

# Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



### E-ISSN: 2278-4136 P-ISSN: 2349-8234 www.phytojournal.com

JPP 2022; 11(6): 42-45 Received: 16-07-2022 Accepted: 18-08-2022

#### Kouame Bi Koffi François Prévost

- LCOSN, UFR SSMT, 08 BP 582 Abidjan 08, Université F. H. B. Abidjan, Côte d'Ivoire
- EBSI Group, CEISAM, LUNAM-CNRS UMR6230, BP 92208, 44322 Nantes, France
- 3. Département de Mathématiques, Physique, Chimie, UFR des Sciences Biologiques, Université Peleforo. GON COULIBALY. Korhogo, BP 1328 Korhogo, Côte d'Ivoire

#### Quattara Zana

- Laboratoire de Chimie BioOrganique et de Substances Naturelles, Université Nangui Abrogoua, Abidjan, Côte d'Ivoire de Chimio Organique Bidgeigue.
- de Chimie Organique Biologique

  2 Université de Corse-Centre National de la
  Recherche Scientifique, Unité Mixte de
  Recherche 6134, Equipe Chimie et
  Biomasse, Ajaccio, France

#### Sari Sari Chardin

UCOSN, UFR SSMT, 08 BP 582 Abidjan 08, Université F. H. B. Abidjan, Côte

#### Gooré Stéphane Gervais

- Laboratoire de Chimie BioOrganique et de Substances Naturelles, Université Nangui Abrogoua, Abidjan, Côte d'Ivoire de Chimie Organique Biologique
- <sup>2</sup> Université de Corse-Centre National de la Recherche Scientifique, Unité Mixte de Recherche 6134, Equipe Chimie et Biomasse, Ajaccio, France

#### Kablan Landry

- LCOSN, UFR SSMT, 08 BP 582 Abidjan 08, Université F. H. B. Abidjan, Côte d'Ivoire
- Département de Mathématiques, Physique, Chimie, UFR des Sciences Biologiques, Université Peleforo. GON COULIBALY. Korhogo, BP 1328 Korhogo, Côte d'Ivoire

#### Redi Gustave

LCOSN, UFR SSMT, 08 BP 582 Abidjan 08, Université F. H. B. Abidjan, Côte d'Ivoire

#### Tea Illa

EBSI Group, CEISAM, LUNAM-CNRS UMR6230, BP 92208, 44322 Nantes, France

#### Tonzibo Feli:

LCOSN, UFR SSMT, 08 BP 582 Abidjan 08, Université F. H. B. Abidjan, Côte d'Ivoire

#### Corresponding Author:

Kouame Bi Koffi François Prévost

- LCOSN, UFR SSMT, 08 BP 582 Abidjan 08, Université F. H. B. Abidjan, Côte d'Ivoire
- EBSI Group, CEISAM, LUNAM-CNRS UMR6230, BP 92208, 44322 Nantes, France
- 3. Département de Mathématiques, Physique, Chimie, UFR des Sciences Biologiques, Université Peleforo. GON COULIBALY. Korhogo, BP 1328 Korhogo, Côte d'Ivoire

# Chemical constituents of essential oils from flowers and stems of an Ivorian species of the genus Crassocephalum

Kouame Bi Koffi François Prévost, Ouattara Zana, Seri Seri Chardin, Gooré Stéphane Gervais, Kablan Landry, Bedi Gustave, Tea Illa and Tonzibo Felix

**DOI:** https://doi.org/10.22271/phyto.2022.v11.i6a.14523

#### Abstract

Essential oils from fresh flowers and stems of *Crassocephalum sp* (Asteraceae) growing wild in Côte d'Ivoire were extracted by hydrodistillation using a Clevenger-type apparatus. Essential oils were analyzed by Gas Chromatography (GC) and then by GC coupled with a Mass Spectrometry (GC-MS). The analyzes of the oils allowed the identification and quantification of 39 and 83 constituents respectively in the oils of the flowers and the stems. The characterization of the oils revealed the predominance of the hydrocarbon monoterpene myrcene with 87.25% in the flowers, 35.55% in the stems. Myrcene alone accounts for almost all of the flower oil. The chemotype of the stem oils is different from that of the flowers with the presence of terpinolene (31.99%) and (*E*)-dec-2-enal (8.01%) which are other major compounds in the oils of the stems of *Crassocephalum sp*.

