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## Efficacy of different fungicides against the *Alternaria solani* under *in vitro* conditions

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**Abstract**

Among the six fungicides, most effective fungicides were found Mancozeb 75% WP which exhibited 100% inhibition in mycelium growth at 0.2% concentration followed by Tebuconazole 50% + Trifloxystrobin 25% w/w (75WG) 85% inhibition in mycelium growth at 0.1% concentration. However, Azoxystrobin (23% SC) shows the least inhibition in mycelium growth 36.11% at 0.1% concentration.

**Keywords:** Fungicides, *in vitro*, *Alternaria solani*, mycelial inhibition

**Introduction**

Tomato (*Lycopersicon esculentum* Mill.) is a commonly cultivated vegetable in the world and is the second largest profitable solanaceous vegetable crop after potatoes (Sahu *et al.*, 2013). Tomato is a model species for classical genetics and genomic research. Tomato is of high medicinal importance. It functions as a booster of gastric secretion and a purifier of blood. It is common because it supplies vitamin C and adds colour and flavour to food.

Tomato suffers from numerous pathogens, including fungi, bacteria, viruses, nematodes, etc., in many countries (Mark *et al.*, 2006). More than 200 diseases have been reported to infect tomatoes worldwide (Atherton and Rudich, 1986). Huge numbers of fungal diseases such as early blight (*Alternaria solani*), late blight (*Phytophthora infestans*), Septoria leaf blight (*Septoria lycopersici*), Powdery mildew (*Oidiopsis taurica*), Fusarium wilt (*Fusarium oxysporum* f. sp. *lycopersici*), Collar rot (*Sclerotium rolfsii*) and Damping off (*Pythium* sp.) trigger significant tomato losses. Amongst these fungal diseases, the early blight caused by *Alternaria solani* is among the most economically significant diseases of the crop nation and the world (Jones *et al.*, 1991). Genus *Alternaria* refers to deuteromycetes of various types, which are harmful plant parasites for families such as Solanaceae, cucurbitaceae, brassicaceae. The members of the *Alternaria* genus are cosmopolitan, living as saprophytes as well as weak parasites. In a variety of instances, tiny dark spots are often produced on pods and tender twigs (Valkonen and Koponen, 1990). *Alternaria*-related diseases are among the most prevalent diseases of many kinds of plants in the world. Absolute gross losses incurred by the numerous *Alternaria* in all their hosts are among the largest caused by any pathogen (Agrios, 2005). Khalid *et al.* (2004) and Deshwal offer a detailed, comparative account of the morphological distinction of the various types of *Alternaria* found in cucurbitaceous, brassicaceous and solanaceous crops (2004). *Alternaria* species are foliar pathogens that trigger comparatively slow degradation of host tissues through reducing photosynthetic ability. Infection contributes to the development of necrotic lesions, which often occur as goal due to growth interruptions due to unfavourable circumstances. The fungus remains in the middle of the lesion, which is surrounded by an uninvaded chlorotic halo, a symptom that is typically found in the course of necrotrophic pathogen infection.

*Alternaria* has no known sexual stage or overwintering spores, but the fungus may live for a long time as mycelium or spores on rotting plant debris or as a latent infection in seeds (Rotem, 1994). In Maharashtra, the disease is predominantly observed in all tomato-growing regions. To manage this disease, many fungicides like mancozeb, captan, carbendazim, propineb, copper oxychloride, tebuconazole and propiconazole are sprayed. Use of such chemicals will be further more in protective agriculture conditions as tomato is one of the vegetable crops promoted under intensive cultivation under protected (polyhouse) conditions. Farmers are already using fungicides for management of this pathogen. However, being a commercial crop, indiscriminate use of these fungicides will lead to residues in the fruits and also environmental, water and soil pollution.

Some alternative methods have been tried in the past but with limited success. Considering the economic importance of the crop and yield losses caused by early blight of tomato and its effect on yield, the present study is focus on *in vitro* efficacy of fungicides against *Alternaria solani*.

## Material and Methods

### *In vitro* evaluation of fungicides

The poisoned food technique (Falck, 1907) was followed to evaluate the efficacy of six different fungicides against *A. solani* at its recommended dose of applications (concentrations). Both non-systemic and systemic fungicides were tested *in vitro* against *A. solani*. Fungicides were added

to the sterilized potato dextrose agar medium as per treatment details. Five mm disc of *A. solani* was taken from seven days old culture and placed at center of petri dish. Simultaneously, a control was also maintained by growing the fungus on fungicide free PDA medium.

