



E-ISSN: 2278-4136

P-ISSN: 2349-8234

www.phytojournal.com

JPP 2020; 9(2): 287-296

Received: 05-09-2019

Accepted: 05-11-2019

Rishi Raj

Junior Research Fellow,
Department of Entomology,
MBAC, Agwanpur, Saharsa,
Bihar, India

Mrinalini Kumari

Assistant Professor cum Junior
Scientist Zoology, Department of
Entomology, MBAC, Agwanpur,
Saharsa, Bihar, India

Blue green algae (BGA) and its application

Rishi Raj and Mrinalini Kumari

Abstract

Cyanobacteria are known from a long evolutionary history and its unique traits, for example, oxygenic photosynthesis, high biomass yield, growth on water sources mainly contaminated and polluted waters), generation of useful by-products and bio-fuels, in agriculture as biofertilizer, wastewater treatment, enhancing the soil fertility, reclamation of saline soil, and in nutrition as food supplements and reducing greenhouse gas emissions, have collectively offered these bio-agents as the precious bio-resource to harness the maximum benefit of cyanobacteria. Furthermore, Cyanobacteria also produce an ample variety of chemicals (secondary metabolites) that are not needed for their normal growth of cyanobacteria which show powerful biological activities such as strong antiviral, antibacterial, antifungal, antimalarial, antitumoral and anti-inflammatory activities useful for therapeutic purposes. Since cyanobacteria grow in simple growth needs, therefore it is cost-effective to make full use of cyanobacteria for the production of recombinant compounds of medicinal and commercial purposes. Recently cyanobacteria have opened new ways to explore in screening, culture, and genetic engineering techniques without many side effects. In this review examines cyanobacteria uses and its application that can be used to deal with current problems and scarcity such as food, energy, and environmental deterioration.

Keywords: Cyanobacteria, agriculture, biofertilizer, biofuels, greenhouse gas

Introduction

Cyanobacteria (CB), known as blue-green algae (BGA), are a group of gram-negative photosynthetic bacteria [1-6]. That has colonized the earth's surface for nearly 3.5 billion years [7-8] and is considered as the predecessors of the modern-day chloroplast [9-13]. They exist in different morphologies including unicellular and filamentous forms [14-15]. Blue-green algae (BGA) have enormous morphological and metabolic diversity (Stal 1995), which makes BGA special choice in economic development and environment management like atmospheric fixation of nitrogen, land reclamation, reclaim salinity affected soils and wastewater treatment. Cyanobacteria are also used in many biotechnological applications that were hoping for possible uses in, Colorants, mariculture, food, fuel, fertilizers and production of various secondary metabolites including toxins, enzymes, and pharmaceuticals. Cyanobacteria is also rich source of bioactive compounds and considered as most promising groups of an organism to produce them [16-17]. Cyanobacteria bioactive compound include antifungal [18], antibacterial [19], antiviral [20], anticancer [21], antiplasmodial [22], algicide [22], and Immuno suppressive agents [23]. BGA have been also investigated for the production of different biofuels including biohydrogen, biodiesel, bioethanol and biomethane.

Agriculture**Nitrogen fixer**

Nitrogen (N₂) is derived from the nitrogen fixation process and present in amino acids, proteins and DNA [24]. The source of soil nitrogen is the Earth's atmosphere and it is the most abundant gas in (about 79% of the total atmospheric gases.), it is extremely unreactive [25]. Unfortunately, no plant species on the earth that can reduce atmospheric dinitrogen into ammonia and use it directly for its growth. It appears that only a number of prokaryotic microorganisms including cyanobacteria and bacteria have been shown to possess the ability to fix dinitrogen [26-29]. Biological fixation of the atmospheric nitrogen can be about 70% of all nitrogen fixed on the Earth per year, the remaining is by some micro-organisms, autotrophs or heterotrophs 'free' fixers [30]. Bacteria known collectively as the "Rhizobia" are famous for their ability to induce nodules on the roots and sometime on the stems of legume plants [31]. Inside these nodules, the differentiated, "bacteroid" forms fix atmospheric nitrogen and the ammonia being formed and used as a source of fixed nitrogen. In this symbiosis relationship bacteria get niche and, in return, the plants obtain a nitrogen source [32]. They occur in the so-called free-living forms e.g. aerobic azotobacter, anaerobic Clostridia or in symbiosis with certain higher plants e.g. Rhizobia with legumes or Azolla Anabaena. Cyanobacteria can be an

Corresponding Author:**Mrinalini Kumari**

Assistant Professor cum Junior
Scientist Zoology, Department of
Entomology, MBAC, Agwanpur,
Saharsa, Bihar, India

important source of nitrogen for supporting aquatic primary productivity^[33] and important for the global nitrogen cycle^[34]. It has been reported that nitrogen availability to plants is increased due to the application of cyanobacteria in agriculture ecosystems, particularly the rice fields. (Stewart *et al.*, 1968; 35-40).

Biofertilizer

The importance of biofertilizers is increasing rapidly since chemical fertilizers (nitrogenous fertilizers) damage the environment. Biofertilizers (BGA) lead to soil enrichment and are compatible with long-term sustainability. Further, they are eco-friendly and pose no danger to the environment. BGA, are the major N fixers in freshwater and marine systems^[15]. BGA has been assessed for use as green fertilizer in rice fields and positive effects have been reported in India^[16] and other countries^[17-18]. The application of BGA in the rice paddy fields promotes rice growth and yield^[41-42]. Many Asian countries like China, Vietnam, India, etc., have been utilizing cyanobacteria in paddy cultivation as the alternative to nitrogen fertilizers^[43-44].

Fertility of tropical rice field soils was given credit to the activity of diazotrophic cyanobacteria^[45] more or less all dominant CB inhabiting rice fields are diazotrophic and it gives an indication how rice has been grown for centuries without any external supply of N₂^[46] Most rice soils have a natural population of BGA, which provides free of cost, nitrogen. So we can say that BGA provides an ideal environment for their luxuriant growth for plant. According to Agronomist of rice field trial BGA commonly contribute nitrogen in the order of 20-30 kg/ha^[47]. Thus using BGA in the rice field, a farmer can save to the extent of 20-30 kg N/ha without compromising with the normal yield. The beneficial effect of BGA, inoculation has also been reported in other crops (oat, tomato, radish, barley, cotton, sugarcane, maize, chili, and lettuce). Besides the contribution of nitrogen, BGA also liberate growth-promoting substances play an important role in sustaining the crop yield. A number of growth-promoting substances [vitamins (B12), nicotinic acid, pantothenic acid, folic acid amino acids, sugars, polysaccharides, etc.), including hormone] have been documented^[48-49].

Soil conditioner

BGA also has huge potential in environmental management as a soil conditioner. Organic changes not only act by improving soil structure but also strongly influence soil microflora^[50]. It had been seen that a continuous layer of colonies of different CB species in rice fields commonly yield a significant amount of biomass^[50]. BGAs excrete a number of compounds (polysaccharides, peptides, lipids during growth in soil^[51-53], which assumed to be diffuse through soil particles and hold them together as macroaggregates. Moreover, polysaccharides can also implicate clay particles and form clusters. Then these clusters or macroaggregates, in turn, grow and take the shape of macroaggregates and subsequently of larger soil aggregates. The interwoven nature of algal filaments may also help in binding soil particles along with organic carbon added through algal biomass. A significant increase has been observed in soil aggregate stability due to an increase in polysaccharide content of the soil as a result of algal inoculation^[54-55].

Reclaim Salinity Affected Soils

BGA also holds the potential to reclaim salinity affected soils.

In the 1950s, a biological approach to the problem of saline soils using BGA was proposed, where in natural populations of nitrogen fixing BGA were employed for reclamation of saline/alkaline lands typical of certain North Indian States.^[56-57] Potential of a brackish-water, BGA, *Anabaena torulosa*, to grow and enrich N status of moderately saline coastal (Kharland) soils have been demonstrated. Most of the sodium removed by BGA remains extracellularly trapped in their mucopolysaccharide sheaths^[58]. Rogers and Burns (1994) CB is also known to solubilize and mobilize phosphorus (P) and make it available to plants. N fixing BGA, *Tolypothrix tenuis*, N fixing CB, *Tolypothrix tenuis*, *Hapalosiphon fontinalis* solubilize Mussoorie rock phosphate (MRP), a source of P₂O₅. Similarly, *Westiellopsis prolifica* and *A. variabilis* showed good growth and N fixing ability in the presence of mineral phosphate sources (MRP and TCP) in addition to solubilizing these insoluble phosphate sources^[59-60].