Keywords: Crassocephalum, chemotype, essential oils, phytochemistry, myrcene, terpinolene

#### Introduction

The genus Crassocephalum to which the species Crassocephalum sp belongs, is part of the very large and widespread family Asteraceae in the tribe Senicioneae. The genus Crassocephalum has been reported to include nearly 24 species (Wagner et al., 1999) [10] some of which have medicinal properties. Several species of Crassocephalum are used in traditional medicine for their therapeutic virtue. Crassocephalum bauchiense Huch is widely used in Cameroonian traditional medicine. The leaf extract has been used to treat several illnesses, including epilepsy, arthritis, intestinal pain and colic (Arbonnier, 2009) [2]. A decoction of the leaves of this herb has been reported to be useful in relieving bronchitis and accompanying fever. According to traditional Cameroonian healers, the plant is also effective in cases of brain deficit, behavioral disorders in mentally retarded children, inflammatory disorders and neuropathic pain (Taïwe et al., 2012) [9]. Among the species of the genus Crassocephalum, several studies on the chemical composition of essential oils have been carried out for the species Crassocephalum crepidiodes. In Vietnam, the main components of essential oils extracted from Crassocephalum crepidioides flowers are myrcene (43.3%),  $\beta$ -phellandrene (10.7%) and cryptone (8.1%) (Hung et al., 2019) [3]. The chemical composition of the essential oils of the stems differs slightly with those of the flowers. Stem oils have as predominant compounds myrcene (26.1%),  $\alpha$ -pinene (10.7%),  $\alpha$ -humulene (5.9%) and (E)- $\beta$ -farnesene (5.2%) (Hung et al., 2019) [3]. On the other hand, the essential oils of the flowers and aerial parts (leaves + stems) of Crassocephalum crepidiodes. harvested in western India (Joshi, 2011) [5] had chemical compositions close to those of Vietnam. The essential oils extracted from the flowers of India contain 46.1% myrcene and 31.0%  $\beta$ -phellandrene and the essential oils from the aerial parts contain 45.3% of myrcene and 20.2% of  $\beta$ -phellandrene (Joshi, 2011) [5]. Cryptone, which was relatively abundant in essential oils from Vietnam, was only observed in small proportions (0.1%) in oils from India. In Africa, essential oils from the leaves of C. crepidioides from Cameroon had  $\alpha$ -phellandrene, p-cymene, myrcene, limonene and (E)- $\beta$ ocimene as major constituents (Zollo et al., 2000) [11]. In another report from Nigeria, the important constituents of oils from the leaves of the same plant were  $\alpha$ -caryophyllene,  $\beta$ cubebene and α-farnesene; while thymol, α-caryophyllene and 4-cyclohexybutyramide were the main constituents of stem oil (Owokotomo et al., 2012) [7]. It should be noted the absence of studies on the essential oils extracted from the flowers and stems of a species of the genus

Crassocephalum in Africa. It is important to note that no chemical study has been carried out on the volatile secondary metabolites (essential oils) of the flowers and stems of a species of the genus Crassocephalum in Côte d'Ivoire. Our research therefore presents for the first time, in Côte d'Ivoire, the chemical composition of essential oils extracted from the flowers and stems of a plant of the genus Crassocephalum, the species *Crassocephalum sp* using a combination of three GC and GC/MS techniques.

# Materials and Methods Plant Materials

Flowers and stems of *Crassocephalum sp* were collected at 8 am in the fallow lands of the village of Gabia in the Oumé sub-prefecture, Gôh region (Centre-West of Côte d'Ivoire). Plant materials were identified in the National Floristic Center of University of Felix Houphouët Boigny, Cocody-Abidjan, department of Botany, Côte d'Ivoire.

#### **Isolation of Essential Oil**

Extraction of essential oils was carried out by steam distillation using a Clevenger-type apparatus. Fresh Flowers or stems (1000 g) were placed on a metal grid under which is distilled water (3000 mL) boiling for 4 hours. Oils are driven by steam. After condensation and liquefaction, the oil overcomes the water in the test tube. The obtained oil was dried over anhydrous sodium sulphate and after filtration, stored in a sealed sample tube at 0 °C until GC and GC-MS analysis.

