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Ravishankar Lanjhiyana

Assistant Professor, Department of Fruit Science, Indira Gandhi Agricultural University, Raipur, Chhattisgarh, India

GD Sahu

Assistant Professor, Department of Fruit Science, Indira Gandhi Agricultural University, Raipur, Chhattisgarh, India

HK Panigrahi

Assistant Professor, Department of Fruit Science, Indira Gandhi Agricultural University, Raipur, Chhattisgarh, India

Pratibha Katiyar

Professor, Department of Plant Physiology, Agri. Bio-chemistry Medicinal Aromatic Plants, Indira Gandhi Agricultural University, Raipur, Chhattisgarh, India

Corresponding Author:**Ravishankar Lanjhiyana**

Assistant Professor, Department of Fruit Science, Indira Gandhi Agricultural University, Raipur, Chhattisgarh, India

Role of pre-sowing seed treatment on germination behavior and seedling vigour of papaya (*Carica papaya* L.)

Ravishankar Lanjhiyana, GD Sahu, HK Panigrahi and Pratibha Katiyar

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Abstract

An experiment entitled “Role of pre-sowing seed treatment on germination behavior and seedling vigour of papaya (*Carica papaya* L.)” was conducted at Instructional cum Research Farm, College of Agriculture and Research Station, (IGKV), Bemetara, Chhattisgarh, India during the year 2018 and 2019. The experiment was laid out in Completely Randomized Design (CRD) with Seventeen different treatments replicated three times. The papaya seeds were sown in polybags which was filled with common media and treated with three different concentration of GA₃ (viz., 50, 100 and 150) and 10 per cent cow urine with addition of bio-fertilizers in media soil. The results revealed that the application of treatment T₁₁ (seed soaking with 150 ppm GA₃ + 12 hours + azotobacter) was given early germination, rate of emergence as well as highest germination percentage, seed vigour index – I and seed vigour index – II. While, T₁₅ (seed soaking with cow urine 10 (%) + 12 hours + azotobacter) had given maximum root: shoot ratio.

Keywords: GA₃, papaya, *Azotobacter*, cow urine, phosphate solubilizing bacteria

Introduction

Papaya (*Carica papaya* L.) is an important fruit crop of the tropical and subtropical regions of the world, in the genus *carica* belonging to the *caricaceae* family and chromosome number is 2n = 18. It is originated from tropics of America (Hofmeyr, 1938) [16]. In India, papaya is being cultivated an area of about 138.4 thousand hectares having annual production of 5989.88 thousand metric tonnes with productivity of 43.30 mt/ha. Andhra Pradesh is leading state in papaya production followed by Gujarat, Karnataka, Madhya Pradesh, Maharashtra and Chhattisgarh. In Chhattisgarh, papaya is cultivated in an area of about 14.40 thousand hectares with production of 381.42 thousand metric tonnes and productivity is 26.48 mt/ha. In Chhattisgarh, top five major papaya producing districts are Durg, Mahasamund, Raipur, Bilaspur and Bemetara (Anon., 2018) [3].

Papaya is a very delicious, nutritious, refreshing fruit and highly valued for its digestive properties. 100g of papaya contains (89.6%) moisture, (9.5%) carbohydrate, (4.0%) calorific value, (0.5%) proteins, (0.1%) fat, (0.4%) minerals, (0.01% calcium, 0.01% phosphorus, 0.4mg iron), 2020 IU Vitamin A (Carotene), 40 IU Vitamin B (Thiamine) 250 IU Vitamin B₂ (riboflavin), 85 mg Vitamin C (ascorbic acid), and 0.2 IU nicotinic acid (Ram, 2005) [5].

Papaya cultivation is restricting some areas in the country due to several limitations on its cultivation. Under present situation, propagation problems, different sex-forms, water logging, frost susceptibility and fungal and viral diseases are well known problems in its cultivation of all over India. Papaya plants react best with integrated management. Because, commercially propagation of papaya is seed and it is highly cross pollinated crop, variability is very much possible. Several internal and external factors such as oxygen, temperature, plant species, water, growth regulators and radiation effect etc. affect the consistency of the plants.

