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Isolation, structural characterization and evaluation of the *in vitro* antioxidant potential of four compounds isolated from the selective leaf extracts of *Bauhinia monandra* Kurz (Fabaceae)

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Abstract

Bauhinia monandra belongs to Fabaceae family, is used in traditional Ivorian medicine to treat diabetes and male infertility. A preliminary study conducted on the leaves, stems and roots showed fluctuating contents of total phytochemicals and flavonoids in the ethyl acetate extract of the leaves. This study made it possible to isolate and elucidate the molecular structures of four phytoconstituents from selective leaf extracts: Vomifoliol (1), guibourtinidol (2), epiafzelechin (3) and D-pinitol (4). The molecular structures of the phytoconstituents were elucidated by NMR ¹H, ¹³C, COSY, HMBC et HMQC. Also, There *in vitro* antioxidant activity was measured using the DPPH and FRAP spectrophotometric methods.

Keywords: *Bauhinia monandra*, structure moléculaire, activité antioxydante, RMN

1. Introduction

Orchids of the genus *Bauhinia*, called Judas tree, are dicotyledonous plants native to Madagascar and widespread in the tropics. The genus includes about 300 species growing in tropical regions around the world [1-2]. They are small trees or shrubs with showy flowers grown as avenue trees. *Bauhinia* species are often used in traditional medicine for the treatment of diabetes [2]. Ethnobotanical information obtained from local traditional herbalists indicates that *Bauhinia monandra* is used to treat diarrhoea, dysentery, fevers, leprosy, smallpox (pox), diabetes, eye diseases. Previous work on *Bauhinia* species has made it possible to isolate different classes of metabolites such as phenolic acids, esters, terpenoids, alkaloids, steroids, tannins, quinines and more commonly flavonoids [2-7]. A recent study carried out on the organs of the Ivorian species made it possible to determine the contents of total phenols and flavonoids, as well as the antioxidant potential by trapping of DPPH and by reduction of Fe³⁺ ions from organic extracts [8]. This work aims to explore the antioxidant potential of four compounds isolated from selective extracts of *Bauhinia monandra* leaves, in relation to their determined molecular structures.

2. Material and Methods**2.1. Plant material**

The plant material used in this study consists of the leaves of *B. monandra* (BMf), harvested in Agboville (city in the south-east of Côte d'Ivoire, capital of the department of Agboville, Agnéby-Tiassa region, 5°55'41" north and 4°13'01" west). After identification by Prof. MALAN Djah François (systematic botanist in NANGUI ABROGOUA University), the organs were cleaned under running water, dried under constant air conditioning (20°C, 20 days), then reduced to powders, then preserved in hermetically sealed glass jars.

2.2. Methods**2.1. Preparation of the crude hydrométhanol extract**

352 g of leaf powder are macerated during 48 hours in 1L of methanol (MeOH, 80%) at room temperature with constant stirring. The operation is repeated three times. After vacuum filtration, the collected filtrates are concentrated using a rotary evaporator (BÜCHI) (40°C, 335 mbar) and drained under liquid nitrogen during 6 h to give 62 g of total hydromethanolic extract (BMF).

2.2. Preparation of selected extracts

To obtain selective extracts, 56 g of BMf are dissolved in 500 mL of distilled water and then successively exhausted with hexane, dichloromethane (DCM), ethyl acetate (AcOEt) and n-butanol (n-BuOH). The organic fractions obtained were dried over anhydrous magnesium sulphate (Na₂SO₄). After filtration on filter paper, hexane (BMf1), DCM (BMf2), AcOEt (BMf3) and n-BuOH (BMf4) fractions are reduced on a rotary evaporator, and dried with a pump under a liquid nitrogen atmosphere for 4 h to provide selective extracts. DCM and AcOEt extracts were used for compound isolation and purification.

2.3. Chromatographic separation of BMf2 and BMf3 fractions

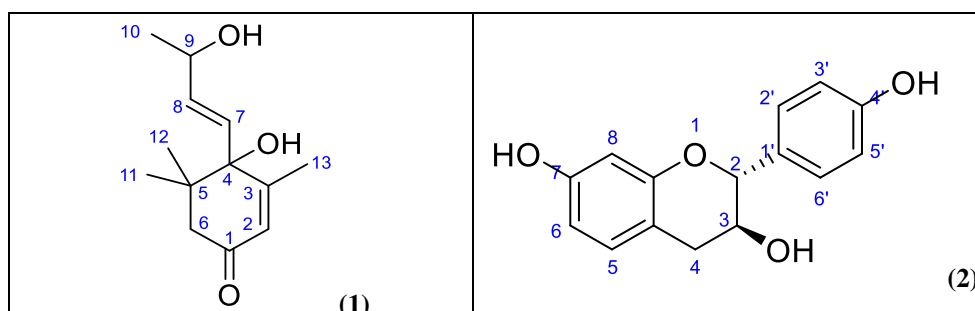
2.3.1. Isolation of compound (1) from the BMf2