The plates were incubated at  $25 \pm 2^{\circ}\text{C}$ . Each treatment was replicated four times. The efficacy of fungicides was recorded by measuring the colony diameter of *A. solani* in each treatment and compared with control. The per cent growth inhibition of the fungus in each treatment in comparison with control was calculated by the equation given by Vincent (1947). *In vitro* evaluation of fungicides treatment details is given below:

### Treatments details:

**Table 1:** Treatments details of Fungicides

Treatment No.	Recommended dose (% Concentration)	Name of Fungicides
T <sub>1</sub>	0.2	Mancozeb (75% WP)
T <sub>2</sub>	0.1	Carbendazim (50% WP)
T <sub>3</sub>	0.3	Copper Oxychloried (50% WP)
T <sub>4</sub>	0.1	Nativo (75 WG)
T <sub>5</sub>	0.25	Propineb (70% WP)
T <sub>6</sub>	0.1	Azoxystrobin (23% SC)
T <sub>7</sub>	-	Control

Replications: Four, Design: CRD

### Observation recorded

The radial growth of the fungus on the poisoned medium was recorded at time of mycelium growth reached 90 mm in control. Per cent inhibition of mycelium growth of the fungus was calculated by using the formula described by Vincent (1947).

$$I = \frac{C - T}{C} \times 100$$

Where,

I = Per cent inhibition

C = Radial growth in control

T = Radial growth in treated (fungicide/ botanicals/ bioagents).

### Result and Discussion

#### Evaluation of fungicides against *Alternaria solani*

Six fungicides *viz.* Mancozeb (75% WP), Carbendazim (50% WP), Copper oxychloride (50% WP), Tebuconazole 50% + Trifloxystrobin 25% w/w (75 WG), Propineb (70% WP) and Azoxystrobin (23% SC) were assessed *In vitro* to find out the most effective fungicide against *A. solani* at its recommended concentrations using poison food technique.

The results are presented in Table 4.20 reveal that the significant difference among fungicides against *A. solani* was observed. Fungus growth was checked in Mancozeb (75% WP) at 0.2%, followed by Tebuconazole 50% + Trifloxystrobin 25% w/w (75 WG) at 0.1% and Copper oxychloride (50% WP) at 0.25% concentration (Plate 4.10).

**Table 2:** *In vitro* evaluation of different fungicides against *Alternaria solani*

Tr. No.	Treatment Details	Recommended Dose. (% concentration)	Average Mycelium growth (mm)*	Average mycelium growth inhibition over control. (%)
T <sub>1</sub>	Mancozeb (75% WP)	0.2	00.00	100 (90.00)
T <sub>2</sub>	Carbendazim (50% WP)	0.1	54.75	39.16 (38.74)
T <sub>3</sub>	Copper oxychloride (50% WP)	0.25	22.00	75.55 (60.37)
T <sub>4</sub>	Tebuconazole 50% + Trifloxystrobin 25% w/w(75 WG)	0.1	13.50	85.00 (67.21)
T <sub>5</sub>	Propineb (70% WP)	0.25	52.00	42.22 (40.52)
T <sub>6</sub>	Azoxystrobin (23% SC)	0.1	57.50	36.11 (36.94)
T <sub>7</sub>	Control	--	90.00	00.00 (0.00)
	S.Em. $\pm$	CD at 5%	CD at 1%	CV (%)
	0.76	2.24	3.05	3.68

CD at 5% level of significance

\*Average of four replications

Values in parenthesis (s) are angular transformed value.

Among the six fungicides, most effective fungicides were found Mancozeb 75% WP which exhibited 100.00 per cent inhibition in mycelium growth at 0.2% followed by Tebuconazole 50% + Trifloxystrobin 25% w/w (75 WG) 85 per cent inhibition in mycelium growth at 0.1% and Copper

oxychloride (50% WP) 75.55 per cent inhibition in mycelium growth at 0.25% concentration. However, Azoxystrobin (23% SC) shows the least inhibition in mycelium growth 36.11 at 0.1 per cent concentration (Fig. 1).



