17 | BC17 | Badami Curd | | |
| 18 | BC18 | Badami Curd | | |
| 19 | BC19 | Badami Curd | | |
| 20 | BC20 | Badami Curd | | |
| 21 | BC21 | Badami Curd | | |
| 22 | BC22 | Badami Curd | | |
| 23 | BC23 | Badami Curd | | |
| 24 | BJC24 | Bijjaragi Curd | | |
| 25 | BJC25 | Bijjaragi Curd | | |
| 26 | BJC26 | Bijjaragi Curd | | |
| 27 | BJC27 | Bijjaragi Curd | | |
| 28 | BJC28 | Bijjaragi Curd | | |
| 29 | BJC29 | Bijjaragi Curd | | |
| 30 | BJC30 | Bijjaragi Curd | | |
| 31 | BJC31 | Bijjaragi Curd | | |
| 32 | BJC32 | Bijjaragi Curd | | |
| 33 | BJC33 | Bijjaragi Curd | | |
| 34 | BJC34 | Bijjaragi Curd | | |
| 35 | BJC35 | Bijjaragi Curd | | |
| 36 | BJC36 | Bijjaragi Curd | | |
| 37 | BJC37 | Bijjaragi Curd | | |
| 38 | BJC38 | Bijjaragi Curd | | |
| 39 | BJC39 | Bijjaragi Curd | | |
| 40 | BJC40 | Bijjaragi Curd | | |
| 41 | BJC41 | Bijjaragi Curd | | |
| 42 | BJC42 | Bijjaragi Curd | | |
| 43 | BJC43 | Bijjaragi Curd | | |
| 44 | BJC44 | Bijjaragi Curd | | |
| 45 | BJC45 | Bijjaragi Curd | | |
| 46 | SPG46 | Sprouted Grains | | |
| 47 | SPG47 | Sprouted Grains | | |
| 48 | SPG48 | Sprouted Grains | | |
| 49 | SPG49 | Sprouted Grains | | |
| 50 | SPG50 | Sprouted Grains | | |
| 51 | SPG51 | Sprouted Grains | | |

| 52 | TCW52 | Tender Coconut Water | | |
|----|--------|----------------------------|--|--|
| 53 | TCW53 | Tender Coconut Water | | |
| 54 | TCW54 | Tender Coconut Water | | |
| 55 | TCW55 | Tender Coconut Water | | |
| 56 | TCW56 | Tender Coconut Water | | |
| 57 | SCR57 | Sugarcane Rhizosphere Soil | | |
| 58 | SCR58 | Sugarcane Rhizosphere Soil | | |
| 59 | SCR59 | Sugarcane Rhizosphere Soil | | |
| 60 | CRS60 | Corn Rhizosphere Soil | | |
| 61 | CRS61 | Corn Rhizosphere Soil | | |
| 62 | CRS62 | Corn Rhizosphere Soil | | |
| 63 | CRS63 | Corn Rhizosphere Soil | | |
| 64 | RSC64 | Rhizosphere Soil Chilli | | |
| 65 | RSC65 | Rhizosphere Soil Chilli | | |
| 66 | RSC66 | Rhizosphere Soil Chilli | | |
| 67 | ORS67 | Orchard Rhizosphere Soil | | |
| 68 | ORS68 | Orchard Rhizosphere Soil | | |
| 69 | ORS69 | Orchard Rhizosphere Soil | | |
| 70 | ORS70 | Orchard Rhizosphere Soil | | |
| 71 | CoRS71 | Cocoa Rhizosphere Soil | | |
| 72 | CoRS72 | Cocoa Rhizosphere Soil | | |
| 73 | CoRS73 | Cocoa Rhizosphere Soil | | |
| 74 | AC74 | Almatti Artisan Curd | | |
| 75 | AC75 | Almatti Artisan Curd | | |
| 76 | AC76 | Almatti Artisan Curd | | |
| 77 | AC77 | Almatti Artisan Curd | | |

Lipolytic activity

LAB isolates were tested for their lipolytic activity as described by Katz *et al.* (2002) ^[5].

Starter culture consortia

Based on EPS production and lipolytic activity, 10 efficient LAB isolates were selected and consortia were prepared. The details of the treatments are given below.