Food

BGA is being used as a food for a human being a long time^[61-65]. Nostoc is a more common dietary supplement for the native populations of Thailand, Peru, China, Ecuador, Fiji, Java, Japan, Mexico, Mongolia, and Siberia are taking different forms of Nostoc like *N. commune* var. *flagelliform*, *N. edule*, *N. ellipso sporum*, *N. verrucosum* and *N. pruniforme*^[66] *N. commune* has a high amount of fiber and protein can play an important physiological and nutritional role in the human diet. Nostoc commune is rich in fibres and *Aphanizomenon* sp. is collected from natural blooms in Lake Klamath (Oregon, The USA) to be used as healthy food^[67].

Spirulina-cyanobacteria have been eaten for centuries by Kanembu people, who live along the shores of Lake Chad in northcentral Africa. Quantities of proteins and other compounds in spirulina are, varies between 50% and 70% of its dry weight. These levels are quite exceptional, even among microorganisms. It has the highest protein of any natural food (65%); far more than animal and fish flesh (15-25%), soybeans (35%), dried milk (35%), peanuts (25%), eggs (12%), grains (8-14%) or whole milk (3%). Moreover, the best sources of vegetable protein achieve only half these levels; for example, you can compare to soya flour contains "only" 35% crude protein. However, the protein content of spirulina varies by 10-15% according to the time of harvesting in relation to daylight^[68]. The highest values were obtained at early daylight. Due to its high protein content and various possible health-promoting effects (alleviation of hyperlipidemia, suppression of hypertension, protection against renal failure, growth promotion of intestinal lactobacilli and lowering of elevated serum glucose level), *Spirulina* has been exploited commercially and recommended as a protein supplement^[69-73]. They can act as the nutritional supplement or represent a source of natural food colorants^[74-81].

Spirulina also contains a considerable amount of carbohydrate (~20%), lipid (~5%), and mineral (~7%). It is a very rich source of β -carotene, thiamine, riboflavin and is one of the richest sources of vitamin B12. (82-83). It also contains 20 antioxidants, 68 minerals, and 70 trace elements, all amino acids, B vitamins, and important enzymes^[84-94].

Hainan Simai Enterprising located in the Hainan province of China is the largest producer of *Spirulina* in the world. On the other hand, Earthrise Company (Caliptera, CA, USA) has the largest *Spirulina* production plant and their products are distributed in over 20 countries. Besides, some other companies selling BGA Commercial preparations like

“Zyulina” and “Recolina” (Zydu Cadila), “Nuclina” (Mapra Laboratories), “Vitalina” containing dry powder of Spirulina is available in the Indian market. Capsules containing dry powder of *Aphanizomenon flos-aquae*, under the trade name of Kalmath’s Best® Blue-Green Algae by Kalmath Valley Botanicals LLC, USA have gained popularity in the USA, Japan, Germany, Canada, Korea, and Austria^[95]

Feed for animals and aquaculture

Spirulina is now mainly used as feed supplements for various organisms and more than half of the total population of spirulina used as feed^[96]. There is no side effect, for e.g. when soy protein in feed for pigs was replaced by spirulina there wasn’t any adverse effect seen apparently on the gastrointestinal. Breeders use spirulina as a tool to change coloration, accelerate growth and attaining earlier sexual maturity & improve fertility rates in parrots, lovebirds & canary, finch. It is also used by breeders to increase reproduction rates in ostrich and turkey. Moreover, the intake of spirulina can increase desirable yellow skin color in chicken & also enhance the deep yellow color of egg yolks^[97]. The digestive system of birds was also enhanced with increased good bacteria population, feather color & its shine was also increased altogether. Spirulina is also being used as feed for bivalve mollusks, penacide shrimp larvae, brine shrimps and marine rotifers etc. spirulina content when increased up to 10% in brine shrimp feed, it increased their survival rates allowing the wing fish to reach market size sooner^[97]. It is now considered as the best food for tiny brine shrimps. Those BGA, who fix nitrogen in marine also used as fish food, therefore, marine BGA used in aquaculture. The Tilapia fish showed high growth rates when fed with marine cyanobacteria in indoor and outdoor cultures^[98]. In India, *Phormidium valderianum* has been used as a complete aquaculture feed source based on its nontoxic nature and nutritional value.

Species of various BGA like *Nostoc*, *Anabaena* and *Calothrix* increase body weight of Telapia hybrid fish with excellent food conversion ratio. As a feed, 5-20% of Spirulina enhanced the color pattern of Champion Koi fish, thereby being fascinating and attractive for a customer, therefore, a farmer can increase its benefit and value in the market by several folds^[97].

Biotechnology

Cyanobacteria have gained a lot of attention in recent years because of their potential applications in biotechnology. Some cyanobacteria found which accumulates polyhydroxyalkanoates (PHA)^[99] which are similar in properties to polyethylene and polypropylene^[100]. These biodegradable plastics could replace by oil derived thermoplastics in some fields. Recent research also shows that cyanobacteria form ideal association with Chemotrophic bacteria can effectively clean oil-contaminated sediments and waste waters^[101-105].

Enzymes

A number of studies also confirmed the detection of enzymes produced by BGA (nitrogenase, hydrogenase, phosphatase, β -Lactamase, elastase, hydroperoxide lyase, arylsulfatase, chitinase, L-asparaginase, L-glutaminase, amylase, protease, lipase, cellulase, urease and lactamase)^[106-110]. An anti-fertility enzyme, elastase, has been purified from *Oscillatoria* sp. Several unique sequence-specific restriction

endonucleases are produced by BGA, and its potential use exploited in recombinant DNA technology.

Biofuel

BGA have been investigated for the production of different biofuels including biohydrogen, biodiesel, bioethanol and biomethane.

Biohydrogen

Hydrogen gas is seen as a future energy source^[111], since it is renewable, does not emit the “greenhouse gas” CO₂ in combustion, liberates large amounts of energy per unit weight in combustion^[112-114]. The advantage of using Hydrogen gas, if used as a fuel, will not cause environmental pollution because its only by-product is water and it is one of the most abundant elements in the universe, and has maximum energy per unit weight (122 KJ g⁻¹). On a weight basis, it is calculated that the heating value of H₂ is 141.65 MJ Kg⁻¹, which is the highest amongst known fuels^[115]. More over it can be stored as gas-metal hydride or as liquid, and has greater energy conversion efficiency than petroleum. Several Cyanobacteria genera including *Anabaena*, *Calothrix*, *Oscillatoria*, *Cyanothece*, *Nostoc*, *Synechococcus*, *Microcystis*, *Gloeobacter*, *Aphanocapsa*, *Chroococcidiopsis*, and *Microcoleus* are known for their ability to produce H₂ under various culture conditions^[116-118]. BGA, can produce different feed-stock for energy generation like H₂ by photosynthesis, lipids for biodiesel, jet fuel and hydrocarbons, isoprenoids for gasoline and carbohydrates for ethanol production^[119-120]. Biological hydrogen production has several benefits over hydrogen production by thermochemical or photoelectrochemical processes. In cyanobacteria, two natural pathways for H₂ production can be used: first, H₂-production as a by-product during nitrogen fixation by nitrogenases; and second, H₂-production directly by bidirectional hydrogenase^[121]. Nitrogenases require ATP whereas bidirectional hydrogenases do not require ATP for H₂-production, hence making them more efficient and favorable for H₂-production with a much higher turnover. *Cyanothece* sp. ATCC 51142, a unicellular, diazotrophic cyanobacterium with capacity to generate high levels of hydrogen under aerobic conditions. Wild-type *Cyanothece* sp. 51142 can produce hydrogen at rates as high as 465lmol/mg of chlorophyll/h in the presence of glycerol. Hydrogen production in this strain is mediated by an efficient nitrogenase system, which can be manipulated to convert solar energy into hydrogen at rates that are several fold higher, compared to other previously described wild-type hydrogen-producing photosynthetic microbes^[122].

