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***In vitro* evaluation of fungicides against mycelial growth and sclerotial viability of *Sclerotinia sclerotiorum* (Lib.) de Bary, the cause of Sclerotinia rot of Rapeseed-mustard**

Kiran Goswami, AK Tewari and Pooja Upadhyay

Abstract

Sclerotinia rot of rapeseed-mustard caused by *Sclerotinia sclerotiorum* (Lib) de Bary has become a serious problem in India since last few successive years. Among eleven fungicides tested *in vitro*, Thiophenate methyl and carbendazim + mancozeb (SAAF) proved to be the best as they completely inhibited the mycelial growth of the pathogen at 250 µg a.i./ml. The fungicides viz., iprodione, hexaconazole, triademefon, tebuconazole, carbendazim, benomyl, propiconazole and mancozeb were also effective in inhibiting mycelial growth (92.6 to 97.7%) at 250 µg a.i./ml. The metalaxyl+ mancozeb (ridomil 72 MZ) was found least effective in checking the growth of the pathogen. Benomyl and carbendazim were found most effective in inhibiting sclerotial germination, while others were ineffective. Complete inhibition of sclerotial germination was observed when sclerotia were dipped in benomyl at 100 µg a.i./ml either 5 or 15 minutes, while carbendazim showed complete inhibition at 250 µg a.i./ml. only in 15 minutes dip treatment.

Keywords: fungicides, rapeseed-mustard, sclerotinia sclerotiorum, sclerotial viability

1. Introduction

Among different crops in India, rapeseed-mustard occupies a premier position accounting for 25 and 30 per cent of total oil seed acreage and production respectively. The disease was of minor importance till few years back, but recently it has turned out to be a serious problem in major rapeseed-mustard growing areas in the country and under severe infestation it causes seed yield losses up to 74 per cent (Chauhan *et al.*, 1992; Lodha *et al.*, 1992; Ghasolia *et al.*, 2004; Kang and Chahal, 2000; Krishnia *et al.*, 2000; Shivpuri *et al.*, 2000, Singh *et al.* 2017a; Singh *et al.* 2017b; Singh *et al.* 2017c; Singh *et al.* 2018; Tiwari *et al.* 2018; Tiwari *et al.* 2019a; Tiwari *et al.* 2019b; Kour *et al.* 2019; Singh *et al.* 2019) [3, 9, 4, 7, 8, 16, 18, 19, 20, 21, 22, 23, 24, 25, 26] or more.

The explosive pathogenicity of the fungus under favourable conditions and ability of sclerotia to withstand under adverse conditions allow it to cause infection on many crops. Use of chemicals has been a practical method of controlling the disease. The fungicides viz, benomyl, carbendazim, thiophenate methyl, iprodione, vinclozolin and mancozeb were found very effective in inhibiting mycelial growth of *S. sclerotiorum*, *in vitro* (Roy and Saikia, 1979; Sharma and Kapoor, 1997; Shivpuri and Gupta, 2001; Chattopadhyay *et al.* 2002) [11, 13, 15, 21]. Sharma and Basandrai (1997) [12] evaluated the effect of fungicides on sclerotial viability of *S. sclerotiorum* and observed that carbendazim and triademefon were most effective in inhibiting sclerotial germination at 100 ppm. The present studies were conducted to screen out the effective fungicides which may completely inhibit the mycelial growth as well as sclerotial germination of *S. sclerotiorum* and could be exploited under natural conditions for the management of the disease.

2. Materials and methods

Eleven systemic and non-systemic fungicides (Table 1) at four different concentrations viz.,

25, 50, 100 and 250 µg a.i./ml were evaluated against *S. sclerotiorum* pathogen *in vitro* to evaluate the relative efficacy of fungicides in inhibiting the radial growth and sclerotial viability. The effect of fungicides on radial growth of the pathogen was evaluated by 'Food Poison Technique' (Schmitz, 1930) [14] and sclerotial viability by dipping the surface sterilized sclerotia in the fungicide solution for 5 and 15 minutes.

2.1 Effect on radial growth

In poisoned food technique, requisite concentration of each fungicide (prepared by stock solution) was incorporated into PDA flasks, mixed thoroughly by shaking and poured 20 ml in each Petri plates. The medium was allowed to solidify and then inoculated with 5 mm discs of 5 day-old culture of the pathogen. The mycelial disc was placed at the centre of each Petri plate and incubated at 20±1°C. The Petri plates inoculated with the pathogen but without any fungicide were served as control. Three replications were kept for each treatment. The observations on radial growth (mm) of fungal colony were measured 5 days after incubation when control Petriplates was filled with the growth of the pathogen. The mycelial inhibition of the pathogen over control was calculated by using formula.