### Chemical Analyses of the essential oil

The essential oil was investigated first by Gas Chromatography (GC) and then by GC coupled with a Mass Spectrometry (GC-MS). Concerning GC analysis, the gas phase chromatography was carried out using a Delsi DI 200 instrument equipped with a flame ionization detector and a DB5 column (25 m x 0.25 mm, df: 0.25 µm) with a split flow rate of 60 mL/min. Nitrogen was used as carrier gas; temperature programming was 5min at 50 °C and 30 °C/min up to 220 °C, injector and detector temperatures were respectively set to 220 °C and 250 °C. GC-MS analysis were performed using a Hewlett-Packard gas Chromatograph Model 6890 coupled to a Hewlett-Packard MS Model 6890 equipped with an HP5 column (30 m x 0.25 mm, df: 0.25 μm). Initial oven temperature was maintained at 50 °C for 5 min and then programmed at 50 °C/min to 300 °C (held 50 min). The carrier gas was helium (1.0 mL/min); a split injection with a split ratio 1:10 was chosen. Injector and detector temperatures were respectively set to 250 °C and 320 °C. The electron multiplier was set at 2200 V with an applied electron ionization voltage of 70 eV, with the ion source temperature at 230 °C. Mass spectral data were acquired in the scan mode in the m/z range of 33-450. Identification of compounds was carried out by calculating Retention Indices (RI) or Koväts Indices (KI) and comparing mass spectra with those in data banks, i.e. Adams (Adams, 1995) [1] or Mc Lafferty and Stauffer (Mc Lafferty et al., 1989) [6]. For quantification purpose, relative area percentages obtained by field ionization detection (FID) were used.

## **Results and Discussions**

The essential oils obtained by hydrodistillation of the flowers and stems of Crassocephalum sp from Côte d'Ivoire are yellow in color. The identification and quantification by GC and GC-MS techniques of the essential oils of the flowers of *Crassocephalum sp*, gave a total of 39 major components

representing 99.72 Essential oils of the stems have been identified 83 constituents representing 99.19% (Table 1). Monoterpenes (98.43% in the leaves, 74.58% in the stems) dominate by their presence the chemical composition of the essential oils extracted from the flowers and stems of Crassocephalum sp from Côte d'Ivoire. The hydrocarbon monoterpene, myrcene (87.25%), is the predominant compound which alone accounts for almost all of the flower oil. Limonene (4.27%) and  $\beta$ -pinene (3.80%), which are also hydrocarbon monoterpenes, are relatively important in proportion compared to the rest of the constituents. Myrcene, limonene and β-pinene together represent 95.32% of the total oil composition of the flowers. Essential oils extracted from Crassocephalum sp flowers from Côte d'Ivoire indicate a monoterpene hydrocarbon chemotype. Another chemotype was obtained for the essential oils extracted from the stems. The main components are myrcene (35.55%), terpinolene (31.99%), (E)-dec-2-enal (8.01%),  $\beta$ -bourbonene (3.28%) and limonene (2.23%). Terpinolene, in small quantity in the flowers 0.83%, is found with a very high content in the stems. The chemical compositions of essential oils from flowers and stems of the species Crassocephalum crepidioides collected in Vietnam (Hung *et al.*, 2019) <sup>[3]</sup>, India (Joshi, 2011) <sup>[5]</sup>, Cameroon (Zollo *et al.*, 2000) <sup>[11]</sup> and in Nigeria (Owokotomo et al., 2012) [7] are different from those obtained in our study on flowers and stems of Crassocephalum sp from Côte d'Ivoire. In Vietnam (Hung et al., 2019) [3], oils extracted from flowers and stems also contain myrcene as the predominant compound but with different proportions. Myrcene content in oils obtained from flowers and stems of Crassocephalum sp from Côte d'Ivoire are higher than those from flowers and stems from Vietnam. In India, the proportion of myrcene in the aerial part (leaves + stems) is greater than that of the stems of Crassocephalum sp in Côte d'Ivoire. The other major compounds reported in Vietnam and India are different from those of the plant in our study. In addition, the compounds in significant amounts in Crassocephalum crepidioides oils from Vietnam and India,  $\beta$ -phelladrene and cryptone were not identified in our study. The results obtained in our work on the flowers and stems of the species Crassocephalum sp from Côte d'Ivoire show different chemotypes to those reported for the oils extracted from the leaves and stems of Crassocephalum crepidoides from Nigeria, a country of the West Africa. The thymol (43.93%),  $\alpha$ -caryophyllene (15.16%) and 4-cyclohexybutyramide (20.94%) chemotype reported for essential oils from the stems of Crassocephalum crepidoides from Nigeria (Owokotomo et al., 2012) [7] presents majority compounds which are in the minority or absent in the oils from the stems of the species studied, of the genus Crassocephalum. In Cameroon, the reported αphellandrene, p-cymene, myrcene, limonene and (E)- $\beta$ ocimene chemotype for seven species of Crassocephalum (Zollo et al., 2000) [11] is different from the myrcene chemotype obtained in the essential oils of the flowers and of the myrcene, terpinolene, (E)-dec-2-enal chemotype identified in the essential oils of the stems of the species Crassocephalum sp studied. Biological activity studies on several monoterpenes have demonstrated that myrcene has antibacterial properties (Rasoul et al., 2012) [8]. In addition, Inoue et al., (2004) [4] also showed that myrcene contributes to the antibacterial activity of essential oils of *Melaleuca* alternifolia (Inoue et al., 2004) [4]. Myrcene, which is the major constituent of the essential oils of the flowers and stems of the Crassocephalum sp plant from Côte d'Ivoire, could transmit its antibacterial properties to the oils of the flowers and stems of the species Crassocephalum sp.