Many unique features of cyanobacteria among them, generation of useful by-products and bio-fuels, enhancing the soil fertility and reducing greenhouse gas emissions, have collectively make Cyanobacteria as the precious bio-resource for sustainable development. Now a days genetically engineered cyanobacteria have been devised with the novel genes for the production of a number of bio-fuels such as bio-diesel, bio-hydrogen, bio-methane, synga, and therefore, open new avenues for the generation of bio-fuels in the economically sustainable manner^[123].

Reduction of global warming

CO₂ sequestration

The CO₂ sequestration by BGA is receiving increased awareness in the current situation of global warming^[124].

Being photosynthetic, contribute a large share of the total photosynthetic conversion of solar energy and assimilation of CO₂. The CO₂ fixation rate in cyanobacteria is about 10-50 times faster than the terrestrial plants. So the use of BGA, can be considered one of the effective method to reduce the concentration of atmospheric CO₂ and thereby, to help in reduction global warming [125]. It is predicted that half of global photosynthesis is fixed by phytoplankton, among them mostly includes Cyanobacteria members [126]. If we consider more specific, about 25% of the total global photosynthesis can be accounted for by only two efficient marine Cyanobacteria genera, *Synechococcus* and *Prochlorococcus* [127]. Cyanobacteria CO₂ fixation in photobioreactors has recently gained more attention for reduction of CO₂ mitigation, because its efficiency increased to many fold. A number of studies have been conducted during the past few decades related to this strategy for CO₂ sequestration [128-131].

Reduction of methane

Methane (CH₄) has negative effect, as it is a potent Green House Gas (GHG) with approximately 20 times more impact of CO₂ [132] cyanobacteria may possibly minimize the emissions of CH₄ from flooded rice soils at the levels of production, transport, and consumption. Bioagents like methanotrophs [133] can play a very significant role to remove significant amount of the most potent and dangerous GHGs such as CH₄ from the soils of various ecosystems in association with cyanobacteria [134-136].

Antibiotics and other potential drugs

Medicinal merit of BGA was first recognized as early as 1500 BC when *Nostoc* species were used to treat gout, fistula and several forms of cancer [137-138] cyanobacteria are a rich source of potentially useful natural products [139-141] And produce a number of bioactive compounds of potential therapeutic use including fatty acids, having activities such as anti-HIV, anticancer, antifungal, antimalarial and antimicrobial [142-149]

Antibacterial activity

Noscomin, a diterpenoid from *N. commune*, showed antibacterial activity against *Bacillus cereus*, *Staphylococcus epidermidis*, and *Escherichia coli* [150] *L. majuscula* shows antibacterial activity against *B. cereus*, *B. subtilis*, *Mycobacterium balnei*, etc. [151]. Norharmane, produced from *Nodularia harveyana* have anti Cyanobacteria activity against other cyanobacteria [152] thus, could be useful in controlling harmful Cyanobacteria blooms.

Fungicidal activity

Natural products of *Nostoc* sp. are effective against *Cryptococcus* sp. as a causal agent of secondary fungal infections in patients with AIDS [153]. Majusculamide-C, a microfilament depolymerizing agent from *L. majuscula*, has shown potent fungicidal activity and may find application in the treatment of resistant fungal-induced diseases of plants and agricultural crops. *Fischerella ambigua* and *Haplosiphon hybernicus* produce Ambiguines, which show anti-fungal activity on *Aspergillus oryzae* and *Candida albicans* [154]. Tjipanazoles produced from *Tolypothrix tjipanasensis* have antifungal activities, especially against fungi responsible for wheat leaf rust rice blast and [155].

Antiviral activity

Cyanovirin-N analogs and Cyanothecae show antiviral activity against a large panel of viruses. The first one, cyanovirin-N

analogs, have been isolated from *Nostoc ellipsosporum* [156] and *Cyanotheca* sp. [157] Cyanovirin show inhibitory activity against HIV-1, HIV-2, simian immunodeficiency virus (SIV), feline immunodeficiency virus, HHV-6, and measles virus (156-157). Also, they inhibit Ebola and influenza viruses. Lipophilic and hydrophilic are strains of cultured CB were examined and they have the ability to inhibit enzyme Reverse Transcriptase (RT) of Avian Myeloblastosis Virus (AMV) and HIV type [156-157-158] Fibre [oxalate oxalic acid-soluble substances (OOSS)], extracted from *N. commune*, and possesses a hypocholesterolemic effect in rats.

Antimalarial activity

Symplocamide A obtained from a BGA, *Symploca* sp. was found to be active against parasite of malaria (*P. falciparum* W2), Chagas disease (*Trypanosoma cruzi*) and leishmaniasis (*Leishmania donovani*) [159-163].

An anti-cancer

An anti-cancer factor has been identified in *Phormidium tenue*, *Scytonema* sp. and *Anabaena variabilis*.

Curacin A, isolated from *L. majuscula* an antimitotic agent (an antiproliferative agent), and its function exploited in inhibiting growth of colon, renal and breast cancer derived cell lines [164-167].

Cryptophycin-1, isolated from a *Nostoc* sp. GSV 224 [165-167], its function had been identified as mitotic inhibitor by blocking the cell cycle at the G₂/M phase via inhibition of tubulin polymerization [168-169].

Nostoc linkia produce borophycine: Can be another anticancer compound (boron containing polyketide) [170]

Tolypothrix nodosa produce Tolyporphin: Its function is to photo sensitising activity against tumour cells and it is 5,000-times more effective than the photodynamic treatment [171].

Calotoxin isolated from *Scytonema ocellatum* depolymerized actin to disturb cell division and seems to be a potent anti-cancer drug.

Anti-inflammatory

BGA also have Anti-inflammatory property, Consumption of BGA has been demonstrated to promote immunity and to protect against inflammatory diseases, such as colitis, arthritis, and allergic rhinitis in animal and human studies [172-175].

Hormone

CB produce Auxins [176-178], Gibberellins [179-180], Cytokinins [181-182] abscisic acids [183]. Ethylene [184]

Acknowledgement

We are thankful to SERB, New Delhi for providing the financial support.

Reference

1. Ferris MJ, Muyzer G, Ward DM. Denaturing gradient gel electrophoresis profiles of 16S rRNA-defined populations inhabiting a hot spring microbial mat community. *Appl Environ Microbiol.* 1996; 62:340-346.
2. Ward DM, Santegoeds CM, Nold SC, Ramsing NB, Ferris MJ, Bateson MM. Biodiversity within hot spring microbial mat communities: molecular monitoring of enrichment cultures. *Antonie Van Leeuwenhoek.* 1997; 71:143-150.

