

E-ISSN: 2278-4136 P-ISSN: 2349-8234 www.phytojournal.com JPP 2022; 11(1): 204-207 Received: 25-11-2021 Accepted: 27-12-2021

Anisa Qadir Janwari Ph.D. Scholar, Division of Veterinary Pathology, SKUAST-K, Jammu and Kashmir, India

Masood Saleem Mir Associate Director Extension, SKUAST-K, Jammu and Kashmir, India

### SA Kamil

H.O.D, Division of Veterinary Pathology, F.V.sc and A.H, SKUAST-K, Jammu and Kashmir, India

### Majid Shafi Kawoosa

Assistant Professor, Division of Veterinary Pathology, SKUAST-K, Jammu and Kashmir, India

Showkat Ah. Shah

Assistant Professor, Division of Veterinary Pathology, FVSc, SKUAST-K, Jammu and Kashmir, India

#### Showkat\_un-Nabi

Assistant Professor Division of Clinical Veterinary Medicine, Ethics and Jurisprudence, SKUAST-K, Jammu and Kashmir, India

Mudasir Ali Rather Assistant Professor, Division Veterinary Public Health. SKUAST-K, Kashmir, India

Zahid Amin Kashoo Assistant Professor, Division of Veterinary Microbiology. SKUAST-K, Jammu and Kashmir, India

### Shaheen Farooq

Assistant Professor division of Veterinary Microbiology, SKUAST-K, Jammu and Kashmir, India

Corresponding Author: Anisa Qadir Janwari Ph.D. Scholar, Division of Veterinary Pathology, SKUAST-K, Jammu and

Kashmir, India

Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



Journal of Pharmacognosy and

Phytochemistry

Anisa Qadir Janwari, Masood Saleem Mir, SA Kamil, Majid Shafi Kawoosa, Showkat Ah. Shah, Showkat\_un-Nabi, Mudasir Ali Rather, Zahid Amin Kashoo and Shaheen Farooq

### Abstract

The present experiment was conducted for 35 days, aimed to investigate the susceptibility of commercial Broiler and Vanraja chicken to Salmonella enterica subsp. enterica serovars Gallinarum infection with supplementation garlic. A total of 160 (80 vanraja and 80 broiler), day old chicks were procured from private local hatcheries (Srinagar). 20 birds each of both vanaraja and broiler were put into four cages and labelled as Control, Control Garlic, Infected and Infected Supplemented with Garlic. Infection was given to infected and infected supplemented with garlic given I/P with 0.2 ml suspension *Salmonella gallinarum* containing  $2.08 \times 10^9$  CFU. Clinical signs along with the mortality pattern was recorded in both vanraja and broiler chickens in all the four groups. Daily observations were made in all the four groups in both vanraja and broiler chickens. There was no clinical signs and mortality observed in both control and garlic control group. There was significant increase in mortality in broiler infected with *S. gallinarum* than in vanraja infected group, while as mortality was more in infection supplemented with garlic in broiler birds than in vanraja feeding to replace antibiotic as a growth promoter.