245 mg of the DCM fraction are introduced into an open column containing 50 g of silica gel 60 GF254 (Merck) (length 45 cm, diameter 4 cm, height of the silica 17 cm), eluted with a gradient of solvents (DCM/ AcOEt) increasing the polarity to 100% AcOEt (90/10-70/30-50/50-40/60-20/80 - 00/100). The elution of the compounds was controlled by TLC (60 F254, aluminum support, Merck). Subsequently, 170 fractions were collected and grouped according to their similarity into 11 sub-fractions (BMf2-1 to BMf2-11). The BMf 2-9 subfraction (23 mg) showed a single molecular spot by TLC visible under UV/256 nm.

2.3.2 Isolation of compounds (1-4) from BMf3

3 g of AcOEt extract dissolved in MeOH are introduced into an open column. The elution was carried out with two solvent gradients: the first elution was carried out with a DCM/AcOEt mixture in the proportions of 20/80 (v/v) with an increasing increase in polarity up to 100% AcOEt. The second elution was performed with AcOEt/MeOH (9/1) up to 50% MeOH. Two elutions gave 81 fractions grouped into 9 sub-fractions, the purification of which made it possible to collect 6 other sub-fractions (BMf3-₁ to BMf3-₆). The BMf3-₁ sub-fraction (53 mg) was subjected to preparative chromatography, eluted with a solvent mixture DCM/AcOEt, 60/40 (v/v) to give 3 bars (BMf3-₁A fractions (12 mg), BMf3-₁B (10 mg), BMf3-₁C (14 mg) of compounds, after migration, observation under UV light 254-366 nm and visualization with phosphomolybdic acid. The BMf3-₁A fraction (12 mg) gave the sole compound (1) (12 mg). Compound (2) (14 mg) is isolated from BMf3-₁C. The BMf3-₂ subfraction (86 mg), purified and analyzed by preparative plate chromatography, provided compound (3) (41 mg) from BMf3-₂D. Purification of the BMf3-₅ subfraction (131 mg) made it possible to isolate compound (4) (45 mg).

2.4. Structural characterization of isolated compounds



The characterization of the isolated compounds was done by nuclear magnetic resonance techniques (NMR ¹H, ¹³C, COSY, HMBC, HMQC) on a spectrometer (Shimadzu, 400 MHz).

2.5. Determination of antioxidant potential (PO) of isolated compounds

DPPH Test

To 1 mL of the ethanolic extract (obtained from each isolated compound) are added 2.5 mL of an ethanolic solution (0.03 mg/mL) of the 2, 2-diphenyl-1-picrylhydrazyl radical (DPPH). The stirred mixture is incubated for 30 min in the dark. The decrease in the absorbance of the violet color of the DPPH solution is measured at 517 nm on a UV-visible spectrophotometer (Spectro AL 800) against a white ^[9-10]. Plant extracts and quercetin are prepared in the concentration range from 1500 to 2.24 µg/mL. The reduction percentage (PR) is determined by the following formula:

$$PR (\%) = 1 - \frac{\text{extract absorbance}}{\text{DPPH absorbance}} \times 100$$

The median DPPH reduction concentration (CR₅₀) reflecting the antioxidant efficacy parameter of the extract, is determined graphically from the PR ^[11-14]. All tests are performed in triplicate.

FRAP Test

The reagent used is a mixture of sodium acetate buffer (0,3 M; pH= 3,6) and 2, 4, 6-Tri (2-pyridyl)-s-triazine (TPTZ) (10 mM) prepared in hydrochloric acid solution (HCl, 40 mM) and iron trichloride (FeCl₃, 20 mM) in a volume ratio 10: 1: 1. 3 mL of FRAP reagent freshly prepared and preheated to 37 °C in a water bath are added to 100 µL of plant extract (0, 25 mg/mL). The absorbance of the intense blue color induced by the reduction of the colorless ferric complex (TPTZ- Fe³⁺) into ferrous cations (Fe²⁺) is read at 593 nm on a UV-visible spectrophotometer (Spectro AL 800) after 4 min of incubation. Linear calibrations are carried out with ferrous sulphate (FeSO₄.H₂O), 80% pure (standard), at concentrations (0,78125; 1,5625; 3,125; 6,25; 12,5 et 25 µg/mL). Ascorbic acid (vitamin C) was the control antioxidant. The results are expressed in micrograms FeSO₄ equivalent per 100 g of extract (µg ESF/100 gE) ^[15-16]. All tests are performed in triplicate.