Treatment combinations formulated based on potential probiotic and functional characteristics

Treatment Details

- T1: Curd prepared using KMF curd culture
- T2: Yoghurt prepared using Standard reference cultures
- T3: Yoghurt prepared using Starter culture consortium- I BJC41+RSC64+BJC35+BJC42+BJC37I)
- **T4:** Yoghurt prepared using Starter culture consortium- I (BJC41+ RSC64+SPG49+ BJC42+ BJC37)
- **T5:** Yoghurt prepared using Starter culture consortium-III (BJC41+ RSC64+BJC40+BJC37+KC6)

Results and discussion

Exopolysaccharide is an important functional trait of LAB. Exopolysaccharide (EPS) produced by LAB with GRAS (generally regarded as safe) status is an important source of natural alternatives. Recently, EPS produced by LAB have gained considerable attention in the fermented dairy industry because of their potential as viscosifiers, texturizers and emulsifying agents. It has been reported that EPS produced yoghurt starter cultures could affect the texture of yoghurt and improve sensory characters such as mouthful, shinyness, clean, cut, ropyness and creamyness. Hence, the LAB collection was subjected to production of EPS. It was observed that many isolates produced EPS (Table. 2). Isolate BC19 and RSC64 produced the highest amount of EPS of 16.5 each and hold promise to be used in the development of starter consortium.

As many as 50 isolates showed lipolytic activity. The highest activity was observed in isolate No. 69 with a zone of

solubilization of 21.5 mm dia. followed by isolate No. 59 (16.5mm dia.) whereas the least activity was recorded in the isolate No. 9 (6.00 mm dia.).

Based on EPS production and lipolytic activity, finally, 10 LAB isolates were selected. Using these 10 isolates, three starter consortia were made and yogurt prepared and organoleptic evaluation of the different yoghurts was done in comparison with the yogurt prepared using the reference yogurt cultures.

Form Table. 4, out of three starter culture consortia, T3 was found to be the best starter consortium (BJC41+RSC64+BJC35+BJC42+BJC37) which produced a yoghurt with good flavour, acceptable color, appearance and texture with the highest overall acceptability of 8.31.

| Table 2: Production of Exopolysaccharide from lactic acid bacteria |
|--|
| isolates (measured in terms of ropyness, mm) |

| LAB | Production of EPS | | Production of EPS | | |
|----------|-------------------|--------------|-------------------|--|--|
| isolates | (mm) | LAB isolates | (mm) | | |
| KC1 | 0.70 | BJC41 | 1.10 | | |
| KC2 | 0.50 | BJC42 | 1.10 | | |
| KC3 | 0.35 | BJC43 | 0.80 | | |
| KC4 | 0.50 | BJC44 | 0.65 | | |
| KC5 | 0.60 | BJC45 | 0.70 | | |
| KC6 | 1.45 | SPG46 | 0.70 | | |
| KC7 | 0.45 | SPG47 | 0.35 | | |
| KC8 | 0.95 | SPG48 | 0.40 | | |
| KC9 | 0.65 | SPG49 | 0.95 | | |
| KC10 | 0.60 | SPG50 | 1.50 | | |
| BC11 | 0.70 | SPG51 | 0.50 | | |
| BC12 | 0.25 | TCW52 | 0.70 | | |
| BC13 | 0.30 | TCW53 | 0.95 | | |
| BC14 | 0.50 | TCW54 | 0.70 | | |
| BC15 | 0.55 | TCW55 | 0.50 | | |
| BC16 | 0.50 | TCW56 | 0.35 | | |
| BC17 | 0.50 | SCR57 | 0.50 | | |
| BC18 | 0.55 | SCR58 | 0.60 | | |
| BC19 | 1.65 | SCR59 | 0.65 | | |
| BC20 | 0.00 | CRS60 | 0.60 | | |
| BC21 | 0.65 | CRS61 | 1.00 | | |
| BC22 | 0.00 | CRS62 | 1.00 | | |
| BC23 | 0.50 | CRS63 | 0.55 | | |
| BJC24 | 0.30 | RSC64 | 1.65 | | |
| BJC25 | 0.00 | RSC65 | 0.65 | | |
| BJC26 | 0.00 | RSC66 | 0.55 | | |
| BJC27 | 0.00 | ORS67 | 0.50 | | |
| BJC28 | 0.55 | ORS68 | 0.45 | | |
| BJC29 | 0.65 | ORS69 | 0.60 | | |
| BJC30 | 0.55 | ORS70 | 0.35 | | |
| BJC31 | 0.30 | CoRS71 | 0.50 | | |
| BJC32 | 0.50 | CoRS72 | 0.40 | | |
| BJC33 | 0.80 | CoRS73 | 0.35 | | |

| BJC34 | 0.45 | AC74 | 0.45 |
|-------|------|------------|------|
| BJC35 | 1.30 | AC75 | 0.50 |
| BJC36 | 0.30 | AC76 | 0.00 |
| BJC37 | 1.45 | AC77 | 0.35 |
| BJC38 | 0.55 | SEM ± | 0.11 |
| BJC39 | 1.15 | CD @ 0.01% | 0.41 |
| BJC40 | 0.55 | | |