Keywords: broiler, vanraja, Allium sativum

## Introduction

Salmonellosis is a big socioeconomic threat worldwide that causes considerable mortality and morbidity in both humans and animals (Smith et al, 2016)<sup>[29]</sup> The gastrointestinal (GI) tract of newly hatched chickens is usually sterile and presents an empty ecological niche that provides easy access for the pathogen to colonize with limited restriction making them highly susceptible to enteric bacterial infections, such as Salmonella (Crhanova M. et al 2011)<sup>[4]</sup>. Among poultry diseases Salmonella is a leading cause of food borne illness worldwide with an estimated annual economic loss of 3.7 billion dollar (Hafiz et al. 2017)<sup>[13]</sup>. Although diseases due to this pathogen have been associated with a wide variety of food sources, poultry, in particular, have been regarded as the single largest cause of human salmonellosis (Gould et al 2013) <sup>[10]</sup>. Salmonellae infecting poultry are S. Gallinarum: major avian serotype in india (Prakash et al., 2005)<sup>[25]</sup>, S. pullorum, S. typhimurium, S. enterititidis. Fowl typhoid a septicemic disease caused by Salmonella enterica subsp. enterica serovar Gallinarum is widely distributed throughout the world, affects all age groups of chicken (Priyantha 2012)<sup>[26]</sup> It is a major avian serotype responsible for immense economic losses due to morbidity and mortality. Many synthetic drugs and growth promoters are supplemented to the broilers to effect rapid growth, but their use have shown many disadvantages like high cost, adverse side effect on health of birds and long residual properties etc. So, scientists are again concentrating on the use of our ancient medicinal system to find beneficial herbs and plants, which can be safely used to increase the production and control various disease. Herbal medicines have always been a form of therapy for livestock among poor small holder farmers. There is, however, little documentation of the use of ethno veterinary medicines, as many researchers and health practitioners view these practices as backward Documentation of herbal plants is necessary because they are likely to be more important in the future, especially given the escalating costs of drugs and the focus on organic products in most developing countries. One of the very important herbs used Spices like Garlic (Allium sativum) are well known for their dietary and medicinal value. The precursor of allicin, which is hydrolysed by enzyme allinase after crushing to its active form, the allicin (S-allyl-2-propenthiosulphinate).

Allicin the most potentially active component of garlic is responsible for its biological properties (Heinrich et al., 2004) <sup>[14]</sup>. Freshly crushed garlic contains allicin, alliin, ajoene, diallylsulfide, dithiin, S-allylcysteine. Garlic (Allium sativum) has been reported to possess useful pharmacological substances (Akhtar et al 1984) <sup>[1]</sup>. Garlic as natural feed additive in poultry nutrition may be of great benefit and value especially for broiler growers. This is due to its antibacterial, anti-parasitic anti-inflammatory, antiseptic, and immunomodulatory properties of garlic. The flavonoids present in Allium sativum exhibits bacteriostatic and bacteriocidal effects on some strains of bacteria by inhibiting the reverse transcriptase and proteases activity in bacteria. (Gazuwa et al; 2003)<sup>[9]</sup>.

# Materials and Methods

A total, 120 (80 commercial broilers and 80 vanraja birds were equally divided ie 20 birds each into four groups, control, garlic control, Infection and Infection supplemeted with garlic @ 176 mg /Kg b/w in their feed. The chicks were reared on cage system under standard management practice though out the experimental period of 35 days. Starter feed was given upto two weeks and finisher ration was fed ad libtum from third to fifth week to both control and treatment groups. Birds were checked for clinical signs and mortality from 0 days post infection during 5 weeks of experimental period. The collected data was stastically analyzed as per procedure given by Snedecor and Cochran (1967).

# **Results and Discussion**

The observations for clinical signs in both vanraja and broiler birds in all four groups, Control, Garlic, Infection and infection supplemented with garlic were noted. No clinical signs were observed in the Vanraja or Broiler chickens from control and garlic throughout the course of study.

In Vanraja birds, clinical signs in both infection and garlic supplemented infection groups appeared from day-1 PI. The birds appeared dull and depressed, and huddled along the front wall of the cage. In the infection group, 13 (65%) birds reluctant to move and stood with sunken head with ruffled feathers. The severity appeared to increase on day-2 and 3 PI. Loss of appetite was evident in all the birds but majority started feeding from day-5PI. However, from day-5 PI and during 2<sup>nd</sup> week PI majority of the birds exhibited increased thirst, and typical loose greenish- yellow diarrhoea. Recovery was evident from day-14 onwards. Birds dead during the study period showed severely ruffled feathers and drooping of wings, before death. Similar changes were observed in birds from garlic supplemented infection group but the signs were comparatively less severe on day-1. Moderate to severe dullness and depression were observed in 9 (45%) birds which exhibited huddling, where as others exhibited milder symptoms and infrequently moved away from other birds. Increase in water intake was noticed in all birds during the second week. Greenish diarrhoea was observed in only few birds. Recovery was markedly evident after 2<sup>nd</sup> week PI.