3. Results and discussion

3.1. Molecular structures of isolated compounds

Figure 1 presents the molecular structures of the compounds isolated from the BMf2 and BMf3 fractions of the leaves of *B. monandra*.

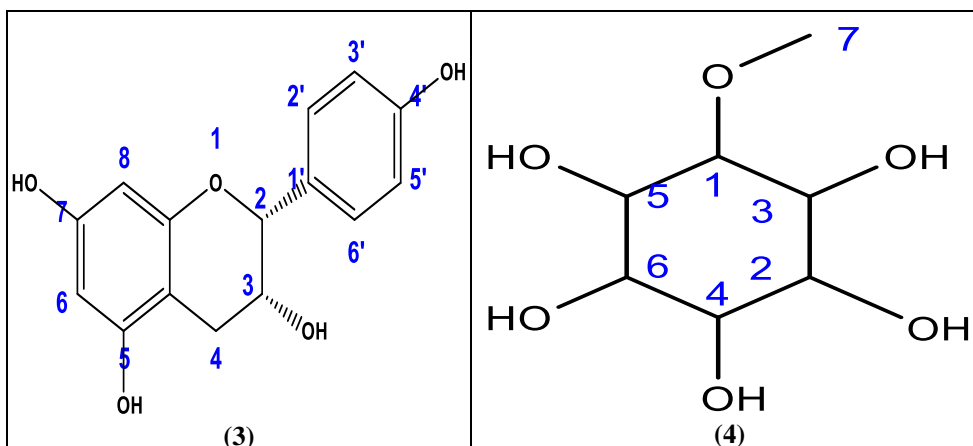


Fig 1: Molecular structures of isolated compounds

Compound (1)

Based on spectral data NMR (^1H , ^{13}C (APT)) (400 MHz, CDCl_3) (Table 1) and COSY, HMBC (Figure 2), and in

comparison, to reported spectral data [17-18] the compound (1) is Vomifoliol.

Table 1: NMR ^1H and ^{13}C spectral data

Position	Compound (1)	
	δ_{C} (ppm)	δ_{H} , mult. (J in Hz)
1	198.3	
2	127.0	5.89 : p; (1.3); 1H
3	163.2	
4	79.2	
5	41.3	
6	49.8	2.22: dd;(17.1; 1.2); 1H _a
		2.43: dd;(17.1; 0.8); 1H _b
7	129.2	5.78: dd;(15.6; 0.2); 1H
8	135.8	5.83: dd; (15.6; 5.5); 1H
9	68.2	4.39: qdd; (6.4; 5.5; 0.2); 1H
10	23.8	1.28 : d; (6.4); 3H
11	24.2	1.00 : s; 3H
12	23.0	1.06 : bs; 3H
13	19.1	1.88 : d; (1.4); 3H