3. Nu'bel U, Garcia-Pichel F, Ku'hl M, Muyzer G. Quantifying microbial diversity: morphotypes, 16S rRNA genes, and carotenoids of oxygenic phototrophs in microbial mats. *Appl Environ Microbiol.* 1999; 65:422-430.
4. Nu'bel U, Garcia-Pichel F, Clavero E, Muyzer G. Matching molecular diversity and ecophysiology of benthic cyanobacteria and diatoms in communities along a salinity gradient. *Environ Microbiol.* 2000; 2:217-226.
5. Abed RMM, Garcia-Pichel F. Long-term compositional changes after transplant in a microbial mat Cyanobacteria community composition revealed using a polyphasic approach. *Environ Microbiol.* 2001; 3:53-62.
6. Garcia-Pichel F, Pringault O. Cyanobacteria track water in desert soils. *Nature.* 2001; 413:380-381.
7. Cockell CS. Ultraviolet radiation, evolution and the π -electron system. *Biol J Linn Soc.* 1998; 63:449-457. [Google Scholar]
8. Hedges SB, Chen H, Kumar S, Wang DY, Thompson AS, Watanabe H. A genomic timescale for the origin of eukaryotes. *BMC Evolutionary Biology.* 1, article. 2001; 4:10. Doi: 10.1186/1471-2148-1-4. [PMC free article][PubMed] [CrossRef][Google Scholar]
9. Tamagnini P, Leitao E, Oliveira P, Ferriera D, Pinto F, Harris DJ *et al.* Cyanobacteria hydrogenases: diversity, regulation and applications. *FEMS Microbiol. Rev.* 2007; 31:692-720.
10. Löffelhardt W, Bohnert HJ. Molecular biology of cyanelles. In: Bryant D A, editor. *The molecular biology of cyanobacteria.* Dordrecht, the Netherlands: Kluwer, 1994, 65-89. [Google Scholar]
11. Löffelhardt W, Bohnert HJ, Bryant DA. The cyanelles of *Cyanophora paradoxa*. *Crit Rev Plant Sci.* 1997; 16:393-413. [Google Scholar]
12. De PK. The role of blue green algae in nitrogen fixation in rice fields, *Proc Roy Soc London,* 127 B (1939) 121-139.
13. Castenholz RW. Phylum BX. Cyanobacteria. Oxygenic photosynthetic bacteria. In *Bergey's Manual of Systematic Bacteriology. The Archaea and the Deeply Branching and Phototropic Bacteria* ed. Garrity G, Boone, 2001, 1.
14. DR, Castenholz RW. New York: Springer-Verlag Stal LJ. *Physiological ecology of cyanobacteria in microbial mats and other communities.* *New Phytol.* 1995; 131:1-32, 474-487.
15. Bhadury P, Wright PC. Exploitation of marine algae: biogenic compounds for potential antifouling applications. *Planta.* 2004; 219:561-578.
16. Dahms HU, Xu Y, Pfeiffer C. Antifouling potential of cyanobacteria: a mini-review. *Biofouling.* 2006; 22:317-327.
17. Kajiyama S, Kanazaki H, Kawazu K, Kobayashi A. Nostifungicidine, an antifungal lipopeptide from the field-grown terrestrial blue-green algae *Nostoc commune*. *Tetrahedron Lett.* 1998; 39:3737-3740.
18. Jaki B, Heilmann J, Sticher O. New antibacterial metabolites from the cyanobacterium *Nostoc commune* (EAWAG 122b). *J Nat Prod.* 2000; 63:1283-1285.
19. Patterson GML, Larsen LK, Moore RE. Bioactive natural products from blue-green algae. *J Appl Phycology.* 1994; 6:151-157.
20. Gerwick WH, Roberts MA, Proteau PJ, Chen JL. Screening cultured marine microalgae for anticancer-type activity. *J Appl Phycol.* 1994; 6:143-149.
21. Papendorf O, Ko'nig GM, Wright AD. Hirridin B and 2,4-dimethoxy-6 heptadecylphenol, secondary metabolites from the cyanobacterium *Phormidium ectocarpi* with antiplasmodial activity. *Phytochem.* 1998; 49:2383-2386.
22. Koehn FE, Lomgley RE, Reed JK. Microcolins A and B, new immunosuppressive peptide from the bluegreen algae *Lyngbya majuscula*. *J Nat Prod.* 1992; 55:613-619.
23. Egamberdieva D, Kucharova Z. Cropping effects on microbial population and nitrogenase activity in saline arid soil. *Turk. J. Biol.* 2008; 32:85-90.
24. Frank IB, Lundgren P, Falkowski P. Nitrogen fixation and photosynthetic oxygen evolution in cyanobacteria. *Research in Microbiol.* 2003; 154:157-164.
25. Capone DG, Burns JA, Montoya JP, Subramaniam A, Mahaffey AC, Gunderson T *et al.* Nitrogen fixation by *Trichodesmium* spp.: an important source of new nitrogen to the tropical and subtropical North Atlantic Ocean. *Global Biogeochem Cycles* 19: GB2024, 2005. Doi: 10.1029/2004GB002331.
26. Wagner, Andreas Otto *et al.* Effects of different nitrogen sources on the biogas production-a lab-scale investigation. *Microbiological research.* 2012; 167(10):630-636.
27. Di Rienzi, Sara C *et al.* The human gut and groundwater harbor non-photosynthetic bacteria belonging to a new candidate phylum sibling to Cyanobacteria. *Elife.* 2013; 2:e01102.
28. Nghia NH, Gyurjan. Problems and perspectives in establishment of nitrogen-fixing symbioses and endosymbiosis. *Endocyt. C. Res.* 1987; 4:131-141.
29. Peter VM, Cassman K, Cleveland C, Crews T, Christopher BF, Grimm BN *et al.* Towards an ecological understanding of biological nitrogen fixation. *Biogeochemistry.* 2002; 57:1-45.
30. Gonzalez LJ, Rodelas B, Pozo C, Salmeron V, Mart2nez MV, Salmeron V. The liberation of amino acids by heterotrophic nitrogen-fixing bacteria. *Amino Acids.* 2005; 28:363-367.
31. Andrew JW, Jonathan D, Andrew R, Lei S, Katsaridou NN, Mikhail S *et al.* Living without Fur: the subtlety and complexity of iron-responsive gene regulation in the symbiotic bacterium *Rhizobium* and other α -proteobacteria. *Biometals.* 2007; 20:501-511.
32. Rodrigo V, Novelo E. Seasonal changes in periphyton nitrogen fixation in a protected tropical wetland. *Biol. Fertil Soils.* 2007; 43:367-372.
33. Stewart WDP, Fitzgerald GP, Burns RH. Acetylene reduction by nitrogen-fixing blue-green algae. *Arch. Microbiol.* 1968; 62:336-348.
34. Affourtit J, Zehr JP, Paerl HW. Distribution of nitrogen-fixing microorganisms along the Neuse river estuary. *North Carolina Microb. Ecol.* 2001; 41:114-123.
35. Peters GA, Toia REJ, Lough SM. The *Azolla-Anabaena* relationship. V.15 N₂ fixation, acetylene reduction and H₂ production. *Plant Physiol.* 1977; 59:1021-1025. Doi: 10.1104/pp.59.6.1021
36. Singh AL, Singh PK. Influence of *Azolla* management on the growth, yield of rice and soil fertility. II. N and P contents of plants and soil. *Plant Soil.* 1987; 102:49-54.
37. Shridhar BS. Review: Nitrogen fixing microorganisms. *Int J Microbiol Res.* 2012; 3:46-52.
38. Pabby A, Prasanna R, Singh PK. *Azolla Anabaena* symbiosis-From traditional agriculture to biotechnology. *Indian J Biotechnol.* 2003; 2:26-37.
39. Reddy PM, Roger PA, Ventura W, Watanabe I. Blue-green algal treatment and inoculation had no significant effect on rice yield in an acidic wetland soil. *Philippine Agr Sci.* 1986; 69:629-32.