The clinical signs in this infection and garlic supplemented infection groups of broiler birds were seen within 12 to 15 hours following infection. In the infection group, all the birds appeared dull, depressed, and huddled together. Moderate to severe depression with ruffled feathers, drooping of wings and reluctance to move or feed was observed in 11 (55%) birds. From day-3 PI, 13 (65%) birds exhibited increased thirst and greenish-yellow diarrhoea. Although the severity of disease decreased progressively especially from day-14 PI,

but some birds continued to show milder symptoms till day-21 PI. Birds dead early during the course of study showed severe depression, stood with sunken head and closed eyes with markedly ruffled feathers and drooping wings and did not move even when forced to do so. Nature and progression of the disease in garlic supplemented infection group was comparable to that in infection group with all the birds exhibiting dullness, depression, fever and huddling together. On day-2 PI, 7 birds showed severe symptoms with ruffling of feathers, drooping of wings and reluctance to move or feed. From day-4 to 14 PI, 14 (70%) birds exhibited increased thirst and greenish-yellow diarrhoea. Recovery was evident after day-14 PI. The birds dead during the course of study exhibited severe depression, complete inappetance, reluctance to move with sunken head and closed eyes, droopy wings, and ruffled feathers before death.

Comparatively analysis of Vanraja and broiler chickens revealed that the nature and progression of clinical disease was similar in both types of birds. However, the symptoms appeared earlier and were comparatively more severe in broiler chickens. Also, relative percentage of birds showing more severe symptoms was more in broilers. Also recovery started comparatively earlier in Vanraja birds.

# Mortality

The number of birds that died in different experimental groups at various intervals of infection is given in Table-2 and graphically depicted in Fig. 1 and 2.

A perusal of this table reveals that no mortality was seen in uninfected birds of control and garlic groups from either Vanraja or broiler chickens.

Total mortality observed, during the period of study, in Vanraja birds was 25% (5/20) in the infection group and 20% (4/20) in garlic supplemented infection group with one bird each dead on day-1, 3, 5, 12 and 14, and day-2, 5, 6 and 16 in the two groups, respectively. Although, mortality was first recorded in Infection group, maximum mortality (3 in each group) in both the groups was noticed during first week following infection as shown in (Fig 1,2)

Total mortality observed, during the period of study, in broiler chickens was 35% (7/20) in the infection group and 25% (5/20) in garlic supplemented infection group. Mortality in both the groups started on day-1 PI. Maximum mortality in the infection group was noticed on days-2 and 3, with 2 birds dead on either day. Also, out of a total of 7 mortalities, 5 died during the first week following infection. In garlic supplemented group 3 out of a total of 5 mortalities were noticed during first week following infection where as two birds died during the  $3^{rd}$  week post infection (day-16 PI) as shown in Fig (1 and 2)