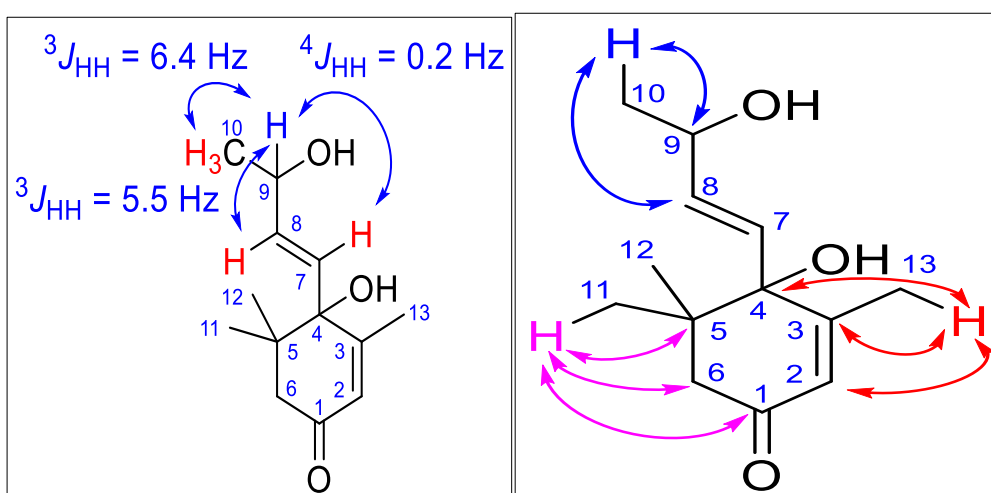


Fig 2: Main COSY and HMBC correlations of Vomifoliol

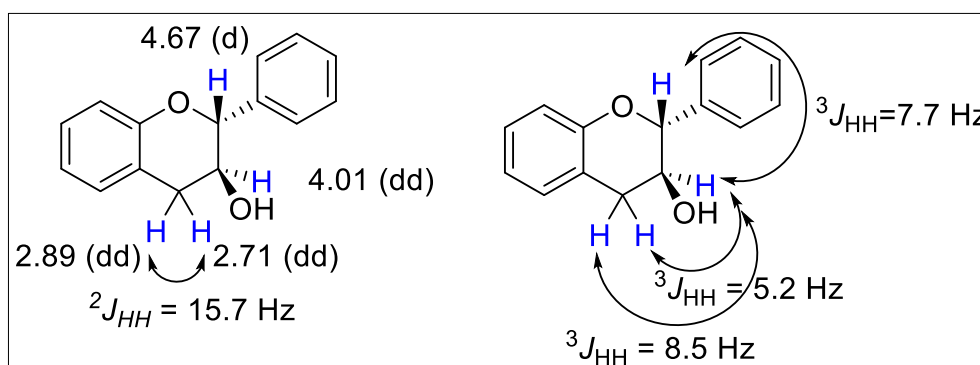
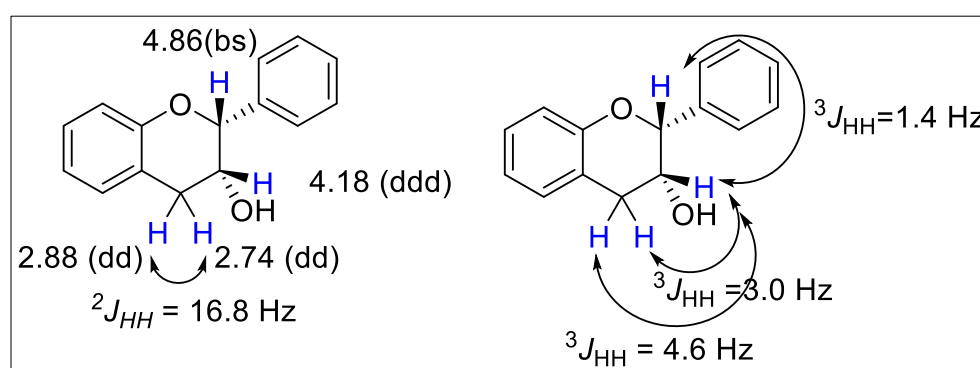
Vomifoliol is a derivative of megastigma [19-20]. It exhibits moderate anticholinesterase and antileishmanial activity [21]. Natural derivatives of megastigma also exhibit antiproliferative effects [22], anticancer and cytotoxic [23].

Compounds (2) and (3): The compounds (2) and (3) are yellow-orange amorphous solids whose spectral data NMR

(^1H , ^{13}C (APT)) (400 MHz, CD_3OD) (Table 2) are consistent with data reported in the literature. The compound (2) (Figure 3) is guibourtinidol ((2R,3S)-4',7-dihydroxyflavan-3-ol) [24-25]. The compound (3) is epiafzelechin (2R,3R)-4',5,7-trihydroxyflavan-3-ol) [24, 26-28].

Table 2: NMR ^1H and ^{13}C spectral data of compounds (2) and (3)

Position	Compound 2		Compound 3	
	δ_{C} (ppm)	δ_{H} , mult. (J in Hz)	δ_{C} (ppm)	δ_{H} , mult. (J in Hz)
2	83.1	4.67 : d; (7.6); 1H	79.9	4.86 : bs; 1H
3	68.8	4.01 : ddd; (8.5; 7.7; 5.2); 1H	67.4	4.18 : ddd; (4.6; 3.0; 1.4); 1H
4	33.6	2.71 : dd; (15.8; 8.5); 1H	29.3	2.74 : dd; (16.8; 3.0); 1H
		2.89 : dd; (15.7; 5.2); 1H		2.88 : dd; (16.8; 4.6); 1H
5	131.2	6.87 : d; (8.3); 1H	157.3	
6	109.5	6.35 : dd; (8.3; 2.4); 1H	96.5	5.92 : d; (2.3); 1H
7	157.9		157.6	
8	103.6	6.28 : d; (2.4); 1H	95.9	5.96 : d; (2.3); 1H
9	156.2		157.9	
10	112.6		100.1	
1'	131.5		131.6	
2'/6'	129.5	7.21 : m; 2H	129.1	7.41-7.26 : m; 2H
3'/5'	116.1	6.79 : m; 2H	115.8	6.83-6.72 : m; 2H
4'	158.5		158.0	