Phosphorus solubilizing bacteria *Pseudomonas spp.* and Nitrogen fixing bacteria *Azotobacter spp.* is known to produce many growth hormones, which often increased root and shoot growth. The useful effect of bio-fertilizers is now well established in fruit crops like banana (Gogoi *et al.*, 2004) [14] and papaya (Sukhade *et al.*, 1995) [28]. Still, very small work has been done on the use of bio-fertilizers in papaya.

For successful seedling production of papaya appropriate seed germination and seedling growth are most imperative consideration under nursery technique. As the germination rate and seedling growth are affected by different pre-sowing seed treatment of papaya. A wide variety of pre-sowing seed treatment, which includes the plant growth regulators and growing media are used to increase the seed germination and seedling growth in several fruit crops.

Papaya seed declines viability rapidly with aging (Begum *et al.*, 1988). The gelatinous sarcotesta (outer seed coat) enclosed with seed which is formed from the outer integument, which can inhibit germination. The papaya seed viability decreases when the sarcotesta remains intact and dry seeds are more prominently affected than fresh ones. GA₃ 200 ppm used as seeds soaking for 12 hours reduced the time taken for 50 per cent germination (Anburani and Shakila, 2010)^[1].

Materials and Methods

The study was carried out during the year 2018 and 2019 at Instructional cum Research Farm, College of Agriculture and Research Station, (IGKV), Bemetara, Chhattisgarh, India. The experiment was laid out in Completely Randomized Design (CRD) with three replications. The treatments included T₁: Untreated (control), T₂: Seed soaking with 50 ppm GA₃ + 06 hours + Azotobacter, T₃: Seed soaking with 50 ppm GA₃ + 12 hours + Azotobacter, T₄: Seed soaking with 50 ppm GA₃ + 06 hours + Phosphate solubilizing bacteria, T₅: Seed soaking with 50 ppm GA₃ + 12 hours + Phosphate solubilizing bacteria, T₆: Seed soaking with 100 ppm GA₃ + 06 hours + Azotobacter, T₇: Seed soaking with 100 ppm GA₃ + 12 hours + Azotobacter, T₈: Seed soaking with 100 ppm GA₃ + 06 hours + Phosphate solubilizing bacteria, T₉: Seed soaking with 100 ppm GA₃ + 12 hours + Phosphate solubilizing bacteria, T₁₀: Seed soaking with 150 ppm GA₃ + 06 hours + Azotobacter, T₁₁: Seed soaking with 150 ppm GA₃ + 12 hours + Azotobacter, T₁₂: Seed soaking with 150 ppm GA₃ + 06 hours + Phosphate solubilizing bacteria, T₁₃: Seed soaking with 150 ppm GA₃ + 12 hours + Phosphate solubilizing bacteria, T₁₄: Seed soaking with Cow urine 10 (%) + 06 hours + Azotobacter, T₁₅: Seed soaking with Cow urine 10 (%) + 12 hours + Azotobacter, T₁₆: Seed soaking with Cow urine 10 (%) + 06 hours + Phosphate solubilizing bacteria and T₁₇: Seed soaking with Cow urine 10 (%) + 12 hours + Phosphate solubilizing bacteria. Observations were recorded in respect to first germination from the date of sowing up to germination of the first seedling, germination percentage at 30 days after sowing by counting number of papaya seeds germinated out of total seed dibbled. Seed vigour index – I, seed vigour index – II and root: shoot ratio at 60 days after sowing. The data generated from these investigations were appropriately computed, tabulated and analyzed as described by Panse and Sukhatme, 1985)^[23] and OPSTAT in Completely Randomized Design (CRD).

Results and Discussion

Effect of pre-sowing seed treatment on germination parameters

On the basis of pooled data (Table 1), gibberellic acid significantly affects the various germination parameters of papaya seeds which is clearly evident from the experiment. In pooled data, Minimum days required for germination 6.83 days was recorded in the treatment T₁₁ (seed soaking with 150 ppm GA₃ + 12 hours + Azotobacter) followed by 7.17 days in treatment T₁₀ (seed soaking with 150 ppm GA₃ + 06 hours + Azotobacter) and 7.39 days in the treatment T₁₃ (seed soaking with 150 ppm GA₃ + 12 hours + PSB). Gibberellic acid acts on the embryo and causes synthesis of hydrolyzing enzymes particularly amylase and protease and this hydrolyzed food is utilized for growth of embryo and thereby enhanced the germination (Paleg, 1965)^[21]. Similar findings were reported by Suryakanth *et al.* (2005)^[29] in guava; Dhankar and Singh (1996)^[11] and Gholap *et al.* (2000)^[13] in aonla; Barche *et al.*

(2010)^[5] and Dhinesh Babu *et al.* (2010)^[12], Anjanwe *et al.* (2013)^[2] and Amit Desai *et al.* (2017)^[9] in papaya. This finding is similar to that of Vasu *et al.* (2010)^[31] reported that 10 g inoculation of *Azotobacter* showed highest germination and also reduced the average time taken to start germination.