Comparative analysis of the mortality pattern in Vanraja and broiler chickens revealed that mortality in general was lower in Vanraja chickens. However, maximum mortality in both Vanraja and broiler chickens was observed during first week post infection. The overall ratio of mortalities in Vanraja vs broiler chickens was 5:7 in infection group and 4:5 in garlic supplemented infection group. The ratio of mortalities during first week was 3:5 and 1:1 in infection and garlic supplemented infection groups, respectively. In present study both Vanaraja and Broiler chickens were found susceptible to *Salmonella gallinarum* infection, and the nature and progression of clinical disease was similar in both types of chickens. Salmonellosis is one of the important disease problems in poultry affecting commercial broiler and layer chicken globally. *S.* Gallinarum causes systemic disease, fowl typhoid, affecting all age groups causing heavy morbidity and mortality (Geetha and Palanivel, 2018; Sreekantapuram et al., 2021) <sup>[11, 31]</sup>. Varying prevalence rates have been reported among broiler farms (Arora et al., 2015; El-Sharkawy et al., 2017; Jibril et al., 2020) [2, 7, 17]. High rearing density and production stress has been frequently incriminated as a cause for higher susceptibility of the commercial chicken (Machado Junior et al., 2020) <sup>[22]</sup>. In contrast, the native chicken traditionally reared in backyards or range are continuously exposed to pathogens and ambient conditions (Ayala et al., 2020) [3], but are considered more resistant to infection or development of more severe clinical disease (Kannaki et al., 2021) <sup>[19]</sup>. Vanaraja are the improved chickens for backyards. Although well adapted to backyard (Singh et al., 2019)<sup>[28]</sup>, Salmonella gallinarum infection has been reported under natural conditions (Dey et al., 2016; Pal et al., 2017) [6, 24]. However, during current study, the symptoms appeared earlier and were comparatively more severe in broiler chickens. Also, the relative percentage of birds showing more severe symptoms was more in broilers as well as recovery started comparatively earlier in Vanaraja birds. Vanaraja birds have been observed to exhibit better resistance against Escherchia coli infection, and a more potent humoral and cell mediated immune response when compared with broiler chicken. However, the resistance has been found to be lower than native breeds (Reddy et al., 2002; De et al., 2013) [27, 5]. Resistance to disease in chicken has been genetically mapped to different quantitative trait loci (QTLs) which interact with environment determines immunopathological response to infections (Jie and Liu, 2011;). Differential expression of these genes in different chicken breeds/ types have been associated with the varied response to Salmonella infection (Hu et al., 2016).

Maximum mortality in both Vanaraja and broiler chickens was observed during first week

gallinarum causes post infection. Salmonella acute septicaemic disease with mortality ranging from 10% to 90%. Maximum mortality due to Salmonella gallinarum has been observed during 1-2 week aged birds (Kumari et al., 2013) <sup>[21]</sup>. Salmonella being an intracellular pathogen with the virulence genes facilitating its adaptation and growth in modified endosomes of macrophages. The activation of phagocytes for bactericidal activity is IFN-y dependent. Development of Salmonella-specific Th1 cells capable of producing sufficient IFN-y for controlling intracellular bacterial replication takes several weeks leading to heavy mortality during early period following infection (Griffin et al., 2009; Jackson et al., 2010) <sup>[12, 16]</sup>. The overall ratio of mortalities in Vanaraja vs broiler chickens was 5:7 in infection group and 4:5 in garlic supplemented infection group. The ratio of mortalities during first week was 3:5 and 1:1 in infection and garlic supplemented infection groups, respectively. The study reflects that Vanaraja chicken are comparatively more tolerant to Salmonella gallinarum infection than commericial broiler but there was no marked difference in susceptibility. Similar observations have been reported for local vs commercial layers as well as among various local chicken ecotypes (Weerasooriya et al., 2017) <sup>[32]</sup>. Garlic supplementation could not prevent the disease but the overall severity of the disease as well as acute mortality was considerably reduced. Garlic has been shown to have beneficial effects against different infections in chicken and other animals with a significant impact on reducing mortality and facilitating recovery (Gautam et al. 2017; Navidshad et al. 2018; Kumar et al., 2022) [8, 23, 20].



Fig 1: Number of birds dead in the different groups of Vanraja and Broiler chickens following *Salmonella enterica subsp. enterica* serovar Gallinarum infection



Fig 2: Per cent Mortality in different groups of Vanraja and Broiler chickens following Salmonella enterica subsp. enterica serovar Gallinarum infection