Per cent inhibition over control

Where,

C = growth of fungus in control

T = growth of fungus in treatment

2.2 Effect on sclerotial viability (germination)

The effect of fungicides and their concentrations on sclerotial viability of the pathogen were evaluated by dip treatment of surface sterilized sclerotia for 5 and 15 minutes. Stock solution of 10,000 µg a.i./ml was prepared by adding required quantity of fungicide in sterilized distilled water and used for the preparation of different concentrations. The treated sclerotia (15 no.) dried under aseptic conditions were placed in Petri plates. Each treatment was replicated thrice. Petri plates inoculated with untreated sclerotia (dipped in sterilized distilled water) were served as control. These Petri plates were incubated at 20±1°C for 5 days. Observations on number of germinated sclerotia (myceliogenic) were recorded and per cent inhibition of sclerotial germination over control was calculated.

3. Results and Discussion

3.1 Effect on radial growth

Pathogen growth in various fungicides was measured and average colony dia. (mm) of the colony in each fungicide was recorded. The data (Table 1 and Fig.1) revealed that all fungicides invariably inhibited radial growth of the pathogen

at different concentration (Plate 1). Among all the fungicides evaluated, thiophenate methyl was found most effective in inhibiting radial growth of fungus (95.5%) followed by iprodione (92.6%) and hexaconazole (90.7%) at 25 µg a.i./ml and were statistically at par but significantly different from other treatments. Same results were obtained at 50 µg a.i./ml. Thiophenate methyl showed 97.7 per cent mycelial inhibition over check at 100 µg a.i./ml followed by iprodione, triademefon (95.1%) and SAAF (94.0%) and were at par with each other but significantly different from other treatments in inhibiting mycelial growth. Complete inhibition of radial growth was observed with thiophenate methyl and SAAF at 250 µg a.i./ml and were at par with each other but significantly different from other treatments whereas, ridomil 72 MZ was found least effective in inhibiting the growth of the pathogen. Among the various concentration of fungicide, no significant differences were observed at 50, 100 and 250 µg a.i./ml in propiconazole, thiophenate methyl, benomyl, iprodione, tebuconazole, hexaconazole and at 100 & 250 µg a.i./ml in triademefon and carbendazim. However, significant differences were observed at 100 and 250 µg a.i./ml with mancozeb and SAAF in inhibiting mycelial growth of the pathogen. Studies made *in vitro* resulted that thiophenate methyl and SAAF (carbendazim + mancozeb) proved to be the best among eleven fungicides evaluated. These two fungicides completely inhibited the mycelial growth of the pathogen at 250 µg a.i./ml. The fungicides *viz.*, iprodione, hexaconazole, triademefon, tebuconazole, carbendazim, benomyl, propiconazole and mancozeb were also found effective at 100 µg a.i./ml as they inhibited more than 90 per cent fungal growth.

The results obtained in the present study are in close agreement with the findings of earlier workers who also reported that benomyl (Roy and Saikia, 1979) [11] and carbendazim and thiophenate methyl (Shivpuri and Gupta, 2001) [15] completely inhibited the mycelial growth of the fungus at 0.05 per cent. However, Sharma and Kapoor (1997) [13] observed, complete inhibition of mycelial growth of the pathogen at higher dose of iprodione (0.5%). Chattopadhyay *et al.* (2002) [2] also reported complete inhibition of fungal growth at higher dose of carbendazim (0.1% a.i.) and mancozeb (0.2% a.i.) Singh (1998) [27] also observed that, iprodione vinclozolin, benomyl, metalaxyl and carbendazim completely inhibited the growth of the fungus only at higher dose (0.2%). Benomyl completely inhibited the fungal growth even at 10 ppm (Singh *et al.*, 1994) [17] while vinclozolin inhibited the growth even at 1.0 µg a.i./ml (Mueller *et al.*, 2002). Goswami *et al.* (2007) [10, 5] observed thiophenate methyl and SAAF (carbendazim + mancozeb) proved to be the best as they completely inhibited the mycelial growth of the pathogen at 250 µg a.i./ml. The hexaconazole and SAAF which not reported earlier in the literature were found effective in the present investigation against the pathogen.