 Table 1: Chemical constituents of essential oil of leaves and stems of Crassocephalum sp

				Flower			
No	KI	RI	Compounds	Conte	nts%	Formula	
1	885	886	styrene	0.02	0.18	C <sub>8</sub> H <sub>8</sub>	
2	909	900	2-heptanol	nd	0.10	C7H16O	
3	922	920	tricyclene	0.02	0.05	$C_{10}H_{16}$	
4	926	929	α-Pinene	0.36	0.05	$C_{10}H_{16}$	
5	945	944	camphene	0.01	nd	$C_{10}H_{16}$	
6	966	971	sabinene	0.39	0.10	$C_{10}H_{16}$	
7	978	976	β-Pinene	3.80	0.79	$C_{10}H_{16}$	
8	983	982	myrcene	87.25	35.55	$C_{10}H_{16}$	
9	987	988	Yomogi alcohol	0.02	nd	$C_{10}H_{18}O$	
10	996	996	2,2,4,6,6-pentamethylheptane	nd	0.20	$C_{12}H_{26}$	
		1005	$\alpha$ -phellandrene	0.04	0.01	$C_{10}H_{16}$	
	1018		α-Terpinene	0.03	0.12	C <sub>10</sub> H <sub>16</sub>	
	1025		paracymene	0.04	1.31	$C_{10}H_{16}$	
	1022		limonene	4.27	2.23	$C_{10}H_{16}$	
	1035		(Z)-β-ocimene	0.02	0.03	$C_{10}H_{16}$	
	1047		(E)-β-ocimene	1.01	0.48	$C_{10}H_{16}$	
	1060		γ-Terpinene	0.07	1.15	$C_{10}H_{16}$	
	1074		p-cymenene	nd	0.22	$C_{10}H_{16}$	
	1081		linalool	0.01	0.05	$C_{10}H_{18}O$	
	1083		trans-linalool oxide	0.01	0.08	$C_{10}H_{18}O_2$	
	1089		terpinolene	0.83	31.99	$C_{10}H_{16}$	
	1095		α-Pinene oxide	0.04	0.04	C <sub>10</sub> H <sub>16</sub> O	
	1113		(E)-4,8-dimethyl-nona-1,3,7-triene	nd	0.04	$C_{11}H_{18}$	
	1163		Terpinen-4-ol	0.04	0.22	$C_{10}H_{18}O$	
	1167		1,4-Dimethoxybenzene	nd	0.06	$C_6H_4(OCH_3)_2$	
	1180		α-Terpineol	0.01	0.03	$C_{10}H_{18}O$	
	1186		decanal	0.16	0.16	$C_{10}H_{20}O$	
	1213		Cumin aldehyde	nd	0.13	C10H <sub>12</sub> O	
	1237		chavicol	nd	0.02	C9H10O	
	1272		(E)-dec-2-enal	0.44	8.01	$C_{10}H_{18}O$	
	1283		carvacrol	nd	0.29	C <sub>10</sub> H <sub>14</sub> O	
	1286		bornyl acetate	nd	1.29	$C_{12}H_{20}O_2$	
	1290		Thymol	nd	0.33	C <sub>10</sub> H <sub>14</sub> O	
	1294		nonyl acetate	nd	0.09	C <sub>11</sub> H <sub>22</sub> O <sub>2</sub>	
	1302		Methyl geranate	nd	0.06	C <sub>11</sub> H <sub>18</sub> O <sub>2</sub>	
	1323		γ-Nonalactone	nd	0.03	C <sub>9</sub> H <sub>16</sub> O <sub>2</sub>	
	1348		Silphin-1-ene	nd	0.46	C <sub>15</sub> H <sub>24</sub>	
	1353		α-Cubebene	0.03	nd	C <sub>15</sub> H <sub>24</sub>	
	1363		(E)-β-Damascenone	nd	0.02	C <sub>13</sub> H <sub>18</sub> O	
	1368		cyclosativene	nd	0.05	C <sub>15</sub> H <sub>24</sub>	
	1375		α-copaene	nd	0.09	C <sub>15</sub> H <sub>24</sub>	
_	1380		Modhephene	0.03	0.74	C <sub>15</sub> H <sub>24</sub>	
	1383		β-cubebene	nd	0.02	C <sub>15</sub> H <sub>24</sub>	
	1388		β-bourbonene	0.13	3.28	C <sub>15</sub> H <sub>24</sub>	
	1393		β-Elemene	nd	0.02	C <sub>15</sub> H <sub>24</sub>	
	1399		Cyperene	nd	0.40	C <sub>15</sub> H <sub>24</sub>	
_	1405		α-Barbatene	0.06	1.89	C <sub>15</sub> H <sub>24</sub>	
	1419		(E)-β-caryophyllene	0.04	0.94	C <sub>15</sub> H <sub>24</sub>	
_	1430		β-copaene	nd	0.11	C15H24	
	1445		Sesquisabinene	nd	0.03	C <sub>15</sub> H <sub>24</sub>	
	1449		α-humulene	0.02	0.43	C <sub>15</sub> H <sub>24</sub>	
	1474		trans-cadina-1(6),4-diene	0.09	0.02	C <sub>15</sub> H <sub>24</sub>	
	1480		germacrene-D	0.02	0.24	C <sub>15</sub> H <sub>24</sub>	
	1491		(Z)-\alpha-Bisabolene	0.10	0.91	C <sub>15</sub> H <sub>24</sub>	
	1494		(E),(E)-α-Farnesene	nd	0.32	C <sub>15</sub> H <sub>24</sub>	
	1506		trans-Calamenene	0.01	1.33	C <sub>15</sub> H <sub>22</sub>	
	1512		7-epi-α-Selinene	nd	0.11	C <sub>15</sub> H <sub>24</sub>	
	1514		δ-Cadinene	0.01	0.16	C <sub>15</sub> H <sub>24</sub>	
	1535		trans-cadina-1,4-diene	nd	0.16	C <sub>15</sub> H <sub>24</sub>	
	1542		Selina-3,7(11)-diene	nd	0.09	C <sub>15</sub> H <sub>24</sub>	
	1548		β-Elemol	nd	0.03	C <sub>15</sub> H <sub>26</sub> O	
_	1551		Elemol	nd	0.16	C <sub>15</sub> H <sub>26</sub> O	
	1563		(E)-Nerolidol	nd	0.19	$C_{15}H_{26}O$	
64	1568	1569	Germacrene-D-4-ol	nd	0.32	$C_{15}H_{26}O$	
	1577	1577	caryophyllene oxide	nd	0.03	C <sub>15</sub> H <sub>26</sub> O	