## Reference

- 1. Akhtar MS, Afzal H, Chaudhary F. Preliminary in vitro antibacterial screening of Bakain, Gilo and Zarisk against Salmonella. Medicos. 1984;9:6-7.
- Arora D, Kumar S, Jindal Naresh, Narang G, Kapoor PK, Mahajan NK. Prevalence and epidemiology of Salmonella enterica serovar Gallinarum from poultry in some parts of Haryana, India. Veterinary World. 2015;8(11):1300-1304.
- Ayala AJ, Yabsley MJ, Hernandez SM. A Review of Pathogen Transmission at the Backyard Chicken–Wild Bird Interface. Frontiers of Veterinary Science. 2020;7:539925.
- 4. Crhanova M, Hradecka M, Faldynova HM, Matulova M, Havlickova H, Sisak F *et al.* Immune response of chicken gut to natural colonization by gut microflora and to *Salmonella enterica serovar Enteritidis* infection. Infection and Immunity. 2011;79:2755-2763.
- 5. De AK, Kundu A, Vasantha RV, Kundu MS, Jeyakumar S, Sunder J. Antibody response to goat erythrocytes in endangered Nicobari fowl, Vanaraja and their various F1 and F2 crosses under the hot humid climate of Andaman and Nicobar Islands, Indian Journal of Applied Animal Research. 2013;41(2):125-132.
- 6. Dey S, Mahanti A, Batabyal K, Joardar SN, Samanta I, Isore DP, Pakhira MC. Identification and antimicrobial susceptibility of *Salmonella gallinarum* isolated from Fowl Typhoid outbreak in backyard Vanaraja fowl.

Exploratory Animal and Medical Research. 2016;6(1):63-67.

- 7. El-Sharkawy H, Amin T, Abd El-Galiel A, El-Gohary, Moshira E, Fares E, *et al.* Epidemiological, molecular characterization and antibiotic resistance of Salmonella enterica serovars isolated from chicken farms in Egypt. Gut Pathogens. 2017;9:8.
- Gautam G, Shrestha N, Bhandari S. Effect of *Allium* sativum on Immune Status Against Newcastle Disease Virus and Productive Performance of Broiler Chicken. International Journal of Poultry Science, 2017;16:515-521. DOI: 10.3923/ijps.2017.515.521
- 9. Gazuwa SY, Makanjuola ER, Jaryum KH, Kutshik JR, Mafulul SG. The Phytochemical composition of Allium cepa/Allium sativum and the effects of their aqueous extracts (cooked and raw forms) on the lipid profile and other hepatic biochemical parameters in female albino wistar rats, Asian Journal of Biological Sciences. 2013;4(3):406-410.
- Gould LH, Walsh KA, Vieira AR, Herman K, Williams IT, Hall AJ. Surveillance for foodborne disease outbreaks - United States, 1998–2008. MMWR Morbidity Mortality Weekly Report. 2013;62:1-34.
- 11. Geetha M, Palanivel KM. A Brief Review on Salmonellosis in Poultry. International Journal of Current Microbiology and Applied. 2018;7(05):1269-1274.
- 12. Griffin A, Baraho-Hassan D, McSorley SJ. Successful treatment of bacterial infection hinders development of acquired immunity. Journal of immunology. 2009;183:1263-1270.
- 13. Hafiz N, Nadarajan A, Ahamed K, Shamila S, Li-oon C, Huda Nurul, *et al.* Prevalence of salmonella in poultry processing environments in wet markets in penang and perlis, Malaysia. Veterinary World. 2017;10(3):286-292.
- Heinrich M, Barnes J, Gibbsons S, Williamson ME. Fundamentals of Pharmacology and Phytotherapy-1<sup>st</sup> Ed., Churchil Livingstone, U.K. 2004;(32, 33):220-221.
- 15. Anil Kumar Jha. Histopathological studies of tapeworm Raillietina tetragona (Molin, 1858) from the gastrointestine of indigenous chicken (Gallus domesticus L.) farming in Kirtipur, Nepal. Int J Vet Sci Anim Husbandry 2019;4(4):01-06.
- 16. Jackson A, Nanton MR, O'Donnell H, Akue AD, McSorley SJ. Innate immune activation during *Salmonella* infection initiates extramedullary erythropoiesis and splenomegaly. Journal of immunology. 2010;185(10):6198-6204.
- 17. Jibril AH, Okeke IN, Dalsgaard A, Kudirkiene E, Akinlabi OC, Bello MB. Prevalence and risk factors of *Salmonella* in commercial poultry farms in Nigeria. PLoS ONE. 2020, 15(9).
- Jie H, Liu YP. Breeding for disease resistance in poultry: Opportunities with challenges. World's Poultry Science Journal, 2011, 67.
- 19. Kannaki TR, Priyanka E, Santosh H. Disease tolerance/resistance and host immune response to experimental infection with *Pasteurella multocida* A:1 isolate in Indian native Nicobari chicken breed. Poultry Science. 2021;100(8):101268
- 20. Kumar A, Sharma NK, Kheraviia SK, Keerqina C, Ionescub C, Blanchard A *et al.* Potential of a mixture of eugenol and garlic tincture to improve performance and intestinal health in broilers under necrotic enteritis challenge. Animal Nutrition. 2022;8(1):26-37.