Table 1: Effect of fungicides on radial growth of *S. sclerotiorum* (5 DAI)

Fungicide	Concentration (µg a.i./ml)							
	25		50		100		250	
	*RG	I (%)	*RG	I (%)	*RG	I (%)	*RG	I (%)
Propiconazole (25% EC)	22.0	75.5	10.3	88.5	6.3	92.6	6.3	92.6
Mancozeb (75% WP)	90.0	0.0	86.6	3.7	7.3	91.8	2.0	97.7
Metalaxyl 8% WP + MZ 64% WP	90.0	0.0	90.0	0.0	50.6	43.7	10.3	88.5
Thiophenate methyl (70% WP)	4.0	95.5	4.0	95.5	2.0	97.7	0.0	100.0
Tebuconazole (25.9% EC)	26.0	71.1	17.3	80.7	4.0	93.3	6.0	95.5
Benomyl (50% WP)	9.0	90.0	7.6	91.4	7.3	91.8	4.6	94.8
Triademefon (25% WP)	17.3	80.7	7.0	92.2	4.3	95.1	2.0	97.7

Iprodione (50% WP)	6.6	92.6	6.3	92.9	6.3	95.1	4.0	95.6
Carbendazim 12% WP + MZ 63% WP	10.6	88.1	9.0	90.0	4.6	93.7	0.0	100.0
Carbendazim (50% WP)	17.6	80.3	8.3	90.7	5.3	94.0	3.3	96.3
Hexaconazole (5% SC)	8.3	90.7	7.6	91.4	6.6	92.5	3.6	95.9
Control	90.0							
CD (P=0.05)	Fungicides 7.1		Concentrations 4.3		Fungicide x Concentration 14.1			
CV (%)	12.7							

* Mean of three replications

Transformed values are used for analysis

RG – Radial growth (mm)