66	1583	1582	Spathulenol	nd	0.06	C <sub>15</sub> H <sub>26</sub> O
67	1594	1593	Viridiflorol	nd	0.10	C <sub>15</sub> H <sub>26</sub> O
68	1601	1601	α-Humulene oxide	nd	0.02	$C_{15}H_{24}O$
69	1610	1607	humulene-1,2-epoxide	nd	0.04	$C_{15}H_{24}O$
70	1615	1614	1,10-di-epi-cubenol	nd	0.12	C <sub>15</sub> H <sub>26</sub> O
71	1625	1629	10- <i>epi</i> -γ-Eudesmol	nd	0.08	C <sub>15</sub> H <sub>26</sub> O
72	1630	1631	$\beta$ -Eudesmol	0.02	0.15	$C_{15}H_{26}O$
73	1635	1635	$\alpha$ -Cadinol	nd	0.14	$C_{15}H_{26}O$
74	1636	1637	$\alpha$ -Eudesmol	nd	0.08	$C_{15}H_{26}O$
75	1651	1653	$\beta$ -Bisabolol	nd	0.04	$C_{15}H_{26}O$
76	1676	1677	andro encecalinol	nd	0.07	$C_{12}H_{12}O_2$
77	1698	1695	β-Sinensal	nd	0.12	C15H22O
78	1715	1715	Mayurone	nd	0.02	$C_{14}H_{22}O$
79	1783	1786	Cis-Cadina-1(2),4-diene	0.05	0.03	$C_{15}H_{24}$
80	1834	1830	6,10,14-Trimethyl pentandecan-2-one	nd	0.06	$C_{18}H_{36}O$
81	1862	1866	1-Hexadecanol	nd	0.08	C <sub>16</sub> H <sub>34</sub> O
82	1899	1884	nonadecane	nd	0.07	C 19H 40
83	1955	1942	isophytol	0.14	0.38	$C_{20}H_{40}O$
84	1978	1976	Ethyl hexadecanoate	0.05	0.02	C18H36O2
85	2107	2106	(E)-phytol	0.04	0.84	C <sub>20</sub> H <sub>40</sub> O
			Total	99.72	99.19	