On the basis of pooled data (Table 1), the maximum rate of emergence 79.38 (%) was recorded in the treatment T₁₁ (seed soaking with 150 ppm GA₃ + 12 hours + Azotobacter) followed by 77.38 (%) in treatment T₁₀ (seed soaking with 150 ppm GA₃ + 06 hours + Azotobacter) and 74.45 in the treatment T₁₃ (seed soaking with 150 ppm GA₃ + 12 hours + PSB). The promising effect of gibberellic acid on seed germination might be due to its participation in the activity of alpha-amylase, which catalyzes the starch conversion in to simple carbohydrates and chemical energy is liberated which is used in the activation of embryo (Anjanawe *et al.*, 2013)^[2]. The results are in conformity with the findings of Babu *et al.* (2010)^[4, 7], Singh *et al.* (2002)^[27], Kadam *et al.* (1992)^[17], Palanisamy *et al.* (1987)^[20].

Similar trend was also found during the year 2018 and 2019 for rate of emergence and on the basis of pooled data (Table 1), the maximum germination percentage 94.17 (%) was recorded in the treatment T₁₁ (seed soaking with 150 ppm GA₃ + 12 hours + Azotobacter) followed by 90.83 (%) in treatment T₁₀ (seed soaking with 150 ppm GA₃ + 06 hours + Azotobacter) and 88.33 in the treatment T₁₃ (seed soaking with 150 ppm GA₃ + 12 hours + PSB). The maximum germination percentage was recorded when seeds soaked in gibberellic acid might be due to fact that GA₃ involved in the activation of cytological enzymes with GA₃ stimulates seed germination of α – amylase enzymes which convert insoluble starch into soluble sugars and it also initiates the radical growth by removing some metabolic blocks. This might be due to fact that gibberellic acid play an important role in leaching out of the inhibitors into the soaking medium and breaking seed dormancy. This result are conformity with Deb *et al.* (2010)^[10], Babu *et al.* (2010)^[4, 7] Barche *et al.* (2010)^[5], and Anjanwe *et al.* (2013)^[2] in papaya and Amit Desai *et al.* (2017)^[9] in papaya. This finding is also supported by Sinish *et al.* (2005)^[26] who reported that inoculation of *Azotobacter* in the potting mixture induced better germination percentage.

Effect of pre-sowing seed treatment on seedling vigour of papaya

On the basis of pooled data (Table 1), at 60 days after sowing of papaya seeds, the maximum shoot/root ratio of papaya 1.07 was recorded in the treatment T₁₅ (seed soaking with cow urine 10 (%) + 12 hours + Azotobacter) followed by 1.05 in treatment T₁₄ (seed soaking with cow urine 10 (%) + 06 hours + Azotobacter) and 1.05 in the treatment T₃ (seed soaking with 50 ppm GA₃ + 12 hours + Azotobacter).

It might be due to the cow urine was maintaining of high water content in cell, increased cell division and cell elongation which had increased the overall growth of seedlings and thereby increased root:shoot ratio. These results were conformity with the findings of Suthesh *et al.* (2016)^[30] in sandal wood and Amit Desai *et al.* (2017)^[9] in papaya.

On the basis of pooled data (Table 2), at 60 days after sowing of papaya seeds, the maximum seedling vigour index-I of the papaya sapling 3939.83 was recorded in the treatment T₁₁ (seed soaking with 150 ppm GA₃ + 12 hours + Azotobacter) followed by 3755.45 in treatment T₁₀ (seed soaking with 150 ppm GA₃ + 06 hours + Azotobacter) and 3008.44 in the

treatment T₇ (seed soaking with 100 ppm GA₃ + 12 hours + Azotobacter).