I- Inhibition

MZ- Mencozeb

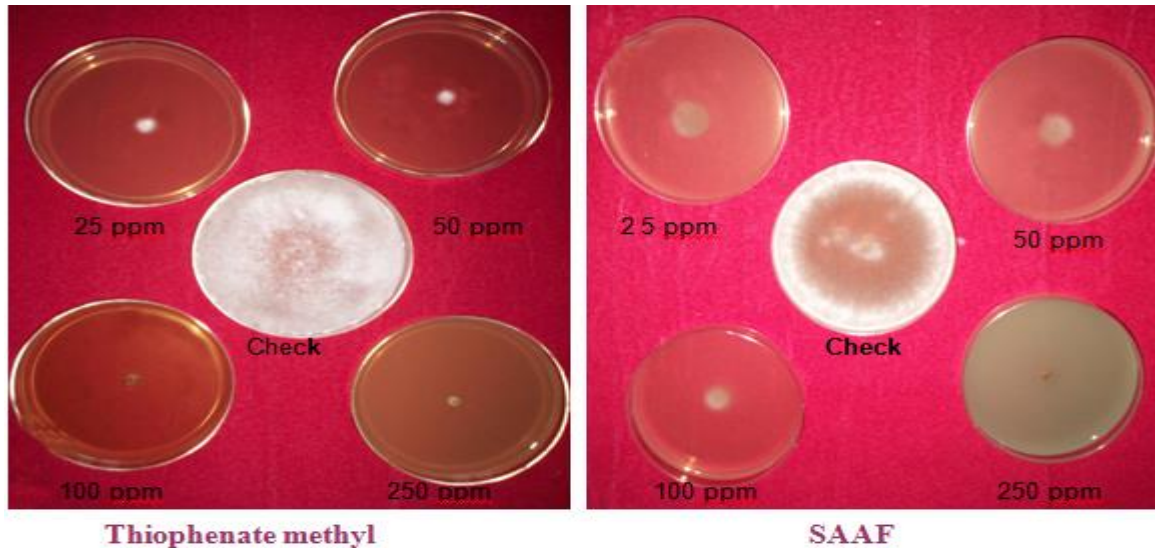


Plate 1: Effect of fungicides on radial growth of *S. sclerotiorum*

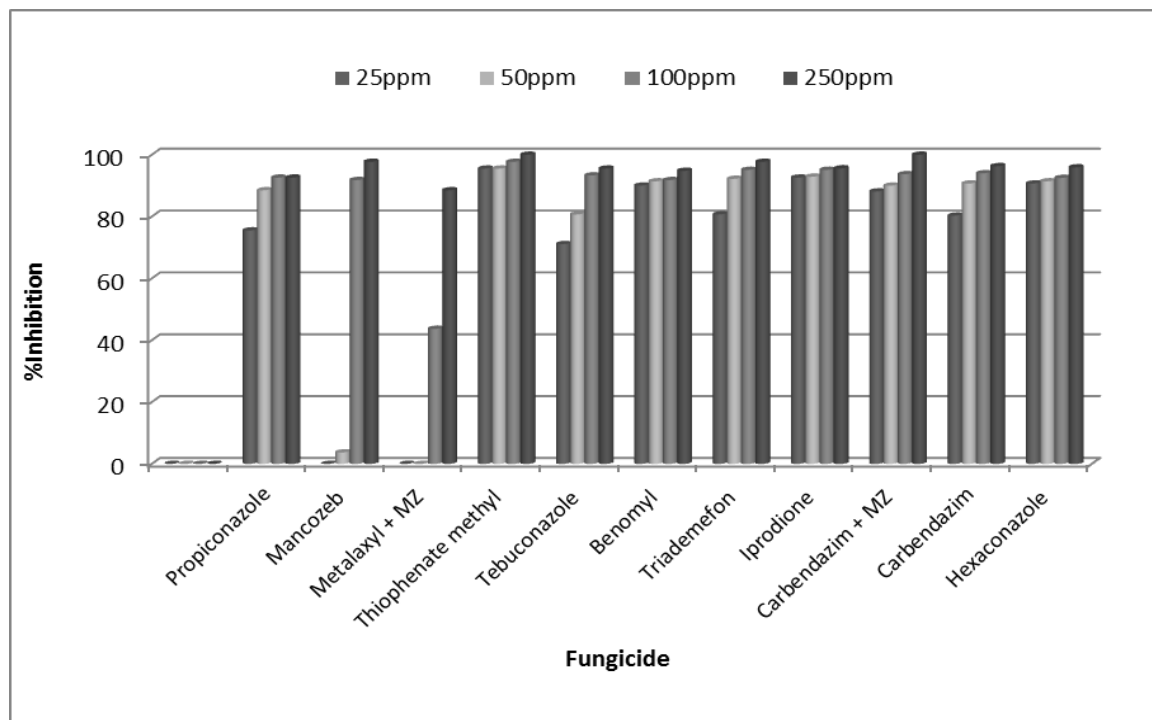


Fig 1: Effect of fungicides on radial growth of *S. sclerotiorum*

3.2 Effect on sclerotial viability

The observation on myceliogenic germination of sclerotia was recorded after 5 days of incubation in sclerotia treated separately with each fungicide by dipping at different concentration for 5 (Table 2a and Fig.2) and 15 minutes (Table 2b and Fig.3).

3.2.1 Effect of 5 minutes dip treatment:

During 5 minute dip treatment among various fungicides, only benomyl at 100 μg a.i./ml completely inhibited the sclerotial germination treated for 5 minutes followed by carbendazim (73.3 & 93.3%), thiophenate methyl (50.0 & 66.6%) and SAAF (46.6 & 63.3%) at 100 & 250 μg a.i./ml respectively. However, other fungicides were found ineffective (Plate 2, Table 2a and Fig.2).

Table 2a: Effect of fungicide on sclerotial viability (5 minutes dip treatment)

Fungicide	Concentration ($\mu\text{g a.i./ml}$)							
	25		50		100		250	
	*SG	I (%)	*SG	I (%)	*SG	I (%)	*SG	I (%)
Propiconazole (25% EC)	15.0	0.0	15.0	0.0	15.0	0.0	15.0	0.0
Mancozeb (75% WP)	15.0	0.0	15.0	0.0	15.0	0.0	15.0	0.0
Metalaxyl 8% WP + MZ 64% WP	15.0	0.0	15.0	0.0	15.0	0.0	15.0	0.0
Thiophenate methyl (70% WP)	14.0	6.6	8.0	46.6	7.5	50.0	5.0	66.6
Tebuconazole (25.9% EC)	15.0	0.0	15.0	0.0	14.0	6.6	13.5	10.0
Benomyl (50% WP)	10.5	30.0	7.0	53.3	0.0	100.0	0.0	100.0
Triadimefon (25% WP)	15.0	0.0	15.0	0.0	15.0	0.0	15.0	0.0
Iprodione (50% WP)	15.0	0.0	15.0	0.0	15.0	0.0	15.0	0.0
Carbendazim 12% WP + MZ 63% WP	15.0	0.0	9.5	36.6	8.0	46.6	5.5	63.3
Carbendazim (50% WP)	6.6	56.6	5.0	66.6	4.0	73.3	1.0	93.3
Hexaconazole (5% SC)	15.0	0.0	14.0	6.6	14.0	6.6	11.5	23.3
Control	15.0							
CD (P=0.05)	Fungicides-6.0		Concentrations - 3.6;		Fungicide x Concentration - 12.0			
CV (%)	30.0							