- 21. Kumari D, Mishra SK, Lather D. Pathomicrobial studies on *Salmonella gallinarum* infection in broiler chickens, Veterinary World. 2013;6(10):725-729.
- 22. Machado J, Chung PC, Hagerman A. Modeling *Salmonella* Spread in Broiler Production: Identifying Determinants and Control Strategies. Frontiers of Veterinary Science. 2020;7:564.
- Navidshad B, Darabighane B, Malecky M. Garlic: An alternative to antibiotics in poultry production, a review. Iranian Journal of Applied Animal Science. 2018;8(1):9-17.
- 24. Pal S, Dey S, Batabyal K, Banerjee A, Narayan JS, Samanta I, *et al.* Characterization of *Salmonella gallinarum* isolates from backyard poultry by polymerase chain reaction detection of invasion (inv A) and Salmonella plasmid virulence (spv C) genes. Veterinary World, 2017;10(7):814-817.
- 25. Prakash T, Suryanarayana LM, Krishnapp G. Evaluation of *Salmonella gallinarum* outer membrane protein based enzyme linked immunosorbent assay for detecting antibodies in vaccinated and infected Chicken. International Journal of Poultry Science. 2005;4(4):222-227.
- 26. Priyantha MAR. An Overview: Vaccination to Kidneys: control fowl typhoid in Commercial layers, Sri Lanka. Grossly, kidneys were enlarged with Wayamba. Journal of Animal Science. 2012, 23-25.
- 27. Reddy MR, Arun Panda, Praharaj NK, Rama Rao SV, Chaudhuri D, Sharma RP. Comparative evaluation of immune competence and disease resistance in dual purpose chicken Vanaraja and Gramapriya vis-à-vis coloured synthetic broiler. The Indian journal of animal sciences. 2002;72(1):6-8.
- Singh M, Islam R, Avasthe R. Socioeconomic Impact of Vanaraja Backyard Poultry Farming in Sikkim Himalayas. International Journal of Livestock. 2019;9(3):243-248.
- 29. Smith SI, Seriki A, Ajayi A. Typhoidal and nontyphoidal *Salmonella* infections in Africa. European Journal of Clinical Microbiology of Infectious Disease. 2016;35:1913-2240.
- Snedecor GW, Cochran WG. Statistical methods (6<sup>th</sup> edition), Lowa State University. Press, Lowa U.S.A. 1994.
- 31. Sreekantapuram S, Berens C, Barth AS, Methner U, Berndt A, Interaction of *Salmonella gallinarum* and Salmonella Enteritidis with peripheral leucocytes of hens with different laying performance. Veterinary Research. 2021;52:123.
- 32. Weerasooriya KMSG, Fernando PS, Liyanagunawardena, N, Wijewardena G, Wijemuni MI, Samarakoon SATC. Natural resistance of Sri Lankan village chicken to *Salmonella gallinarum* infection, British Poultry Science, 2017. DOI: 10.1080/00071668.2017.1376034.
- 33. Hu Y, Chen WW, Liu HX, Shan YJ, Zhu CH, Li HF. *Et al.* Genetic differences in ChTLR15 gene polymorphism and expression involved in *Salmonella enterica* natural and artificial infection respectively of Chinese native chicken breeds, with a focus on sexual dimorphism. Avian Pathology. 2016;45:13-25.