In present study, the seed vigour index-I of seedling due to GA₃ pre-soaking can be correlated with higher seed germination, higher shoot length and root length and number of leaves has lead to over all assimilation and distribution of food material with the plant (Brain and Hemming,1955)^[8] and hence resulted in higher seedling vigour (Pampanna and Sulikeri, 2001)^[22]. The results were in close agreement with the Pampanna and Sulikeri (2001)^[22] in sapota; Kumar *et al.* (2011), Padma Lay *et al.* (2013)^[19] and Amit Desai *et al.* (2017)^[9] in papaya.

On the basis of pooled data (Table 2), at 60 days after sowing of papaya seeds, the maximum seedling vigour index-II of the papaya sapling 259.61 was recorded in the treatment T₁₁ (seed

soaking with 150 ppm GA₃ + 12 hours + Azotobacter) followed by 239.20 in treatment T₁₀ (seed soaking with 150 ppm GA₃ + 06 hours + Azotobacter) and 200.65 in the treatment T₁₅ (seed soaking with cow urine 10 (%) + 12 hours + Azotobacter).

In present study, the seed vigour index-II of sapling due to GA₃ pre-soaking can be correlated with higher seed germination, higher shoot length and root length and number of leaves has lead to over all assimilation and distribution of food material with the plant (Brain and Hemming,1955)^[8] and hence resulted in higher seedling vigour (Pampanna and Sulikeri, 2001)^[22]. The results were in close agreement with the findings of Amit Desai *et al.* (2017)^[9] in papaya; Patil *et al.* (2012) in Rangpur lime and Gurung *et al.* (2014) in passion fruit.

Table 1: Effect of pre-sowing seed treatment on germination behavior on papaya

Treatments	Days taken to germination			Rate of emergence			Germination percentage		
	2018	2019	Pooled	2018	2019	Pooled	2018	2019	Pooled
T ₁	11.67	12.33	12.00	61.57	57.87	59.73	65.00	63.33	64.17
T ₂	10.00	10.67	10.33	70.71	67.68	69.20	75.00	78.33	76.67
T ₃	09.33	09.67	09.50	71.15	68.59	69.87	80.00	78.33	79.17
T ₄	10.33	11.00	10.67	71.72	68.69	70.21	76.67	75.00	75.83
T ₅	09.33	10.00	09.67	68.79	65.76	67.28	80.00	76.67	78.33
T ₆	08.33	08.67	08.50	73.93	71.15	72.54	81.67	80.00	80.83
T ₇	07.33	07.67	07.50	73.26	70.70	71.98	81.67	83.33	82.50
T ₈	08.33	08.67	08.50	73.08	70.30	71.69	80.00	78.33	79.17
T ₉	07.33	07.67	07.50	65.81	63.25	64.53	81.67	80.00	80.83
T ₁₀	07.00	07.33	07.17	78.57	76.19	77.38	90.00	91.67	90.83
T ₁₁	06.67	07.00	06.83	80.42	78.33	79.38	95.00	93.33	94.17
T ₁₂	08.33	08.67	08.50	74.60	72.22	73.41	85.00	88.33	86.67
T ₁₃	07.33	07.33	07.39	75.64	73.26	74.45	86.67	90.00	88.33
T ₁₄	09.00	09.33	09.17	65.76	62.73	64.25	78.33	76.33	77.50
T ₁₅	08.67	09.00	08.83	68.69	65.66	67.18	80.00	78.33	79.17
T ₁₆	09.33	09.67	09.50	65.76	62.73	64.25	73.33	71.67	72.50
T ₁₇	09.00	09.33	09.17	65.56	62.22	63.89	75.00	73.00	74.17
SEm±	0.291	0.323	0.252	2.739	2.65	2.293	2.021	2.858	1.739
C.D. at 5%	0.841	0.933	0.729	7.907	7.650	6.619	5.834	8.250	5.018