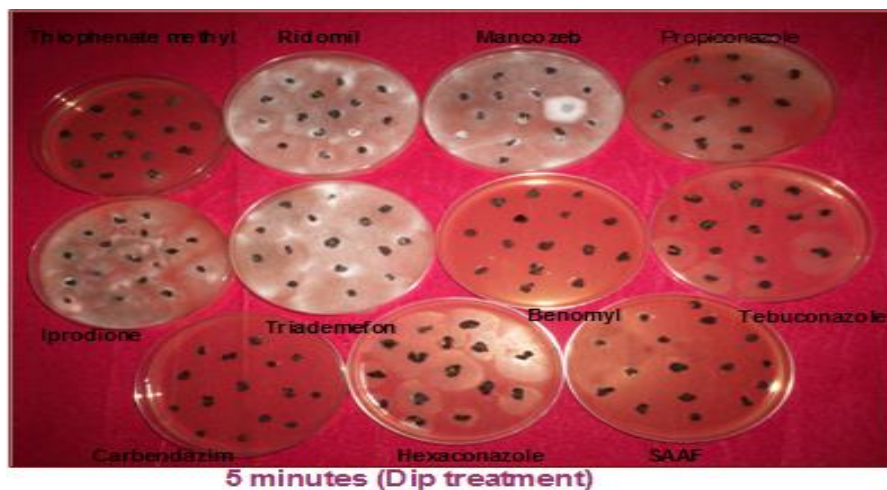
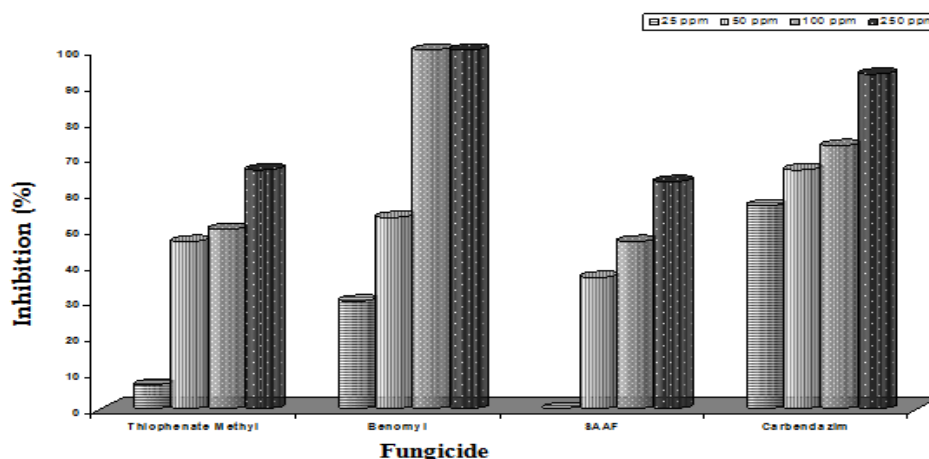
* Mean of three replications (15 sclerotia / plate)

Transformed values are used for analysis

SG – Sclerotia germinated

I- Inhibition

MZ- Mencozeb

**Plate 2:** Effect of fungicides on sclerotial germination (5 minutes dip treatment)**Fig.2:** Effect of fungicides on sclerotial germination (5 minutes dip treatment)

3.2.2 Effect of 15 minutes dip treatment:

When sclerotia were treated for 15 minutes, benomyl and carbendazim were found significantly most effective in inhibiting sclerotial germination (66.6 and 66.6%) at 25 $\mu\text{g a.i./ml}$ and 91.6 and 75.0 per cent at 50 $\mu\text{g a.i./ml}$ respectively, as compared to remaining fungicides. Significantly higher inhibition of sclerotial germination was

observed at 100 $\mu\text{g a.i./ml}$ with SAAF (66.6%) followed by thiophenate methyl (41.6%) and propiconazole (41.6%). Complete inhibition of sclerotial germination was observed with benomyl and carbendazim followed by propiconazole (91.6%), thiophenate methyl (75.0%), tebuconazole (75.0%), SAAF (66.6%) and hexaconazole (58.7%) at 250 $\mu\text{g a.i./ml}$ (Plate 3, Table 2b and Fig.3). However, mancozeb,

triadimefon and iprodione were found significantly less effective. Ridomil 72 MZ was found ineffective even at higher concentration. Among various concentration of effective fungicides no significant differences were observed

at 100 & 250 $\mu\text{g a.i./ml}$ with benomyl and SAAF in inhibiting sclerotial germination. However, significant differences were observed with carbendazim at 100 & 250 $\mu\text{g a.i./ml}$.