**Fig 3:** Main COSY correlations of guibourtinidol**Fig 4:** Main COSY correlations of epiafzelechin

Guibourtinidol and epiafzelechin are the flavanols which have been described and isolated, respectively in *Cassia sieberiana* [25, 29], *Cassia sieberiana* [30-31] and *Celastrus orbiculatus* [27]. These compounds are known for their pharmacological interest [32-35].

Table 3: NMR ^1H and ^{13}C spectral data of compound (4)

Position	Compound 4	
	δ_{C} (ppm)	δ_{H} , mult. (J in Hz)
1	84.9	3.26 : t; (9.6; 9.1); 1H
2	72.6	3.76 : dd; (9.7; 2.6); 1H
3	73.7	3.91 : d; (3.4); 1H
4	73.4	3.60 : t; (9.4); 1H
5	74.3	3.90 : s; (3.4); 1H
6	72.0	3.70 : dd; (9.7; 2.6); 1H
O-CH ₃	60.6	3.61 : s; 3H

Compound (4)

The compound (4) is D-pinnitol (yellow amorphous solid). NMR (^1H , ^{13}C (APT)) (400 MHz, CD_3OD) Spectral data in Table 3 are consistent with those reported in the literature [36-37].

3.2. Antioxidant potential of isolated compounds

The antioxidant potential of the compounds was evaluated using the DPPH and FRAP tests.

3.2.1. DPPH antioxidant profile

In general, the isolated compounds reduce DPPH. Figure 5 presents the significant and variable reduction percentages (PR) of the different compounds isolated: 87.332±0.179% for vomifoliol; 89.053±0.182% for epiafzelechin; 66.719±0.175% for guibourtinidol and 51.126±0.061% for D-pinnitol.

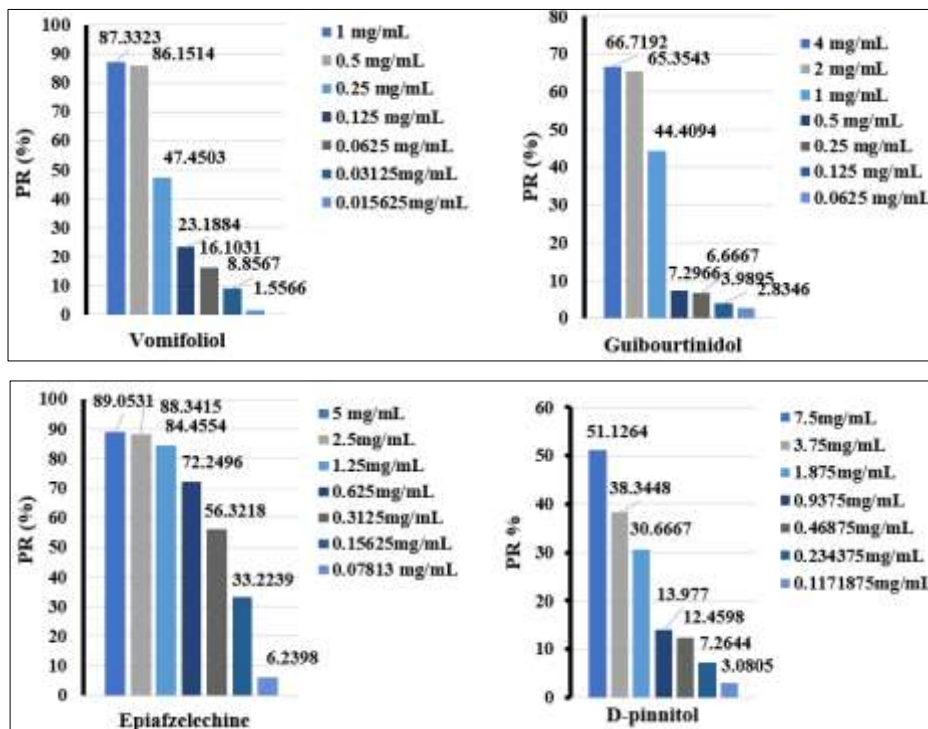


Fig 5: DPPH reduction percentages of molecules isolated from *B. monandra* leaves

Table 4 highlights the antioxidant efficacy of the compounds isolated, which is reflected by the determination of the CR₅₀. This parameter reflects the efficiency of the sample. The lower its value, the more significant the antioxidant activity [38, 14].