**Fig 1:** Least inhibition in mycelium growth 36.11 at 0.1 per cent concentration

The present findings are similar with the result of Choulwar and Datar (1994) [6] reported that the Mancozeb (1000 ppm) was the most effective fungicide with 77 per cent growth inhibition followed by Captafol. Kamble *et al.* (2000) [13] tested different fungicide and reported that mancozeb was most effective followed by copper oxychloride inhibiting the mycelial growth of *A. alternata* causing leaf spot of tomato. Prasad and Naik (2003) found iprodione, mancozeb and copper oxychloride as most effective and thus inhibited more than 75.00 per cent of mycelial growth at 0.25 per cent concentration. Similar type of result were also obtained by Roopa *et al.* (2014) [28] found that, there was a significant difference among contact fungicides in inhibiting the growth of *A. solani*. Among the three contact fungicides evaluated, mancozeb (63.20%) was significantly superior over other treatments.

Sadana and Didwania (2015) [29] tested seven fungicides against *Alternaria solani* and highest reduction in the disease was achieved by applying Mancozeb (1500 ppm) that caused 86.4 per cent inhibition of mycelial growth of *Alternaria solani*. Similar results were also obtained by El-Nazar *et al.* (1970) [8], Lodha and Prasad (1975) [27] found that Dithane Z-78 effectively checked the growth of *A. solani* in pot trials and *In vitro*, Petkar and Utikar (1976) [26] reported that mancozeb was effective against *A. solani*, Kalra and Sohi (1984), Mohammad (1988), Abhinandan *et al.* (2004) [1], Chethana *et al.* (2012) [4].

## References

1. Abhinandan D, Randhawa HS, Sharma RC. Incidence of *Alternaria* leaf blight in tomato and efficacy of commercial fungicides for its control. *Annals of Biology* 2004;20:211-218.
2. Arunakumara KT, Kulkarni MS, Thammaiah N, Hegde Y. Fungicidal management of early blight of tomato. *Indian Phytopath* 2010;63(1):96-97.
3. Arunakumara KT, Kulkarni MS, Thammaiah N, Hegde Y. Fungicidal management of early blight of tomato. *Indian Phytopath* 2010;63(1):96-97.

4. Chethana BS, Girija Ganeshan, Rao AS, Bellishree K. *In vitro* evaluation of plant extracts, Bioagents and Fungicides against *Alternaria porri* (Ellis) Cif. Causing purple blotch disease of onion. *Pest Manag. in Horti. Ecosystem* 2012;18(2):194-198.
5. Chohan S, Perveen R, Mehmood MA, Naz N, Akram N. Morpho-physiological studies, management and screening of Tomato germplasm against *Alternaria solani* the causal agent of Tomato early blight. *Int. J Agril. Bio* 2015;17(1):111-118.
6. Choulwar AB, Datar VV. Tolerance of *Alternaria solani* to fungicides, *J. Maharashtra Agril.Unvi* 1994;19(1):133-134.
7. Dubey SC, Patel B, Jha DK. Chemical management of *Alternaria* blight of broad bean. *Indian Phytopath* 2000;53(2):213-215.
8. El-Nazar, Tantawy I, Zield G, Eldeprawy MS, Elals ABD. Chemical control of early and late blight of tomato. *Acta. Agron. Hung* 1970;19(34):384-385.
9. Ganeshan G, Chethana BS. Bioefficacy of Pyraclostrobin 25% EC against early blight of tomato. *World Applied Sci. J* 2009;7(2):227-229.
10. Ghazanfar MU, Raza W, Ahmed KS, Qamar J, Haider N. Evaluation of different fungicides against *Alternaria solanisora* cause of early blight of tomato under laboratory conditions. *Int. J. Zoology* 2016;1(5):08-12.
11. Hassan M, Farooq M, Gul JF. *In vitro* evaluation of some fungicides against common fungal pathogen of early blight and fruit rot of Tomatoes. *Appl. Environ. Biol. Sci* 2014;4(9):424-429.
12. Islam R, Kawser-E-Jahan, Ali S. Efficacy of fungicides for controlling early blight of tomato. *Bangladesh J. Plant Patho* 2012;28(1-2):25-27.
13. Kamble PV, Ramiah M, Patil DV. Efficacy of fungicide controlling leaf spot disease of tomato caused by *A. alternata* (Fr.) Keissler. *J. Soils and Crops* 2000;10:36-38.
14. Kamble SB, Pawar DR, Sankeshware SB, Sawant UK. *In vitro* efficacy of fungitoxicants against *Alternaria solani*. *Int. J. Res. Agril. Sci* 2009;5(1):137-139.
15. Karla JS, Sohi HS. Studies on post-harvest rots of tomato fruits rot. *Indian J. Mycol. and Pl. Pathol* 1985;15:176-178.
16. Khatri DNK, Jagdish Prasad, Maheshwari SK. Efficacy of fungicide against early blight of tomato caused by *Alternaria solani*. *Indian J. Ann. Pl. Protect. Sci* 2013;22(1):148-151.
17. Kodmelwar RV, Wangikar PD, Shukla VN. *In vitro* studies of some copper-based fungicides against *Colletorichum capsici* and *Alternaria solani*. *PKV Research J* 1973;1:212-213.
18. Lodha PC, Prasad N. Efficacy of some fungicides and antibiotics against alternariosis of tomato. *Phytopathologia Mediterranea* 1975;14:21-22.
19. Mallikarjun G. Studies on *Alternaria alternata* (Fr.) Keissler a causal agent of leaf blight of turmeric (*Curcuma longa* L.). M. Sc. (Agri.) Thesis, Univ. Agril. Sci., Dharwad, India 1996.
20. Mesta RK, Benagi VI, Shankergroud I, Megeri SN. Effect of dates of sowing and correlation of weather parameters on the incidence of *Alternaria* blight of sunflower. *Karnataka J. Agric. Sci* 2009;22(2):441-443.
21. Mohammad A, Chawla S, Zaidi NW, Rayar JK, Variar M, Singh US, *et al.* Development of specific primers for genus *Fusarium* and *F. solani* using rDNA sub-unit and