Table 2b: Effect of fungicide on sclerotial viability (15 minutes dip treatment)

Fungicide	Concentration ($\mu\text{g a.i./ml}$)							
	25		50		100		250	
	*SG	I (%)	*SG	I (%)	*SG	I (%)	*SG	I (%)
Propiconazole (25% EC)	5.5	8.3	5.0	16.3	3.5	41.6	0.5	91.6
Mancozeb (75% WP)	6.0	0.0	6.0	0.0	5.5	8.3	5.0	16.6
Metalaxyl 8% WP + MZ 64% WP	6.0	0.0	6.0	0.0	6.0	0.0	6.6	0.0
Thiophenate methyl (70% WP)	6.0	0.0	5.0	16.6	3.5	41.6	1.5	75.0
Tebuconazole (25.9% EC)	5.5	8.3	5.0	16.6	4.5	25.0	1.5	75.0
Benomyl (50% WP)	2.0	66.6	0.5	91.6	0.0	100.0	0.0	100.0
Triadimefon (25% WP)	5.5	8.3	5.5	16.6	4.5	25.0	2.5	33.3
Iprodione (50% WP)	5.5	8.3	5.0	8.3	5.0	25.0	4.0	25.0
Carbendazim 12% WP + MZ 63% WP	6.0	0.0	2.5	58.3	2.0	66.6	2.0	66.6
Carbendazim (50% WP)	2.0	66.6	1.5	75.0	1.5	75.0	0.0	100.0
Hexaconazole (5% SC)	6.0	0.0	4.5	25.0	4.5	25.0	2.5	58.7
Control	6.0	0.0	6.0	0.0	6.0	0.0	6.0	0.0
CD (P=0.05)	Fungicides 15.2		Concentrations 9.2		Fungicide x Concentration 30.5			
CV (%)	30.0							

* Mean of three replications (15 sclerotia / plate)

Transformed values are used for analysis

SG – Sclerotia germinated I- Inhibition

MZ- Mancozeb

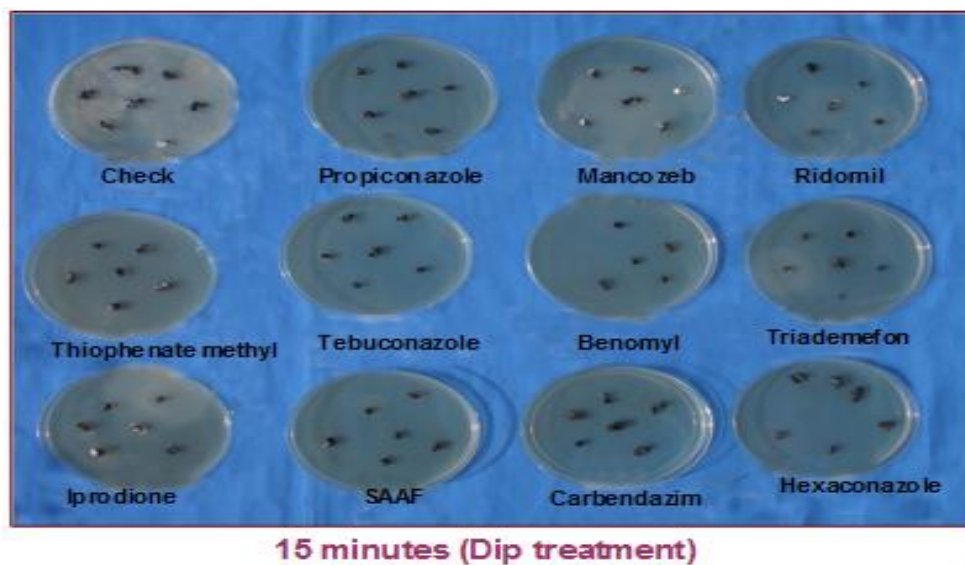


Plate 3: Effect of fungicides on sclerotial germination (15 minutes dip treatment)