Table 2: Effect of pre-sowing seed treatment on seedling vigour of papaya

Treatments	Seedling vigour index-I			Seedling vigour index-II			Root/ shoot ratio		
	2018	2019	Pooled	2018	2019	Pooled	2018	2019	Pooled
T ₁	1242.15	1215.23	1228.69	75.83	79.10	77.47	0.91	0.86	0.89
T ₂	2538.25	2540.47	2539.36	154.58	162.18	158.38	1.04	1.03	1.03
T ₃	2575.42	2579.28	2577.35	169.80	174.52	172.16	1.05	1.04	1.05
T ₄	1655.33	1649.13	1652.23	98.12	102.48	100.30	0.98	0.95	0.96
T ₅	1794.93	1802.73	1798.83	112.27	112.70	112.48	0.98	0.96	0.97
T ₆	2928.82	2876.00	2902.41	185.68	179.47	182.58	1.02	1.01	1.02
T ₇	2973.18	3043.70	3008.44	191.62	190.23	190.93	1.03	1.05	1.04
T ₈	1973.07	1940.28	1956.68	119.65	118.62	119.13	0.93	0.88	0.91
T ₉	2092.85	2057.33	2075.09	126.88	124.27	125.58	0.95	0.90	0.92
T ₁₀	3718.80	3792.10	3755.45	237.90	240.50	239.20	0.98	1.00	0.99
T ₁₁	3983.98	3895.68	3939.83	260.3	258.92	259.61	1.00	1.01	1.01
T ₁₂	2138.40	2225.45	2181.93	146.77	154.03	150.40	0.97	1.00	0.98
T ₁₃	2242.07	2337.60	2289.83	158.85	168.00	163.43	0.99	1.01	1.00
T ₁₄	2550.40	2518.80	2534.60	191.98	192.13	192.06	1.05	1.06	1.05
T ₁₅	2640.00	2615.35	2627.68	202.13	199.17	200.65	1.06	1.08	1.07
T ₁₆	1757.98	1723.28	1740.63	117.78	117.97	117.88	0.96	0.98	0.98
T ₁₇	1784.83	1753.50	1769.17	126.77	125.48	126.13	0.97	1.00	0.99
SEm±	61.719	65.511	51.665	4.585	5.787	4.555	0.011	0.030	0.019
C.D. at 5%	178.145	189.089	149.126	13.235	16.704	13.148	0.031	0.080	0.055

Conclusion

From the present study it may be concluded pre-sowing seed treatment of papaya gave significant result on improving the germination parameters and seedling vigour of papaya. Application of GA₃, with bio-fertilizers provide the additional

benefits on media and proved better in the germination under condition of Chhattisgarh. The treatment T₁₁ (Seed soaking with 150 ppm GA₃ + 12 hours + Azotobacter) may be recommended for the enhancement of seed germination and vigour of papaya.