- transcription elongation factor (TEF-1 $\alpha$ ) gene. Afr. J. Biotechnol 2012;11(2):444-447.
22. Mohammad SE. Control of tomato early blight under plastic house condition. Mesopotamia J Agric 1988;20(2):359-366.
  23. Patel HV. Studies on early blight of tomato caused by *A. solani* (Ellis and Mart.) under north Gujarat conditions. M.Sc. (Agri.) Thesis, Submitted to Sardarkrushinagar, Dantewada Agricultural University, Sardarkrushinagar 2007, pp. 68.
  24. Patel RL, Choudhary RF. Management of *Alternaria solani* causing early blight of tomato with fungicides. J. Plant Dis. Sci 2010;5(1):65-67.
  25. Patel RL, Choudhary RF. Management of *Alternaria solani* causing early blight of tomato with fungicides. J. Plant Dis. Sci 2010;5(1):65-67.
  26. Petkar AS, Utikar PG. Note on efficacy of fungicides for the control of *Alternaria* blight of tomato. Pantanagar J. Res 1976;1(1):77-78.
  27. Prasad Y, Naik MK. Evaluation of genotypes, fungicides and plant extracts against early blight of tomato caused by *Alternaria solani*. J. Pl. Protect 2003;31:49-53.
  28. Roopa RS, Yadahalli KB, Kavyashree MC. Evaluation of natural plant extracts, antagonists and fungicides against early blight caused by *A. solani in vitro*. Int. J. Lif. Sci 2014;9(3):1309-1312.
  29. Sadana D, Didwania N. Bio efficacy of fungicides and plant extracts against *Alternaria solani* causing early blight of Tomato. J. Agro. Ecol 2015;2(3):181-186.
  30. Sahu DK, Khare CP, Singh HK, Thakur MP. Evaluation of newer fungicide for management of early blight of tomato in Chhattisgarh. The Bioscan 2013;8(4):1255-1259.
  31. Sharma RN, Gaur RB. Management of post-harvest core rot, *Alternaria alternata* in Kinnow, Citrus deliciosa fruits. Indian J. Plant Protect 2009;37(1-2):207-210.
  32. Singh PC, Singh D. *In vitro* evaluation of fungicides against *Alternaria alternata*. Annals of Pl. Protect. Sci 2006;14(2):500-502.
  33. Thippeswamy M, Krishnappa CN, Chakravarthy AM, Sathisha Jyothi SU, Kumar V. Pathogenicity and management of phomopsis blight and leaf spot in brinjal caused by *Phomopsis vexans* and *Alternaria solani*. Indian Phytopathol., 2006;59(4):475-481.
  34. Vijay Kumar, Gurvinder Singh, Ankur Tyagi. Evaluation of different fungicides against *Alternaria* leaf blight of Tomato (*Alternaria solani*). Int. J Curr. Microbiol. App. Sci 2017;6(5):2343-2350.