Table 4: CR₅₀ of isolated compounds and quercetin

Compounds	CR ₅₀ (µg/mL)
Vomifoliol	278.035
Guibourtinidol	1266.89
Epiafzelechin	269.73
D-pinnitol	7136.44
Quercetin	3.519

According to the analysis of Table 4, epiafzelechin and vomifoliol show the best antioxidant efficacy with regard to DPPH. However, in comparison to quercetin, an antioxidant

potential, epiafzelechin and Vomifoliol exhibit low antioxidant efficacy. One study reported the antioxidant behavior of epiafzelechin extracted from the root bark of *Cassia siberiana* harvested in Togo [30].

3.3.2. FRAP antioxidant profile

The antioxidant potential of the isolated compounds was explored by the reduction of ferric ions (Fe³⁺) of the (Fe³⁺ TPTZ) complex to ferrous ions (Fe²⁺) of the Fe²⁺ TPTZ complex. For this purpose, a linear calibration ($y = 0.0197x - 0.0187$; $R^2 = 0.9991$) was carried out. Figure 6 presents the results obtained.

In light of Figure 6, Vomifoliol (268.42±0.689 µMolEFeSO₄/g ES) and epiafzelechin (702.578±4.196 µMolEFeSO₄/g ES) show weak reducing capacities compared to vitamin C. However, the reducing activity of epiafzelechin is 2.61 times greater than that of vomifoliol.

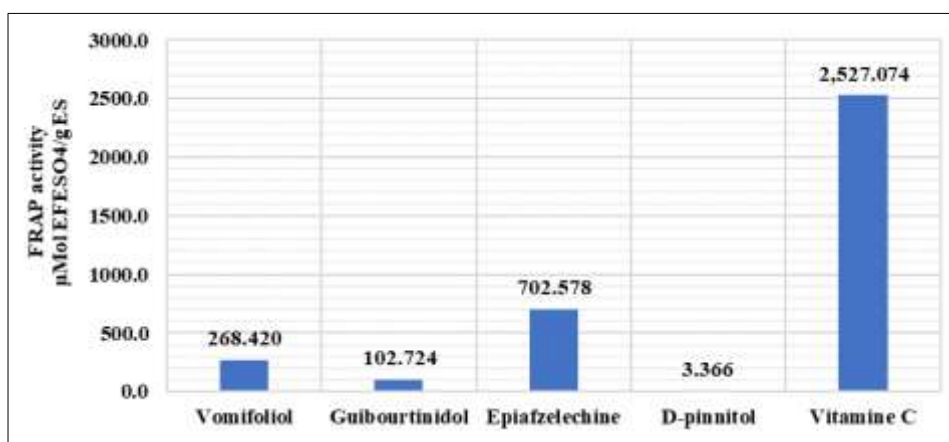


Fig 6: FRAP antioxidant profile of samples and reference

4. Conclusion

Bauhinia monandra is a plant species used in traditional Ivorian medicine in Côte d'Ivoire for the treatment of male infertility and diabetes. This study carried out on the selective

extracts of the leaves made it possible to isolate, purify and elucidate the molecular structures of four compounds. These compounds were isolated from the leaves of the species distributed in Côte d'Ivoire for the first time. Two

complementary spectrophotometric methods for measuring free radical scavenging (DPPH test) and ferric reduction (FRAP test) have made it possible to assess their antioxidant potential. The antioxidant properties clearly expressed through these methods by epiafzelechin and vomifoliol, isolated from the leaves of *Bauhinia monandra* would explain, among other things, the use of the plant and its benefits in traditional medicinal practices in response to pathological conditions.

5. Acknowledgment

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6. Conflict of interest

The authors have not declared any conflict of interests.