40. Asuming-Brempong S. Sustainable growth of rice in Ghana: The role of biofertilizers (phosphate solubilizing microorganisms and *Azolla Anabaena*) to rice improvement. *Global Adv Res J Agr.* 2014; *Sci* 3:1-7.
41. Ghosh TK, Saha KC. Effects of inoculation with N₂ fixing cyanobacteria on the nitrogenase activity in soil and rhizosphere of wet land rice (*Oryza sativa* L.). *Biol Fertil Soils.* 1993; *16*:16-20.
42. Begum ZNT, Mandal R, Islam S. Effect of Cyanobacteria biofertilizer on the growth and yield components of two HYV of rice. *J Algal Biomass Util.* 2011; *2*:1-9.
43. Venkataraman GS. *Algal Biofertilizers and Rice Cultivation.* New Delhi: Today and tomorrow Printers and Publishers, 1972.
44. Lumpkin TA, Plucknett DL. *Azolla as a Green Manure: Use and Management in Crop Production: Westview Tropical Agriculture Series No. 5.* Boulder, Colo: Westview Press, 1982, 230.
45. De PK. The role of blue green algae in nitrogen fixation in rice fields, *Proc Roy Soc London.* 1939; *127B*:121-139.
46. Watanabe I, Yoneyama T, Padre B, Ladha JK. The difference in the natural abundance of ¹⁵N in several rice (*Oryza sativa* L.) varieties: Application for evaluating N₂ fixation, *Soil Sci Plant Nutr.* 1987; *33*:407-415.
47. Goyal SK. Algal biofertilizer for vital soil and free nitrogen, *Proc Ind Natl Sci Acad.* 1993; *B59*:295-302.
48. Misra S, Kaushik BD. Growth promoting substances of cyanobacteria: Detection of amino acids, sugars and auxins, *Proc Ind Natl Sci Acad.* 1989; *B55*:499-504.
49. Kaushik BD. Blue green algal (Cyanobacteria) biofertilizer and nutrient management in rice crop, in *Soil-Plant-Microbe Interaction in Relation to Integrated Nutrient Management*, edited by B D Kaushik (Venus Printers and Publishers, New Delhi), 1998, 55-63.
50. Crecchio C, Curci M, Mininni R, Ricciuti P, Ruggiero P. Short term effects of municipal solid waste compost amendments on soil carbon and nitrogen content, some enzyme activities and genetic diversity, *Biol Fertil Soils.* 2001; *34*:311-318.
51. Marathe KV. Role of some BGA in soil aggregation, in *Taxonomy and Biology of BGA*, edited by T V Desikachary, (Madras University Press, Madras, India), 1972, 328-331.
52. Mehta VG, Vaidya BS. Cellular and extracellular polysaccharides of the blue-green alga *Nostoc*, *J Exp Bot.* 1978; *113*:1423-1430.
53. Bertocchi C, Navarini L, Cesaro A, Anastasio M. Polysaccharides from cyanobacteria, *Carbohydr Polym.* 1990; *12*:127-153.
54. Rao DLN, Burns RG. Use of blue-green algae and bryophyte biomass as a source of nitrogen for oil-seed rape, *Biol Fertil Soils.* 1990; *10*:61-64.
55. Rogers SL, Burns RG. Changes in aggregate stability, nutrient status, indigenous microbial populations and seedling emergence, following inoculation of soil with *Nostoc muscorum*, *Biol Fertil Soils.* 1994; *18*:209-215.
56. Singh RN. Reclamation of "usar" lands in India through blue green algae, *Nature.* 1950; *165*:325-326.
57. Singh RN. Reclamation of usar lands, in *Role of Blue Green Algae in Nitrogen Economy of Indian Agriculture* (Indian Council of Agricultural Research, New Delhi, 1961, 83-98.
58. Apte SK, Thomas J. Possible amelioration of coastal soil salinity using halotolerant nitrogen-fixing cyanobacteria, *Plant Soil.* 1997; *189*:205-211.
59. Yandigeri MS, Yadav AK, Meena, KK, Pabbi S. Effect of mineral phosphate on growth and nitrogen fixation of diazotrophic cyanobacteria *Anabaena variabilis* and *Westiellopsis prolifica*, *Antonie Leeuwenhoek.* 2010; *97*:297-306.
60. Yandigeri MS, Pabbi S. Response of diazotrophic Cyanobacteria to alternative sources of phosphorus, *Ind J Microbiol.* 2005; *45*:132-134.
61. Radwan SS, Al-Hasan RH. "Oil pollution and cyanobacteria," in *The Ecology of Cyanobacteria*, eds B. A. Whitton and M. Potts (Berlin: Kluwer Academic Publishers), 2000, 307-319.
62. Carmichael WW, Gorham PR. *Freshwater Cyanophyte Toxins.* Algal Biomass, 1980, 437-448. New York: Elsevier.
63. Chakdar, Hillol *et al.* "Potential applications of blue green algae", 2012.
64. Singh S, Kate BN, Banerjee UC. Bioactive compounds from cyanobacteria and microalgae: An overview. *Crit. Rev. Biotechnol.* 2005; *25*:73-95. [Cross Ref] [PubMed]
65. Spolaore P, Joannis-Cassan C, Duran E, Isambert A. Commercial applications of microalgae. *J Biosci. Bioeng.* 2006; *101*:87-96. [Cross Ref] [PubMed]
66. Chisti Y. Microalgae as sustainable cell factories. *Environ. Eng. Manag. J.* 2006; *5*:261-274.
67. Milledge JJ. Commercial application of microalgae other than as biofuels: A brief review. *Rev. Environ. Sci. Biotechnol.* 2011; *10*:31-41. [Cross Ref]
68. Jassby A. *Spirulina: A model microalgae as human food*, in *Algae and Human affairs*, edited by Lembi CA, Waaland JP (Cambridge University Press, Cambridge), 1988, 149-179.
69. Watanabe F, Katsura H, Takenaka S *et al.*, "Pseudovitamin B12 is the predominant cobamide of an algal health food, spirulina tablets," *Journal of Agricultural and Food Chemistry.* 1999; *47*(11):4736-4741.
70. Singh Jay Shankar *et al.* "Cyanobacteria: a precious bio-resource in agriculture, ecosystem, and environmental sustainability". *Frontiers in microbiology.* 2016; *7*:529.
71. Brown MR, Jeffrey SW, Volkman JK, Dunstan GA. Nutritional properties of microalgae for mariculture. *Aquaculture.* 1997; *151*:315-331. 10.1016/S0044-8486(96)01501-3 [Cross Ref] [Google Scholar]
72. Bandaranayake WM. Mycosporines: are they nature's sunscreens? *Nat. Prod. Rep.* 1998; *15*:159-172. 10.1039/a815159y [PubMed] [CrossRef] [Google Scholar]
73. Sinha RP, Klisch M, Groniger A, Hader DP. Ultraviolet-absorbing/ screening substances in cyanobacteria, phytoplankton and macroalgae. *J. Photochem. Photobiol.* 1998; *47*:83-94. 10.1016/S1010-6030(98)00332-3
74. Kulshreshtha A, Zacharia J, Jarouliya U, Bhadauriya P, Prasad GBKS, Bisen PS. *Spirulina in healthcare management.* *Curr. Pharm. Biotechnol.* 2008; *9*:400-405. 10.2174/138920108785915111
75. Nelis HJ, DeLeenheer AP. Microbial sources of carotenoid pigments used in foods and feeds. *J. Appl. Bacteriol.* 1991; *70*:181-191. 10.1111/j.1365-2672.1991.tb02922.x [CrossRef] [Google Scholar]
76. Borowitzka MA. Commercial production of microalgae: ponds, tanks, tubes and fermenters. *J Biotechnol.* 1999; *70*:313-321. 10.1016/S0168-1656(99)00083-8 [CrossRef] [Google Scholar]