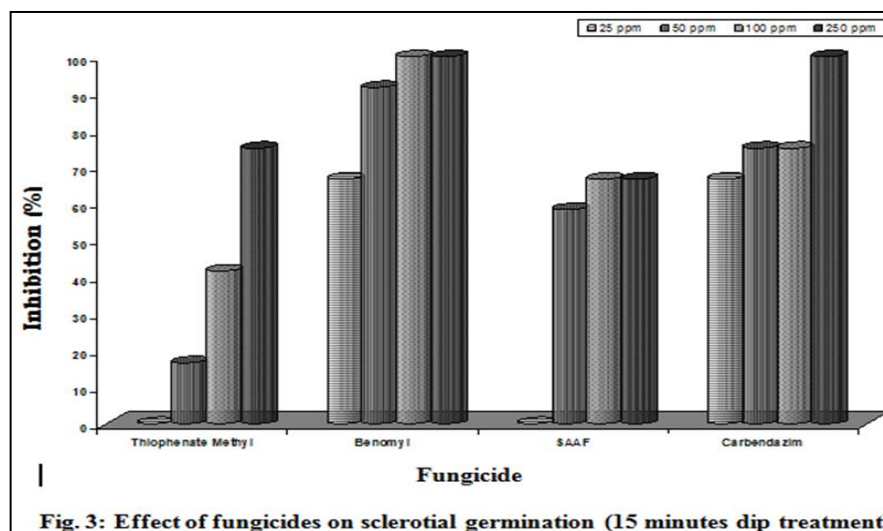


Fig. 3: Effect of fungicides on sclerotial germination (15 minutes dip treatment)

In the present study, complete inhibition of sclerotial germination was found with benomyl at 100 µg a. i./ml in both the treatments (5 and 15 minutes) sclerotial dip treatment. However, carbendazim at 250 µg a. i./ml completely inhibited the sclerotial germination only in 15 minutes dip treatment. The other fungicides were found ineffective in inhibiting sclerotial germination. Sharma and Basandrai (1997) [12] also reported carbendazim as most effective in inhibiting sclerotial germination (90%) at 100 ppm. However, Goswami *et al.* (2008) [6] observed 100 per cent inhibition of sclerotial germination with benomyl and carbendazim. In the present study, the increased efficacy of fungicides was observed when sclerotia were treated for 15 minutes. Thus it is possible to reduce sclerotia viability by judicious use of fungicides. Not much of the works has been done on sclerotial dip treatment against the pathogen.