nd: no detected

#### Conclusion

This research work contributes to the promotion of tropical plant species and little-known plants of the genus Crassocephalum. This study presents for the first time phytochemical results ondf essential oils extracted from the flowers and stems of a species of the genus Crassocephalum harvested in Côte d'Ivoire. Our work shows a myrcene, limonene and  $\beta$ -pinene monoterpene chemotype for the oils extracted from the flowers and a myrcene, terpinolene and (E)-dec-2-enal chemotype for the oils extracted from the stems. Myrcene, which represents almost all of the volatile matter in the oils of the flowers of Crassocephalum sp, is also the main compound of the species Crassocephalum crepidioides with a lower percentage. The high amount of myrcene, an antibacterial, in the oils extracted from the flowers of Crassocephalum sp could indicate a strong antibacterial activity of the flower oil.

#### References

- Adams RP. Identification of Essential oil Components by Gas Chromatography/Mass Spectroscopy. Illinois, USA: Allured Publishing Co; c1995.
- Arbonnier M. Arbres, arbustes et lianes des zones sèches d'Afrique de l'Ouest, Versailles: Ed. Quae; c2009. p. 574.
- 3. Hung N, Satyal P, DN D, Tai T, Huong L, Chuong N, et al. Chemical Compositions of Crassocephalum crepidioides Essential Oils and Larvicidal Activities Against Aedes aegypti, Aedes albopictus, and Culex quinquefasciatus. Natural Product Communications. 2019;14(6):1-5.
- 4. Inoue Y, Shiraishi A, Hada T, Hamashima H, Shimada J. The Antibacterial Effects of Myrcene on Staphylococcus aureus and Its Role in the Essential Oil of the Tea Tree (*Melaleuca alternifolia*). Natural Medicines. 2004;58(10-14).
- 5. Joshi R. Terpene composition of *Crassocephalum crepidioides* from Western Ghats region of India. International Journal of Natural products Research. 2011;1(2):19-22.
- 6. Mc Lafferty FW, Stauffer DB. The Wiley NBS registry of mass spectral data. New York: 2ème Edition John Wiley and Sons; c1989.

- 7. Owokotomo IA, Ekundayo O, Oladosu IA, Aboaba SA. Analysis of the Essential Oils of Leaves and Stems of *Crassocephalum crepidioides* Growing in South Western Nigeria. International Journal of Chemistry. 2012;4(2):34-37.
- 8. Rasoul M, Marei GIK, Abdelgaleil S. Evaluation of antibacterial properties and biochemical effects of monoterpenes on plant pathogenic bacteria. African Journal of Microbiology Research. 2012;6(15):3667-3672.
- 9. Taïwe G, Bum E, Talla E, Dimo T, Sidiki N, Dawe A, et al. Evaluation of antinociceptive effects of Crassocephalum bauchiense Hutch (Asteraceae) leaf extract in rodents. Journal of Ethnopharmacology. 2012;141(1):234-241.
- 10. Wagner W, Herbs D, Sohmer S. Manual of the Flowering Plants of Hawaii, Revised Edition, Bernice P. Bishop Museum Special Publication. University of Hawaii Bishop. Museum Press, Honolulu. 1999;2:1919.
- Zollo PHA, Kuiate JR, Menut C, Bessiere JM. Aromatic Plants of Tropical Central Africa. XXXVI. Chemical Composition of Essential Oils from Seven Cameroonian Crassocephalum Species. Journal of Essential Oil Research. 2000;12(5):533-536.