77. Muller-Feuga A. The role of microalgae in aquaculture: situation and trends. *J Appl. Phycol.* 2000; 12:527-534. 10.1023/A:1008106304417 [Cross Ref] [Google Scholar]
78. Branen LA, Davidson MP, Salmine NS, Thorngate HJ. *Food Additives*. New York, NY: Marcel Dekker, 2002. [Google Scholar]
79. Becker W. "Microalgae in human and animal nutrition," in *Handbook of Microalgal Culture*, ed. Richmond A., editor. (Oxford: Blackwell), 2004, 312-351. [Google Scholar]
80. Rangel-Yagui CO, Godoy Danesi ED, Carvalho JCM, Sato S. Chlorophyll production from *Spirulina platensis*: cultivation with urea addition by fed-batch process. *Bioresour. Technol.* 2004; 92:133-141. 10.1016/j.biortech.2003.09.002 [PubMed] [CrossRef] [Google Scholar]
81. Bhaskar SU, Gopalswamy G, Raghu R. A simple method for efficient extraction and purification of C-phycoerythrin from *Spirulina platensis* Geitler. *Indian J. Exp. Biol.* 2005; 43:277-279. [PubMed] [Google Scholar]
82. Soletto D, Binaghi L, Lodi A, Carvalho JCM, Converti A. Batch and fed-batch cultivations of *Spirulina platensis* using ammonium sulphate and urea as nitrogen sources. *Aquaculture.* 2005; 243:217-224. 10.1016/j.aquaculture.2004.10.005 [CrossRef] [Google Scholar]
83. Plavsic M, Terzic S, Ahel M, Van Den Berg CMG. Folic acid in coastal waters of the Adriatic Sea. *Mar. Freshw. Res.* 2004; 53:1245-1252. 10.1071/MF02044 [CrossRef] [Google Scholar]
84. Prasanna R, Sood A, Jaiswal P, Nayak S, Gupta V, Chaudhary V *et al.* Rediscovering Cyanobacteria as valuable sources of bioactive compounds (Review). *Appl. Biochem. Microbiol.* 2010; 46:119-134. 10.1134/S0003683810020018 [PubMed] [CrossRef] [Google Scholar]
85. Nakamura H, Kobayashi J, Hirata Y. Separation of mycosporine-like amino acids in marine organisms using reversed phase high performance liquid chromatography. *J Chromatogr.* 1982; 250:113-118. 10.1016/S0021-9673(00)95219-1
86. Garcia-Pichel F, Wingard CE, Castenholz RW. Evidence regarding the UV sunscreen role of a mycosporine-like compound in the cyanobacterium *Gloeocapsa* sp. *Appl. Environ. Microbiol.* 1993; 59:170-176. [PMC free article] [PubMed] [Google Scholar]
87. Bohm GA, Pfliegerer W, Boger P, Scherer S. Structure of a novel oligosaccharide-mycosporine-aminoacid ultraviolet A/B sunscreen pigment from the terrestrial cyanobacterium *Nostoc commune*. *J Biol. Chem.* 1995; 270:8536-8539. 10.1074/jbc.270.15.8536 [PubMed] [CrossRef] [Google Scholar]
88. Rimbau V, Camins A, Pubill D, Sureda FX, Romay C, Gonzalez R *et al.* C-phycoerythrin protects cerebellar granule cells from low potassium/serum deprivation-induced apoptosis. *Arch. Pharmacol.* 2001; 364:96-104. 10.1007/s002100100437 [PubMed] [CrossRef] [Google Scholar]
89. Sinha RP, Klisch M, Helbling EW, Hader DP. Induction of mycosporine-like amino acids (MAA's) in cyanobacteria by ultraviolet B radiation. *J. Photochem. Photobiol.* 2001; 60:129-135. 10.1016/S1011-1344(01)00137-3 [PubMed] [CrossRef] [Google Scholar]
90. Kedar L, Kashman Y, Oren A. Mycosporine-2-glycine is the major mycosporine-like amino acid in a unicellular cyanobacterium (*Euhalothece* sp.) isolated from a gypsum crust in a hypersaline saltern pond. *FEMS Microbiol. Lett.* 2002; 208:233-237. 10.1111/j.1574-6968.2002.tb11087.x [PubMed] [CrossRef] [Google Scholar]
91. Rissanen T, Voutilainen S, Nyyssonen K, Salonen JT. Lycopene atherosclerosis, and coronary heart disease. *Exp. Biol. Med.* 2002; 227:900-907. [PubMed] [Google Scholar]
92. Romay CP, Gonazalez R, Ledon N, Ramirez D, Vimbaré V. C-phycoerythrin: a biliprotein with antioxidant, anti-inflammatory and neuroprotective effects. *Curr. Protein Peptide Sci.* 2003; 4:207-216. 10.2174/1389203033487216 [PubMed] [CrossRef] [Google Scholar]
93. Benedetti S, Benvenuti F, Pagliarani S, Francogli S, Scoglio S, Canestrari F. Antioxidant properties of a novel phycoerythrin extract from the blue green alga *Aphanizomenon flos-aquae*. *Life Sci.* 2004; 75:2353-2362. 10.1016/j.lfs.2004.06.004 [PubMed] [CrossRef] [Google Scholar]
94. Rajeev KJ, Xu Z. Biomedical compounds from marine organisms. *Mar. Drugs.* 2004; 2:123-146. 10.3390/md203123 [CrossRef] [Google Scholar]
95. Subhashini J, Mahipal SV, Reddy MC, Mallikarjuna Reddy M, Rachamalla A, Reddanna P. Molecular mechanisms in C-phycoerythrin induced apoptosis in human chronic myeloid leukemia cell line-K, 2004.
96. Hillol Chakdar, Shrikrishna D, Jadhav, Dolly Wattal Dhar, Sunil Pabbi. Centre for Conservation and Utilisation of Blue Green Algae, Indian Agricultural Research Institute, New Delhi 110 012.
97. Yamaguchi K. Recent advances in microalgal bioscience in Japan with special reference to utilization of biomass and metabolites: A review, *J Appl Phycology.* 1997; 8:487-502.
98. Official web page: <http://www.spirulinasource.com>, 2009.
99. Thajuddin N, Subramanian G. "Cyanobacteria biodiversity and potential applications in biotechnology". *Current science* 4, 2005, 7-57.
100. Vincenzini, Massimo *et al.* "Occurrence of poly-beta-hydroxybutyrate in *Spirulina* species". *Journal of bacteriology.* 1990; 172.5:2791-2792.
101. Steinbüchel A, Fuchtenbusch B, Gorenflo V, Hein S, Jossek R, Langenbach S, Rehm BHA. Biosynthesis of polyesters in bacteria and recombinant organisms. *Polymer Degrad Stabil.* 1997; 59:177-182.
102. Yan GA, Jiang JW, Wu G, Yan X. Disappearance of linear alkylbenzene sulfonate from different cultures with *Anabaena* sp. HB 1017. *Bulletin of Environmental Contamination and Toxicology.* 1998; 60:329-334.
103. Radwan SS, Al-Hasan RH. Oil pollution and cyanobacteria. In: Whitton, B.A., Potts, M. (Eds.). *The Ecology of Cyanobacteria*. Kluwer, the Netherlands, 2000, 307-319.
104. Raghukumar C, Vipparthy V, David JJ, Chandramohan D.. Degradation of crude oil by marine cyanobacteria. *Applied Microbiology and Biotechnology.* 2001; 57:433-436.
105. Mansy AE, El-Bestway E. Toxicity and biodegradation of fluometuron by selected Cyanobacteria species. *World Journal of Microbiology and Biotechnology.* 2002; 18:125-131.
106. Abed RMM, Koester J. The direct role of aerobic heterotrophic bacteria associated with cyanobacteria in