7. References

- Cechinel-Filho V. Chemical composition and biological potential of plants from the genus *Bauhinia*. *Phytotherapy Research*. 2009;23(10):1347-1354.
- Silva KL, Cechinel-Filho V. Plantas do gênero *Bauhinia*: Composição Química E Potencial Farmacológico. *Química Nova*. 2002;25(3):449-454.
- Yadav S, Bladoria BK. Two dimeric flavonoids from *Bauhinia purpurea*. *Indian Journal of Chemistry*. 2005;44b:2604-2607.
- Zhao YY, Cui CB, Cai B, Han B, Sun QS. A new phenanthraquinone from the stems of *Bauhinia variagata* L. *Journal of Asian Natural Products Research*. 2005;7(6):835-838.
- Shang XY, Li S, Wang YH, Wang SJ, Yang YC, Shi JG. Chemical constituents of *Bauhinia aurea*. *China Journal of Chinese of Materia Medica*. 2006;31(23):1953-1955.
- Menezes FS, Minto ABM, Ruela HS, Kuster M, Sheridan H, Frankish N. Hypoglycemic activity of two Brazilian *Bauhinia* species: *Bauhinia forficata* L. and *Bauhinia monandra* Kurz. *Revista Brasileira de Farmacognosia*. 2007;17(1):8-13.
- Taweel AMA, Shafae AME, Perveen S, Fawzy GA, Khan SI. Anti-inflammatory and cytotoxic constituents of *Bauhinia retusa*. *International Journal of Pharmacology*. 2015;11(4):372-376.
- Cherif-Soumahoro A, N'gaman-Kouassi KCC, Mamyrbekova-Bekro JA, Pirat JL, Virieux D, Békro YA, et al. Evaluation of the phenolic content and *in vitro* antioxidant potential of *Bauhinia monandra* Kurz (Fabaceae). *Journal of Pharmacognosy and Phytotherapy*. 2022;14(3):37-43.
- Espin JC, Soler-Rivas C, Wichers HJ. Characterization of the Total Free Radical Scavenger Capacity of Vegetable Oils and Oil Fractions Using 2, 2-Diphenyl-1-picrylhydrazyl Radical. *Journal of Agriculture and Food Chemistry*. 2000;48(3):648-656.
- Sladjana MS, Gordana ŠĆ, Jasna MČB, Sonja MD. Comportement cinétique de l'activité de balayage des radicaux DPPH d'extraits de déchets de tomate. *Journal of the Serbian Chemical Society*. 2012;77(10):1381-1389.
- Etepo SD, N'Gaman-Kouassi CKC, Mamyrbekova-Békro JA, Yves-Alain B. Antioxidant profiles of Alcoholic tinctures from *Heterotis rotundifolia* (SM.) Jacq. -Fél (Melastomataceae) by DPPH Radical Trapping. *European Journal of Biomedical and Pharmaceutical Sciences*. 2018;5(10):39-40.
- Tanoh SK, Amani BK, N'gaman-Kouassi CKC, Boa D, Mamyrbekova-Békro JA, Yves-Alain B, et al. DPPH• Scavenging Activity of *Mallotus oppositifolius*: A Kinetic Study. *European Journal of Scientific Research*. 2019a;153(2):196-206.
- Tanoh SK, N'Gaman-Kouassi CKC, Boa D, Mamyrbekova-Békro JA, Yves-Alain B. Activité antioxydante des extraits bruts hydroéthanoliques et hydroacétoniques des organes de quatre plantes de Côte d'Ivoire médicinales. *Revue Nature et Technologie*. 2019b;11(2):28-34.
- Etepo SD, N'Gaman-Kouassi KCC, Yao TR, Mamyrbekova-Bekro JA, Bekro YA. Phytochemical, Antioxidant and pharmacological study of *Heterotis rotundifolia* (Sm.) Jacq.-FeL. (Melastomataceae). *Journal of Pharmaceutical and Biological Sciences*. 2022;10(1):37-43.
- Benzie IFF, Strain JJ. Ferric reducing/antioxidant power assay: Direct measure of total antioxidant activity of biological fluids and modified version for simultaneous measurement of total antioxidant power and ascorbic acid concentration. *Methods in Enzymology*; c1999. p. 15-27.
- Gong J, Huang J, Xiao G, Chen F, Lee B, You Y, et al. Antioxidant Capacities of Fractions of Bamboo Shaving Extract and Their Antioxidant Components. *Molecules*. 2016;21(996):1-14.
- Hammami S, Jannet HB, Bergaoui A, Ciavatta L, Cimino G, Mighri Z, et al. Isolation and Structure Elucidation of a Flavanone, a Flavanone Glycoside and Vomifoliol from *Echinochloa fruticosum* Growing in Tunisia. *Molecules*. 2004. p. 602-608.
- Maria S, Islam F, Qais N, Hasan CM. Isolation of Vomifoliol: A Megastigmane from Leaves of *Antidesma ghaesembilla*, *Asian Journal of Chemistry*. 2013;25(6):3533-3534.
- Stuart KL, Coke LB. The effect of Vomifoliol on stomatal aperture. *Planta*. 1975;122(3):307-310.
- Coke LB, Stuart KL, Whittle YG. Further effects of Vomifoliol on stomatal aperture and on the germination of lettuce and the growth of cucumber seedlings, *Planta*. 1975;127:21-25.
- Mogana R, Adhikari A, Debnath S, Hazra S, Hazra B, Teng-jin K, et al. The anti-acetylcholinesterase and antileishmanial activities of *Canarium patentinervium* Miq. *BioMed Research International*; c2014. p. 1-7.
- Chen WL, Liu QF, Wang J, Zuo JP, Zhu ZX, Zhao WM. New Guaiane, Magastigmane and Eudesmane-type sesquiterpenoids and anti-inflammatory constituents from *Youngia japonica*. *Planta Medica*. 2006;72(2):143-150.
- Ito H, Kobayashi E, Li SH, Hatano T, Sugita D, Kubo N, et al. Antitumor activity of compounds isolated from leaves of *Eriobotrya japonica*. *Journal of Agricultural Food chemistry*. 2002;43(8):2400-2403.
- Rensburg HV, Steynberg PJ, Burger JFW, Heerden PSV, Ferreira D. Circular dichroic properties of flavan-3-ols. *Journal of Chemical Research*; c1999. p. 450-451.
- Reinier JN, Makhosazana M, Johan C, Hendrik VR, Elfranco M, Daneel F, et al. The novel flavan-3-ol, (2R, 3S)-guibourtinidol and its Diastereomers. *Phytochemistry*. 1999;52(6):1153-1158.
- Sethi VK, Taneja SC, Dhar KL, Atal KC. (-)-Epiafzelechin-5-O-β-D-glucoside from *crataeva religiosa*. *Phytochemistry*. 1984;23(10):2402-2403.