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5. References

1. Anonymous. Annual progress report 2006-07. All India co-ordinated research project (Rapeseed-Mustard). GBPUAT, Pantnager, 2006, 127p.
2. Chattopadhyay C, Meena PD, Sudeer K. Management of *Sclerotinia* stem rot of mustard using eco-freindly strategies. *J. Mycol. and pl. pathol.* 2002; 32:194-200.
3. Chauhan LS, Singh J, Chandra DR. Assesment of losses due to stem rot to yellow sarson. *Proc. Natl Symp on Management of Microbes in Service of Mankind*, 19-21 Nov 1992. Allahabad, India, 1992, 65-66p.
4. Ghasolia RP, Shivpuri A, Bhargava AK. *Sclerotinia* rot of Indian mustard (*Brassica juncea*) in Rajasthan. *Indian Phytopath.* 2004; 57:76-79.
5. Goswami K, Tewari AK, Awasthi RP. *In vitro* screening of fungicides and bioagents against *Sclerotinia sclerotiorum* (stem rot of Rapeseed-mustard). In: *Proc IPS (MEZ) annual meet and Natl Symp on Advancing frontiers of plant disease management*, 15- 17 Nov 2007, NDUA&T, Faizabad, 2007, 93p (Abs).
6. Goswami K, Tewari AK, Awasthi RP. *In vitro* evaluation of fungicides and culture filtrate of *Trichoderma* isolates against sclerotia of *Sclerotinia sclerotiorum* causing stem rot of Rapeseed-mustard. *3rd J & K Congress on Science and Technology for Sustainable Development*, 26-28 Feb 2008 University of Jammu, Jammu, India, 2008, 299-300p (Abs).
7. Kang IS, Chahal SS. Prevalence and incidence of white rot of rapeseed and mustard incited by *Sclerotinia sclerotiorum* in Punjab. *Plant Dis. Res.* 2000; 15:232-233.
8. Krishnia SK, Meena PD, Chattopadhyay C. Seed- yield and yield-attributes of Indian mustard affected by *Sclerotinia* rot. *J. Mycol. Pl. Pathol.* 2000; 30:265.
9. Lodha BC, Bhatanager MK, Mathur K, Doshi A, Mathur S, Bairwa LN *et al.* *Plant Pathological thoughts and News.* Deptt. of Plant Pathology, Rajasthan Collage of Agric., Udaipur (India), 1992, 52p.
10. Mueller DS, Dorrance AE, Derksen RC, Ozlan E, Kurle JE. Efficacy of fungicides on *Sclerotinia sclerotiorum* and their potential for control of sclerotinia stem rot on soybean. *Plant Dis.* 2002; 86:26-31.
11. Roy AK, Saikia UN. White blight of mustard and its control. *Indian J. Agric. Sci.* 1976; 46:274-277.
12. Sharma BK, Basandrai AK. Effect of biocontrol agents, fungicides and plant extracts on sclerotial viability of *Sclerotinia sclerotiorum*. *Indian J. Agric. Sci.* 1997; 67:132-133.
13. Sharma CL, Kapoor AS. *In vitro* sensitivity of *Sclerotinia sclerotiorum* to fungicides. *Himanchal J. Agric. Res.* 1997; 23:50-55.
14. Schmitz H. A suggested toximetric method for wood preservations. *Inus and Engin. Chem. Anlyt. Ed. II.* 1930, 361-663.
15. Shivpuri A, Gupta RBL. Evaluation of different fungicides and plant extracts against *Sclerotinia sclerotiorum* causing stem rot of mustard. *Indian Phytopath.* 2001; 54:272-274.
16. Shivpuri A, Sharma KB, Chhipa HP. Some studies on the stem rot (*Sclerotinia sclerotiorum*) disease of rapeseed/ mustard in Rajasthan. *J. Mycol. Pl. Pathol.* 2000; 30:268.
17. Singh R, Tripathi NN, Kaushik CD. Management of *Sclerotinia* rot of Indian mustard (*Brassica juncea* (L.) Czern and Coss.) by fungicides. *Crop Res.* 1994; 7:276-281.
18. Singh C, Tiwari S, Boudh S, Singh JS. Biochar application in management of paddy crop production and methane mitigation. In: Singh, J.S., Seneviratne, G. (Eds.), *Agro-Environmental Sustainability: Managing Environmental Pollution*, second ed. Springer, Switzerland, 2017a, 123-146p.
19. Singh C, Tiwari S, Singh JS. Impact of Rice Husk Biochar on Nitrogen Mineralization and Methanotrophs Community Dynamics in Paddy Soil, *International Journal of Pure and Applied Bioscience.* 2017b; 5:428-435.
20. Singh C, Tiwari S, Singh JS. Application of Biochar in Soil Fertility and Environmental Management: A review, *Bulletin of Environment, Pharmacology and Life Sciences.* 2017c; 6:07-14
21. Singh C, Tiwari S, Gupta VK, Singh JS. The effect of rice husk biochar on soil nutrient status, microbial biomass and paddy productivity of nutrient poor agriculture soils *Catena.* 2018; 171:485-493.
22. Tiwari S, Singh C, Singh JS. Land use changes: a key ecological driver regulating methanotrophs abundance in upland soils. *Energy, Ecology, and the Environment.* 2018; 3:355-371.
23. Tiwari S, Singh C, Boudh S, Rai PK, Gupta VK, Singh JS. Land use change: A key ecological disturbance declines soil microbial biomass in dry tropical uplands. *Journal of Environmental Management.* 2019a; 242:1-10.
24. Tiwari S, Singh C, Singh JS. Wetlands: A Major Natural Source Responsible for Methane Emission A.K. Upadhyay *et al.* (Eds.), *Restoration of Wetland Ecosystem: A Trajectory towards a Sustainable Environment*, 2019b, 59-74p.
25. Kour D, Rana KL, Yadav N, Yadav AN, Rastegari AA, Singh C *et al.* *Technologies for Biofuel Production: Current Development, Challenges, and Future Prospects.* A. Rastegari *et al.* (Eds.), *Prospects of Renewable Bioprocessing in Future Energy Systems, Biofuel and Biorefinery Technologies.* 2019a; 10:1-50.
26. Singh C, Tiwari S, Singh JS. Biochar: A Sustainable Tool in Soil 2 Pollutant Bioremediation R. N. Bharagava, G. Saxena (Eds.), *Bioremediation of Industrial Waste for Environmental Safety*, 2019b, 475-494p.
27. Singh Y. Management of *Sclerotinia* rot of rapeseed and mustard through chemicals. *Plant Dis. Res.* 1998; 13:149-150.