- the degradation of oil compounds. *Int Biodeterior Biodegrad.* 2005; 55:29-37.
107. Bergman B *et al.* "N₂ fixation by non-heterocyst cyanobacteria". *FEMS Microbiology reviews.* 1997; 19.3:139-185.
 108. Lindblad P, Tamagnini P. "Cyanobacteria hydrogenases and biohydrogen: Present status and future potential". *Biohydrogen II.* Pergamon, 2001, 143-169.
 109. Sarma TA, Swaran Kanta, Usha Kiran. "AMYLASE ACTIVITY IN A BLUE-GREEN ALGA, 1977.
 110. Kushner DJ, Colette Breuil. "Penicillinase (β -lactamase) formation by blue-green algae". *Archives of microbiology.* 1977; 112.2:219-223.
 111. Beneytout, Jean-Louis *et al.* "Properties of a lipoxxygenase in green algae (*Oscillatoria* sp.)". *Plant physiology.* 1989; 91.1:367-372.
 112. Dutta D, De D, Chaudhuri S, Bhattacharya SK. Hydrogen production by cyanobacteria. *Microb cell Fact.* 2005; 4:36 Doi:10.1186/1475-2859-4-36
 113. Tamagnini P, Leit-ao E, Oliveira P, Ferreira D, Pinto F, Harris DJ *et al.* Cyanobacteria hydrogenases: diversity, regulation and applications. *FEMS Microbiology Review.* 2007; 31(6):692e720.
 114. Levin DB, Lawrence P, Murry L. Biohydrogen production: prospects and limitations to practical application. *Int. J Hydrogen Energy.* 2004; 29:173-185.
 115. Sakurai H, Masukawa H. Promoting R&D in photobiological hydrogen production utilizing mariculture-raised cyanobacteria. *Mar. Biotechnol.* 2007; 2:128-145.
 116. Ali I, Basit MA. Significance of hydrogen content in fuel combustion. *Inter. J Hydrogen Energ.* 1993; 18:1009-1011. Doi: 10.1016/0360-3199(93)90083-M
 117. Masukawa H, Nakamura K, Mochimaru M, Sakurai H. "Photobiological hydrogen production and nitrogenase activity in some heterocystous cyanobacteria," in *Biohydrogen II*, eds J Miyake T, Matsunaga, San Pietro A (Oxford: Elsevier Science Ltd), 2001, 63-66.
 118. Parmar A, Singh NK, Pandey A, Gnansounou E, Madamwar D. Cyanobacteria and microalgae: a positive prospect for biofuels. *Bioresour. Technol.* 2011; 102:10163-10172. Doi: 10.1016/j.biortech.2011.08.030
 119. Nozzi NE, Oliver JWK, Atsumi S. Cyanobacteria as a platform for biofuel production. *Front. Bioeng. Biotechnol.* 2013; 1:7. doi:10.3389/fbioe.2013.00007
 120. Parmar A, Singh NK, Pandey A, Gnansounou E, Madamwar D. Cyanobacteria and microalgae: a positive prospect for biofuels. *Bioresour. Technol.* 2011; 102:10163-10172. Doi: 10.1016/j.biortech.2011.08.030
 121. Rosgaard L, De Porcellinis AJ, Jacobsen JH, Frigaard NU, Sakuragia Y. Bioengineering of carbon fixation, biofuels, and biochemicals in cyanobacteria and plants. *J. Biotechnol.* 2012; 162:134-147. Doi:10.1016/j.jbiotec.2012.05.006
 122. Angermayr SA, Hellingwer KJ, Lindblad P, Teixeira De Mattos MJ. Energy biotechnology with cyanobacteria. *Curr. Opin. Biotechnol.* 2009; 20:257-263.
 123. Bandyopadhyay A, Stöckel J, Min H, Sherman LA, Pakrasi HB. High rates of photobiological H₂ production by a cyanobacterium under aerobic conditions. *Nat. Commun.* 2010; 1:139
 124. Kumar A, Singh JS. "Microalgae and cyanobacteria biofuels: a sustainable alternate to crop-based fuels," in *Microbes and Environmental Management*, eds Singh JS, Singh DP (New Delhi: Studium Press Pvt. Ltd.), 2016, 1-20.
 125. Kumar K, Dasgupta CN, Nayak B, Lindblad P, Das D. Development of suitable photobioreactors for CO₂ sequestration addressing global warming using green algae and cyanobacteria. *Bioresour. Technol.* 2011; 102:4945-4953. Doi: 10.1016/j.biortech.2011.01.054
 126. *Biotechnol. Adv.* 25, 294-306. Doi:10.1016/j.biotechadv.2007.02.001
 127. Fuhrman J. Genome sequences from the sea. *Nature.* 2003; 424:1001-1002. doi: 10.1038/4241001a
 128. Rohwer F, Thurber RV. Viruses manipulate the marine environment. *Nature.* 2009; 459:207-212. Doi: 10.1038/nature08060
 129. Hanagata N, Takeuchi T, Fukuju Y, Barnes DJ, Karube I. Tolerance of microalgae to high CO₂ and high temperature. *Photochemistry.* 1992; 31:3345-3348. Doi: 10.1016/0031-9422(92)83682-O
 130. Maeda K, Owada M, Kimura N, Omata K, Karub I. CO₂ fixation from the flue gas on coal-fired thermal power plant by microalgae. *Energ. Convers. Manage.* 1995; 36:717-720. Doi:10.1016/0196-8904(95)00105-
 131. Mirata S, Hayashitani M, Taya M, Tone S. Carbon dioxide fixation in batch culture of *Chlorella* sp. using a photobioreactor with a sunlight-collection device. *J. Ferment. Bioeng.* 1996a; 81:470-472. Doi: 10.1016/0922-338X(96)85151-8
 132. Hirata S, Taya M, Tone S. Characterisation of *Chlorella* cell cultures in batch and continuous operations under a photoautotrophic condition. *J Chem. Eng. Jpn.* 1996b; 29:953-959. Doi: 10.1252/jcej.29.953
 133. Rangel-Yagui CO, Godoy Danesi ED, Carvalho JCM, Sato S. Chlorophyll production from *Spirulina platensis*: cultivation with urea addition by fed-batch process. *Bioresour. Technol.* 2004; 92:133-141. Doi:10.1016/j.biortech.2003.09.002
 134. Singh JS. Methanotrophs: the potential biological sink to mitigate the global methane load. *Curr. Sci.* 2011; 100:29-30
 135. Tiwari S, Singh JS, Singh DP. Methanotrophs and CH₄ sink: effect of human activity and ecological perturbations. *Climate Change Environ. Sustain.* 2015; 3:35-50. Doi: 10.5958/2320-642X.2015.00004.6
 136. Singh JS. Anticipated effects of climate change on methanotrophic methane oxidation. *Climate Change Environ. Sustain.* 2013a; 1:20-24. Doi:10.5958/j.2320-6411.1.1.003
 137. Singh JS, Pandey VC. Fly ash application in nutrient poor agriculture soils: impact on methanotrophs density dynamics and paddy yields. *Ecotoxicol. Environ. Safe.* 2013; 89:43-51. doi: 10.1016/j.ecoenv.2012.11.011
 138. Singh JS, Singh DP. Impact of anthropogenic disturbances on methanotrophs abundance in dry tropical forest ecosystems, India. *Expert Opin. Environ. Biol.* 2013a; 2:1-3.
 139. Singh, Jay Shankar *et al.* "Cyanobacteria: a precious bio-resource in agriculture, ecosystem, and environmental sustainability". *Frontiers in microbiology.* 2016; 7:529.
 140. Parmar, Asha *et al.* "Cyanobacteria and microalgae: a positive prospect for biofuels". *Bioresource technology.* 2011; 102(22):10163-10172.
 141. Pietra F. *A Secret World: Natural Products of Marine Life* (first ed), Birkhäuser, Basel, Switzerland, 1990. Google Scholar