References

- Anburani A, Shakila A. Influence of seed treatment on the enhancement of germination and seedling vigour of papaya. *Acta Hort* 2010;851:295-298.
- Anjanawe SR, Kanpure RN, Kachouli BK, Mandloi DS. Effect of plant growth regulators and growth media on seed germination and growth vigour of papaya. *Ann. Pl. and Soil Res* 2013;15(1):31-34.
- Anonymous, Horticultural Statistics at a Glance. Horticulture Statistics Division Department of Agriculture, Cooperation & Farmers' Welfare Ministry of Agriculture & Farmers' Welfare Government of India, 2018.
- Babu KD, Patel RK, Singh A, Yadav DS, De LC, Deka BC. Seed germination, seedling growth and vigour of papaya under North East Indian condition. *Acta Hort* 2010;851:299-306.
- Barche SK, Singh K, Singh DB. Response of seed treatment on germination, growth, survivability and economics of different cultivars of papaya (*Carica papaya* L.). *Acta Hort* 2010;851:279-281.
- Begum H, Lavania ML, Babu Ratan, GHVR. Effect of pre-sowing treatments on seed and seedling vigour in papaya. *Seed Research* 1987;15(1):9-15
- Babu KD, Patel RK, Singh A, Yadav DS, De LC, Deka BC. Seed germination, seedling growth and vigour of papaya under North East Indian condition. *Acta Hort* 2010;851:299-306.
- Brain PW, Hemming HG. The effects of GA on shoot growth of pea seedling. *Physiology of plant*, 1955;8:669-681.
- Desai Amit, Ashwin Trivedi, Bharat Panchal, Velji Desai. Improvement of papaya seed germination by different growth regulator and growing media under net house condition. *International Journal of Current Microbiology and Applied Sciences* 2017;6(9):828-834.
- Deb P, Das A, Ghosh SK, Suresh CP. Improvement of seed germination and seedling growth of papaya (*Carica papaya* L.) through different pre-sowing seed treatments. *Acta Hort* 2010;581:313-316.
- Dhankhar DS, Singh M. Seed germination and seedling growth of aonla (*Phyllanthus emblica* Linn.) as influenced by gibberellic acid and thio-urea. *Crop Research* 1996;12(3):363-366.
- Dhinesh Babu K, Patel RK, Singh A, Yadav DS, Singh A, Yadav DS *et al.* Seed germination, seedling growth and vigour of papaya under north east indian condition. *Acta Hort* 2010;851:299-306.
- Gholap SV, DOD VN, Bhuyar SA, Bharad SG. Effect of plant growth regulators on seed germination and seedling growth in aonla (*Phyllanthus emblica* L.) under climatic condition of Akola. *Crop Res* 2000;20(3):546-548.
- Gogoi D, Kotoky U, Hazarika. Effect of bio-fertilizers on productivity and soil characteristics of banana. *Indian J. Hort* 2004;61:354-56.
- Gurung N, Swamy GSK, Sarkar SK, Ubale NB. Effect of chemicals and growth regulators on germination, vigour and growth of passion fruit (*Passiflora edulis* Sims). *Bioscan* 2014;9(1):155-157.
- Hofmeyr JDJ. In Fruits – Tropical and Subtropical, Eds. Bose, T. K., Mitra, S. K. and Sanyal, D 1938;1:497.
- Kadam SS, Arumugam R, Balamohan TN. Effect of seed treatment with chemical on germination of papaya seed cv. Washington. National Seminar on Production and Utilization of Papaya. T.N.A.U., Coimbatore. 6-7 March, 1992, 26.
- Kumar P, Sehrawat SK, Dahiya OS, Dahiya DS. Papaya seed priming in relation to seed vigour. *Haryana J. Hort. Sci* 2011;40(1, 2):4-9.
- Padma Lay, Basvaraju GV, Sarika G, Amrutha N. Effect of seed treatment to enhance seed quality of papaya (*Carica papaya* L.) cv. Surya. *G.J.B.A.H.S* 2013;2(3):221-225.
- Palaniswamy V, Ramamoorthy K. Seed germination studies in papaya. *Progressive Horticulture* 1987;19(3-4):253-255.
- Paleg L. Physiological effects of gibberellins, *Annual Review of Plant Physiology* 1965;16:291-322.
- Pampanna Y, Sulikeri GS. Effect of growth regulators on seed germination and seedling growth of Sapota. *Karnataka J Agric. Sci* 2001;14:1030-1036.
- Panse VG, Sukhatme PV. *Statistical Method for agricultural workers*. Fourth edition. Indian Council of Agricultural Research, New Delhi 1985;1-381.
- Patil SR, Sonkamble AM, Waskar DP. Effect of growth regulators and chemicals on germination and seedling growth of Rangpur lime under laboratory conditions. *International J Agric. Sci.* 2012;8(2):494-497.
- Ram M Papaya. Indian council of Agricultural Research, New Delhi, 2005, 142-143.
- Sinish MS, Mercy G, John PS. Organic methods for cashew root stock production. *Cashew*.2005;19(1):8-15.
- Singh DK, Bhattacharya B, Mondal K. Role of pre-sowing seed treatment with different chemicals on germination behaviour and seedling growth of jackfruit (*Artocarpus heterophyllus* Lam). *Environment and Ecology* 2002;20(3):741-743.
- Sukhada M, Shivananda TN, Iyenger BRV. Uptake of ³²P labelled superphosphate by endomycorrhizal papaya (*Carica papaya* cv. Coorg Honey Dew). *Journal Nuclear of Agriculture* 1995;24(4):30-31.
- Suryakanth LB, Mukunda GK, Raghavendraprasad GC. Studies on seed germination in guava cvs. Taiwan guava and Allahabad safeda. *Karnataka J Horti* 2005;1(3):47-50.
- Sutheesh VK, Jijeesh CM, Divya TP. Evaluation of organic and inorganic pre-treatments for better seed germination and seedling vigour in (*Santalum album* L.) *Plant Archives* 2016;16(1):143-150.
- Vasu D, Kumari M, Zia-ul-Hasan. Effect of bio-fertilizers (Azotobacter and PSB) and their combinations on germination and survival of *Lens culinaris* Medic. *Environment and Ecology* 2010;28(1B):703-705.