 Table 3: Lipolytic activity of LAB isolates measured in mm (Zone of solubilisation, mm=dia)

| LAB isolates | Lipolytic activity | LAB isolates | Lipolytic activity | | |
|--------------|--------------------|--------------|--------------------|--|--|
| KC1 | 10.50 | BJC41 | 10.00 | | |
| KC2 | 0.00 | BJC42 | 6.50 | | |
| KC3 | 0.00 | BJC43 | 8.50 | | |
| KC4 | 9.00 | BJC44 | 0.00 | | |
| KC5 | 0.00 | BJC45 | 11.50 | | |
| KC6 | 0.00 | SPG46 | 0.00 | | |
| KC7 | 0.00 | SPG47 | 12.50 | | |
| KC8 | 6.50 | SPG48 | 0.00 | | |
| KC9 | 6.00 | SPG49 | 0.00 | | |
| KC10 | 0.00 | SPG50 | 0.00 | | |
| BC11 | 0.00 | SPG51 | 0.00 | | |
| BC12 | 9.00 | TCW52 | 0.00 | | |
| BC13 | 7.00 | TCW53 | 6.50 | | |
| BC14 | 0.00 | TCW54 | 13.00 | | |
| BC15 | 0.00 | TCW55 | 0.00 | | |
| BC16 | 13.50 | TCW56 | 9.00 | | |
| BC17 | 0.00 | SCR57 | 0.00 | | |
| BC18 | 0.00 | SCR58 | 0.00 | | |
| BC19 | 10.00 | SCR59 | 16.50 | | |
| BC20 | 0.00 | CRS60 | 0.00 | | |
| BC21 | 8.50 | CRS61 | 0.00 | | |
| BC22 | 0.00 | CRS62 | 0.00 | | |
| BC23 | 12.00 | CRS63 | 0.00 | | |
| BJC24 | 8.50 | RSC64 | 0.00 | | |
| BJC25 | 0.00 | RSC65 | 0.00 | | |
| BJC26 | 0.00 | RSC66 | 0.00 | | |
| BJC27 | 0.00 | ORS67 | 0.00 | | |
| BJC28 | 0.00 | ORS68 | 0.00 | | |
| BJC29 | 8.50 | ORS69 | 21.50 | | |
| BJC30 | 8.00 | ORS70 | 0.00 | | |
| BJC31 | 7.00 | CoRS71 | 0.00 | | |
| BJC32 | 0.00 | CoRS72 | 0.00 | | |
| BJC33 | 0.00 | CoRS73 | 0.00 | | |
| BJC34 | 8.50 | AC74 | 0.00 | | |
| BJC35 | 0.00 | AC75 | 0.00 | | |
| BJC36 | 10.50 | AC76 | 0.00 | | |
| BJC37 | 9.00 | AC77 | 0.00 | | |
| BJC38 | 0.00 | SEM ± | 0.59 | | |
| BJC39 | 0.00 | CD @ 0.01% | 2.19 | | |
| BJC40 | 0.00 | | | | |

Table 4: Organoleptic evaluation of the yoghurt

| Treatments | | Hedonic scale values | | | | |
|---|------|----------------------|------------|---------|--------------------|--|
| | | Color | Appearance | Texture | Overall acceptance | |
| T1: Curd prepared using KMF curd culture | 6.17 | 6.67 | 7.63 | 7.33 | 7.30 | |
| T2: Yoghurt prepared using Standard Reference cultures | 8.33 | 8.13 | 7.67 | 8.33 | 8.25 | |
| T3: Yoghurt prepared using starter culture consortium-I (BJC41+RSC64+BJC35+BJC42+BJC37) | 7.97 | 8.00 | 7.33 | 7.80 | 8.31 | |
| T4: Yoghurt prepared using starter culture consortium-II (BJC41+ RSC64+SPG49+ BJC42+ BJC37) | 6.33 | 6.00 | 6.67 | 6.00 | 6.50 | |
| T5: Yoghurt prepared using Starter culture consortium-III (BJC41+ RSC64+BJC40+ BJC37+KC6) | 7.00 | 6.67 | 6.33 | 6.67 | 6.73 | |
| S. Em± | 0.28 | 0.34 | 0.42 | 0.39 | 0.34 | |
| CD@0.01 | 1.21 | 1.45 | 1.80 | 1.67 | 1.5 | |

Conclusion

In conclusion, most of the 77 LAB isolates used in the study showed EPS production and lipolytic activity. Using the selected isolates, different consortia were made and the yoghurt prepared by using consortia no. I exhibited the highest quality and the overall acceptance as revealed by organoleptic evaluation. These isolates have a great potential and prospective candidates as one of the probiotic traits besides good overall acceptability to carry forward for further study in the preparation of yoghurt.

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