142. Namikoshi M, Rinehart KL. "Bioactive compounds produced by cyanobacteria". *Journal of Industrial Microbiology*. 1996; 17(5-6):373-384.
143. Chen, Lanzhou, Dunhai Li, Yongding Liu. "Salt tolerance of *Microcoleus vaginatus* Gom. a cyanobacterium isolated from desert algal crust, was enhanced by exogenous carbohydrates". *Journal of arid environments*. 2003; 55(4):645-656.
144. Burja, Adam M *et al.* "Marine cyanobacteria-a prolific source of natural products". *Tetrahedron*. 2001; 57(46):9347-9377.
145. Donia, Marwa, Mark T. Hamann. "Marine natural products and their potential applications as anti-infective agents". *The Lancet infectious diseases*. 2003; 3(6):338-348.
146. Skulberg, Olav M. "Microalgae as a source of bioactive molecules-experience from cyanophyte research". *Journal of Applied Phycology*. 2000; 12(3-5):341-348.
147. Newman DJ, Cragg GM, Snader KM. Natural products as sources of new drugs over the period 1981-2002. *J Nat Prod*. 2003; 66:1022-1037.
148. Newman DJ, Cragg GM. Marine natural products and related compounds in clinical and advanced preclinical trials. *J Nat Prod*. 2004a; 67:1216-1238.
149. Tan LT. Bioactive natural products from marine cyanobacteria for drug discovery. *Phytochemistry*. 2007; 68:954-979.
150. Gademan K, Portman C. Secondary metabolites from cyanobacteria: complex structures and powerful bioactivities. *Curr Org Chem*. 2008; 12:326-341.
151. Jones AC, Gu L, Sorrels CM, Sherman DH, Gerwick WH. New tricks from ancient algae: natural products biosynthesis in marine cyanobacteria. *Curr Opin Chem Biol*. 2009; 13:216-223.
152. Tan LT. Filamentous tropical marine cyanobacteria: a rich source of natural products for anticancer drug discovery. *J Dembitsky VM, Rezanka T. "Metabolites produced by nitrogen-fixing Nostoc species". Folia microbiologica*. 2005; 50(5):363-391.
153. Moikeha SN, Chu GW. Dermatitis-producing alga *Lyngbya majuscula* Gomont in Hawaii. II. Biological properties of the toxin factor, *J Phycol*. 1971; 7:8-13.
154. Volk, Rainer-B, Franz H, Furkert. "Antialgal, antibacterial and antifungal activity of two metabolites produced and excreted by cyanobacteria during growth". *Microbiological Research*. 2006; 161(2):180-186.
155. Kuwaki, Shinsuke *et al.* "Antifungal activity of the fermentation product of herbs by lactic acid bacteria against tinea". *Journal of bioscience and bioengineering*. 2002; 94(5):401-405.
156. Bonjouklian, Rosanne *et al.* "Tjipanazoles, new antifungal agents from the blue-green alga *Tolypothrix tjipanasensis*". *Tetrahedron*. 1991; 47(37):7739-7750.
157. Smitka, Tim A *et al.* "Ambiguine isonitriles, fungicidal hapalindole-type alkaloids from three genera of blue-green algae belonging to the Stigonemataceae". *The Journal of Organic Chemistry*. 1992; 57(3):857-861.
158. Boyd, Michael R *et al.* "Discovery of cyanovirin-N, a novel human immunodeficiency virus-inactivating protein that binds viral surface envelope glycoprotein gp120: potential applications to microbicide development". *Antimicrobial agents and chemotherapy*. 1997; 41(7):1521-1530.
159. Dey, Barna *et al.* "Multiple antiviral activities of cyanovirin-N: blocking of human immunodeficiency virus type 1 gp120 interaction with CD4 and coreceptor and inhibition of diverse enveloped viruses". *Journal of virology*. 2000; 74(10):4562-4569.
160. Matei, Elena *et al.* "Structure and glycan binding of a new cyanovirin-N homolog". *Journal of Biological Chemistry*. 2016; 291(36):18967-18976.
161. Pettit GR. The Dolastatins. *Prog. Chem. Org. Nat. Prod*. 1997; 70:2-79.
162. Fennell BJ *et al.* "Effects of the antimitotic natural product dolastatin 10, and related peptides, on the human malarial parasite *Plasmodium falciparum*". *Journal of Antimicrobial Chemotherapy*. 2003; 51(4):833-841.
163. Lehr F, Posten C. Closed photo-bioreactors as tools for biofuel production. *Curr Opin Biotechnol*. 2009; 20:280-285.
164. McPhail KL, Correa J, Linington RG, Gonzalez J, Ortega-Barría E, Capson TL, Gerwick WH. Antimalarial linear lipopeptides from a Panamanian strain of the marine cyanobacterium *Lyngbya majuscula*. *J Nat Prod*. 2007; 70:984-988.
165. Barbaras D, Kaiser M, Brunb R, Gademann K. Potent and selective antiplasmodial activity of the Cyanobacteria alkaloid nostocarboline and its dimers. *Bioorg Med Chem Lett*. 2008; 18:4413-4415.
166. Linington RG, Edwards DJ, McPhail SCF, KL MT, Gerwick WH. Symplocamide A, a potent cytotoxin and chymotrypsin inhibitor from the marine cyanobacterium *Symploca* sp. *J Nat Prod*. 2008; 71:22-27.
167. Gerwick WH, Proteau PJ, Nagle DG, Hamel E, Blokhin A, Slate DL. Structure of Curacin A, a novel antimitotic, antiproliferative and brine shrimp toxic natural product from the marine cyanobacterium *Lyngbya majuscula*. *J Org Chem*. 1994; 59:1243-1245.
168. White JD, Kim T-S, Nambu M. Absolute configuration and total synthesis of (+)-curacin A, an antiproliferative agent from the cyanobacterium *Lyngbya majuscula*. *J Am Chem Soc*. 1997; 119:103-111.
169. Verdier-Pinard P, Sitachitta N, Rossi JV, Sackett DL, Gerwick WH, Hamel E. Biosynthesis of radiolabeled curacin A and its rapid and apparently irreversible binding to the colchicine site of tubulin. *Arch Biochem Biophys*. 1999; 370:51-58.
170. Jones AC, Gu L, Sorrels CM, Sherman DH, Gerwick WH. New tricks from ancient algae: natural products biosynthesis in marine cyanobacteria. *Curr Opin Chem Biol*. 2009; 13:216-223.
171. Schwartz RE, Hirsch CF, Sesin DF *et al.* Pharmaceuticals from cultured algae. *J Ind Microbiol*. 1990; 5:113-124.
172. Trimurtulu G, Ogino J, Helzel CE *et al.* Structure determination confirmational analysis chemical stability studies and anti-tumor evaluation of the cryptophycins: Isolation of 18 new analogues from *Nostoc* sp. strain GSV 224. *J Am Chem Soc*. 1995; 117:12030-12049.
173. Barrow RA, Hemscheidt T, Liang J, Paik S, Moore RE, Tius MA. Total synthesis of cryptophycins. Revision of the structures of cryptophycins A and C. *J Am Chem Soc*. 1995; 117:2479-2490.
174. Hemscheidt T, Puglisi MP, Larsen LK, Patterson GML, Moore RE, Rios JL *et al.* Structure and biosynthesis of borophycin, a new boeseken complex of boric acid from a marine strain of the blue-green alga *Nostoc linckia*. *J Org Chem*. 1995; 59:3467-3471.
175. Morliere P, Maziere JC, Santus R *et al.* Tolyporphin: a natural product from cyanobacteria with potent

- photosensitizing activity against tumor cells *in vitro* and *in vivo*. *Cancer Res.* 1998; 58:3571-3578.
176. Selmi, Carlo *et al.* The effects of Spirulina on anemia and immune function in senior citizens. *Cellular & molecular immunology.* 2011; 8(3):248.
177. Coskun ZK *et al.* The study of biochemical and histopathological effects of spirulina in rats with TNBS-induced colitis. *Batıslavskie lekarske listy.* 2011; 112(5):235-243.
178. Rasool, Mahaboobkhan, Evan Prince Sabina, and Balaji Lavanya. Anti-inflammatory effect of Spirulina fusiformis on adjuvant-induced arthritis in mice. *Biological and Pharmaceutical Bulletin.* 2006; 29(12):2483-2487.
179. Ahmad, Mirza Rashid, Alan Winter. Studies on the hormonal relationships of algae in pure culture. *Planta.* 1968; 78(3):277-286.
180. Misra S, Kaushik BD. "Growth promoting substances of cyanobacteria II. Detection of amino acids, sugars and auxins". *Proc. Indian Sci. Acad. B,* 1989, 55.
181. Kumar, Gagan *et al.* "Secondary Metabolites from Cyanobacteria: A Potential Source for Plant Growth Promotion and Disease Management". *Secondary Metabolites of Plant Growth Promoting Rhizomicroorganisms.* Springer, Singapore, 2019, 239-252.
182. Sergeeva, Elena, Anton Liaimer, Birgitta Bergman. Evidence for production of the phytohormone indole-3-acetic acid by cyanobacteria. *Planta.* 2002; 215(2):229-238.
183. Singh VP, Trehan T. Effects of extracellular products of *Aulosira fertilissima* on the growth of rice seedlings. *Plant Soil.* 2002; 38:457-464. 10.1007/BF00779027
184. Mohan M, Mukherji KG. Some biologically active extracellular products of blue-green algae. *Phykos.* 1978; 18:73-82.
185. Rodgers GA, Bergman B, Henriksson E, Udriş M. Utilization of blue-green algae as bio-fertilizers. *Plant Soil.* 1979; 52:99-107. 10.1007/BF02197736
186. Selykh IO, Semenova LR. *Problems of Ecology and Physiology of Microorganisms.* Moscow: Dialog-MGU, 2000, 94.
187. Huang, Tan-Chi, Te-Jin Chow. Ethylene production by blue-green algae. *Bot. Bull. Academia Sinica.* 1984; 25:81-86.
188. Chakdar, Singh, Jay Shankar *et al.* Cyanobacteria: a precious bio-resource in agriculture, ecosystem, and environmental sustainability. *Frontiers in microbiology.* 2016; 7:529.