27. Min KR, Hwang BY, Lim HS, Kang BS, Oh GJ, Lee J, *et al.* (-)- Epiafzelechin: Cyclooxygenase-1-inhibitor and anti-inflammatory agent from aerial parts of *Celastrus orbiculatus*. *Planta Medica*. 1999;65(5):460-462.
28. Kengo H, Mikiyo W, Shoji Y, Takashi W, Hari PD. Antioxydant phenolic compounds from the rhizomes of *Astilbe rivularis*. *Natural Product Research*. 2018;32(4):453-456.
29. Yang X, He Z, Zheng Y, Wang N, Mulinge M, Schmit JC, *et al.* Chemical Constituents of *Cassia abbreviata* and their Anti-HIV-1 Activity. *Molecules*. 2021;26:2455.
30. Kafui K, Amegnona A, Ana GP, Etchri A, Messanvi G, Gloria P, *et al.* Epiafzelechin from the root bark of *Cassia sieberiana*: Detection by dart mass spectrometry, spectroscopy characterization, and antioxidant properties. *Journal of Natural Products*. 2011;74:455-459.
31. Traore L, Bekro YA, Guiffrey P, Pirat JL, Vireux D, Meudec E, *et al.* Flavanols LC/MS and 1H-RMN from *Ivorian Cassia sieberiana*. *International Journal of Green and Herbal Chemistry*. 2017;6(1):27-34.
32. Youdim KA, Shukitt-Hale B, Joseph JA. Flavonoids and the brain: Interactions at the blood-brain barrier and their physiological effects on the central nervous system, *Free Radical Biology and Medicine*. 2004;37(11):1683-1693.
33. Gossé F, Roussi S, Guyot S, Schoenfelder A, Mann A, Jean-Pierre B, *et al.* Potentiation of apple procyanidin-triggered apoptosis by the polyamine oxidase inactivator MDL 72527 in human colon-derived metastatic cells. *International Journal of Oncology*. 2006;29(2):423-428.
34. Ottaviani JJ, Actis-Goretta L, Villordo JJ, Fraga CG. Procyanidin structure defines the extent and specificity of angiotensin I converting enzyme inhibition. *Biochimie*. 2006;88(4):359-365.
35. Aron PM, Kennedy JA. Flavan-3-ols: Nature, occurrence and biological Activity. *Molecular Nutrition and Food Research*. 2008;52:79-104.
36. Sharma N, Verma MK, Gupta DK, Satti NK, Khajuria RK. Isolation and quantification of pinitol in *Argyrolobium roseum* plant, by 1H-NMR. *Journal of Saudi Chemistry Society*. 2016;20(1):81-87.
37. Akram MK, Ezzat AMG, Hazem AK, El-Sayed ME. New phytoconstituents, anti-microbial and cytotoxic activities of *Acacia etbaica* Schweinf. *Natural Product Research*. 2020;35(25):5571-5580.
38. Falleh H. Antioxidant activity and polyphenol content in the different organs of the *Cynara* wild artichoke cardunculus. *Arid Regions Review*. 2006;61:341-344.